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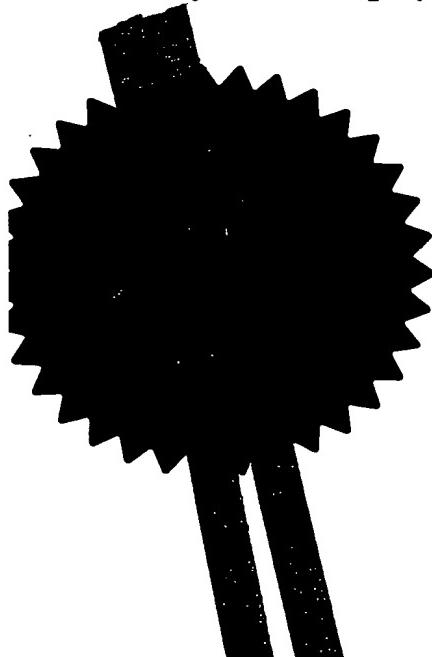
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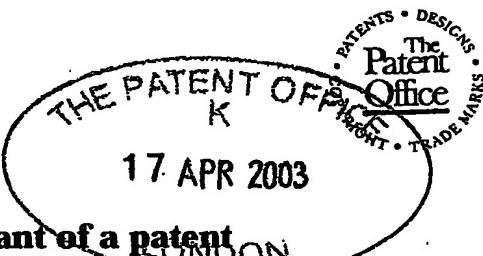
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Molecule

5. Name of your agent (if you have one)

D Young & Co

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DUPLICATE

MOLECULE

FIELD

The present invention relates to the fields of microbiology. It also relates to the fields of medicine, especially therapy and diagnosis.

5 BACKGROUND

Some microorganisms are capable of acting as immunomodulating agents, such as *Mycobacterium smegmatis* used in Freund's complete adjuvant and OK432 from *Streptococcus pygens* as the anti-tumor potentiator. Many polysaccharide immunomodulating agents have also been detected and isolated from *Basidiomycetes* class 10 of fungi, such as lentinan, schizophyllan, TML and SF AI. A novel family of fungal immunomodulatory proteins has been isolated from the edible mushrooms, such as Vvo from *Volvariella volvacea* (grass mushroom), LZ-S from *Ganoderma lucidum* (Ling-Zhi), Gts from *Ganoderma tsugae* (songshan lingzhi), and Fve from *Flammulina velutipes* (golden needle mushroom).

15 Although the therapeutic value of a number of mushrooms has been documented, the active components that confer such therapeutic effects are not well understood.

Ko et al (Eur. J. Biochem., 228, 244-2419) describes the isolation and purification of a protein known as FIP-fve from Golden Needle Mushroom extracts. The authors describe a method of extracting this protein, as well as some biochemical properties of 20 FIP-fve. The amino acid sequence of FIP-fve is presented. FIP-fve is shown to cause proliferation of human peripheral blood lymphocytes, and mice sensitised to BSA are protected against anaphylactic shock by periodic injections of FIP-fve. A hind-paw edema test shows that FIP-fve inhibits antibody production against antigen 48/80. Finally, the authors show that FIP-fve induces expression of IL-2 and IFN- γ in spleen cells from 25 mouse.

An amino acid sequence of FIP-fve is found as GenBank accession numbers:S69147 immunomodulatory protein FIP-fve - golden needle mushroom gi|7438667|pir||S69147[7438667] and P80412 IMMUNOMODULATORY PROTEIN FIP-FVE gi|729544|sp|P80412|FVE_FLAVE[729544].

5 **SUMMARY**

According to a first aspect of the present invention, we provide an Fve polypeptide comprising at least one biological activity of native Fve protein, and being a fragment, homologue, variant or derivative thereof.

Preferably, the Fve polypeptide comprises an immunomodulatory activity.

- 10 Preferably, the biological activity is selected from the group consisting of: up-regulation of expression of Th1/Tc1 cytokines, preferably IFN- γ and TNF- α , down-regulation of expression of Th2/Tc2 cytokines, preferably IL-4 and IL-13, up-regulation of expression of T regulatory (Tr) cytokines IL-10 and TGF- β , hemagglutination activity, cell aggregation activity, lymphocyte aggregation activity, lymphoproliferation activity, up-regulation of expression of IL-2, IFN- γ , TNF- α , but not IL-4 in CD3 $^+$ T cells, interaction with T and NK cells, adjuvant activity, stimulation of CD3 $^+$ CD16 $^+$ CD56 $^+$ natural killer (NK) T cells and CD3 $^+$ CD8 $^+$ CD18 $^{+bright}$ T cells, and up-regulation of allergen specific Th1 immune responses.
- 15

- 20 Preferably, the polypeptide comprises between 2 to 20 residues of amino acid sequence flanking the glycine residue corresponding to position 28 of Fve.

Preferably, the polypeptide comprises the sequence RGT or the sequence RGD.

Preferably, the polypeptide has a sequence as set out in Appendix A or Appendix B.

There is provided, according to a second aspect of the present invention, a Fve polypeptide comprising an sequence selected from the group consisting of: Fve R27A, Fve T29A, GST-Fve (wild type), GST-Fve R27A, and GST-Fve T29A, and fragments, homologues, variants and derivatives thereof.

5 We provide, according to a third aspect of the present invention, a polypeptide comprising a first portion comprising at least a portion of Fve and a second portion comprising at least a portion of an allergen.

10 Preferably, the allergen comprises an allergen from a mite, preferably from Family *Glycyphagidae* or Family *Pyroglyphidae*, preferably a group 1 allergen (Der p 1, Der f 1, Blo t 1, Eur m1, Lep d 1), a group 2 allergen (Der p 2, Der f 2, Blo t 2, Eur m 2, Lep d 2), a group 5 allergen (Blo t 5, Der p 5, Der f 5, Eur m 5, Lep d 5) a group 15 allergen (Der p 15, Der f 15, Blot 15, Eur m 15, Lep d 15).

15 Preferably, the Fve polypeptide or a polypeptide is selected from the group consisting of: Blo t 5-Fve, Blo t 5-FveR27A, Blo t 5-FveT29A, GST-Der p 2-FveR27A, GST-Der p 2-FveT29A, Blo t 5-Der p 2-FveR27A, and Blo t 5-Der p 2-FveT29A. More preferably, it comprises Blo t 5-FveT29A, Der p 2-FveT29A, or Blo t 5-Der p 2-FveT29A.

20 Preferably, the allergen is selected from the group consisting of: tree pollen allergen, Bet v 1 and Bet v 2 from birch tree; grass pollen allergen, Phl p 1 and Phl p 2 from timothy grass; weed pollen allergen, antigen E from ragweed; major feline antigen, Fel'd; major fungal allergen, Asp f1, Asp f2, and Asp f3 from *Aspergillus fumigatus*.

25 As a fourth aspect of the present invention, there is provided a polypeptide comprising a first portion comprising at least a portion of Fve and a second portion comprising at least a portion of a viral antigen selected from the group consisting of: E6 and E7 from HPV; core Ag and E2 from HCV; core and surface antigens from HBV; LMP-1, LMP-2, EBNA-2, EBNA-3 from EBV; and Tax from HTLV-1.

Preferably, it comprises HCV Core23-FveT29A, or HPV E7-FveT29A.

We also provide a polypeptide comprising a first portion comprising at least a portion of Fve and a second portion comprising at least a portion of a viral antigen selected from the group consisting of antigens from Adenovirus, Parainfluenza 3 virus, Human 5 Immunodeficiency Virus (HIV), Herpes simplex virus, HSV, Respiratory syncytial virus, RSV, and Influenza A, Flu A.

We provide, according to a fifth aspect of the present invention, a polypeptide comprising a first portion comprising at least a portion of Fve and a second portion comprising at least a portion of a tumour-associated antigen selected from the group 10 consisting of: MAGE-1, MAGE-2, MAGE-3, BAGE, GAGE, PRAME, SSX-2, Tyrosinase, MART-1, NY-ESO-1, gp100, TRP-1, TRP-2, A2 melanotope, BCR/ABL, Proteinase-3/Myeloblastin, HER2/neu, CEA, P1A, HK2, PAPA, PSA, PSCA, PSMA, pg75, MUM-1, MUC-1, BTA, GnT-V, β -catenin, CDK4, and P15.

Preferably, it comprises MAGE3-FveT29A, MART1-FveT29A or CEA-FveT29A.

15 The present invention, in a sixth aspect, provides a nucleic acid encoding a Fve polypeptide or a polypeptide according to any preceding statement of invention.

Preferably, the nucleic acid comprises CGT GGT ACC, or a sequence which differs from the above by virtue of the degeneracy of the genetic code and which encodes a sequence RGT.

20 In a seventh aspect of the present invention, there is provided a nucleic acid comprising a sequence encoding at least a portion of Fve and a sequence encoding at least a portion of an allergen.

Preferably, it comprises Blo t 5-FveT29A, Der p 2-FveT29A, or Blo t 5-Der p 2-FveT29A.

According to an eighth aspect of the present invention, we provide a nucleic acid comprising a sequence encoding at least a portion of Fve and a sequence encoding at least a portion of a viral antigen selected from the group consisting of: E6 and E7 from HPV; core Ag and E2 from HCV; core and surface antigens from HBV; LMP-1, LMP-2, EBNA-2, EBNA-3 from EBV; and Tax from HTLV-1.

5

Preferably, it comprises HCV Core23-FveT29A, or HPV E7-FveT29A.

We also provide a nucleic acid comprising a sequence encoding at least a portion of Fve and a sequence encoding at least a portion of a viral antigen selected from the group consisting of antigens from Adenovirus, Parainfluenza 3 virus, Human Immunodeficiency Virus (HIV), Herpes simplex virus, HSV, Respiratory syncytial virus, RSV, and Influenza A, Flu A.

10 We provide, according to a ninth aspect of the invention, a nucleic acid comprising a sequence encoding at least a portion of Fve and a sequence encoding at least a portion of a tumour associated antigen selected from the group consisting of: MAGE-1, MAGE-2, MAGE-3, BAGE, GAGE, PRAME, SSX-2, Tyrosinase, MART-1, NY-ESO-1, gp100, TRP-1, TRP-2, A2 melanotope, BCR/ABL, Proteinase-3/Myeloblastin, HER2/neu, CEA, P1A, HK2, PAPA, PSA, PSCA, PSMA, pg75, MUM-1, MUC-1, BTA, GnT-V, β -catenin, CDK4, and P15.

15

Preferably, it comprises MAGE3-FveT29A, MART1-FveT29A or CEA-FveT29A.

20

There is provided, in accordance with a tenth aspect of the present invention, a nucleic acid selected from the group consisting of: Fve R27A, Fve T29A, GST-Fve (wild type), GST-Fve R27A, GST-Fve T29A, Blo t 5-Fve, Blo t 5-FveR27A, Blo t 5-FveT29A, GST-Der p 2-FveR27A, GST-Der p 2-FveT29A, Blo t 5-Der p 2-FveR27A, Blo t 5-Der p 2-FveT29A, and fragments, homologues, variants and derivatives thereof.

As an eleventh aspect of the invention, we provide a vector, preferably an expression vector, comprising a nucleic acid sequence as set out above.

We provide, according to a twelfth aspect of the invention, there is provided DNA vaccine comprising a nucleic acid encoding Fve, a nucleic acid, or a vector as set out
5 above.

According to a thirteenth aspect of the present invention, we provide host cell comprising a nucleic acid encoding Fve, a nucleic acid, or a vector as set out above.

There is provided, according to a fourteenth aspect of the present invention, transgenic non-human organism comprising a nucleic acid encoding Fve, a nucleic acid, or
10 a vector as set out above.

Preferably, the transgenic non-human organism is a bacterium, a yeast, a fungus, a plant or an animal, preferably a mouse.

According to a sixteenth aspect of the present invention, we provide a pharmaceutical composition comprising a polypeptide, a nucleic acid, a vector, a DNA
15 vaccine, or a host cell as set out above, together with a pharmaceutically acceptable carrier or diluent.

According to a seventeenth aspect of the present invention, we provide the use of a native Fve polypeptide, or an Fve polypeptide, nucleic acid, vector, DNA vaccine, host
cell, transgenic organism, or a pharmaceutical composition as set out above as an
20 immunomodulator.

According to an eighteenth aspect of the present invention, we provide the use of a native Fve polypeptide, or an Fve polypeptide, nucleic acid, vector, DNA vaccine, host cell, transgenic organism, or a pharmaceutical composition as set out above to enhance an immune response in a mammal.

According to a nineteenth aspect of the present invention, we provide the use of a native Fve polypeptide, or an Fve polypeptide, nucleic acid, vector, DNA vaccine, host cell, transgenic organism, or a pharmaceutical composition as set out above to stimulate proliferation of CD3⁺ CD8⁺ CD18^{+ bright} T cells.

5 According to a twentieth aspect of the present invention, we provide the use of a native Fve polypeptide, or an Fve polypeptide, nucleic acid, vector, DNA vaccine, host cell, transgenic organism, or a pharmaceutical composition as set out above to stimulate proliferation of CD3⁺ CD16⁺ CD56⁺ natural killer (NK) T cells.

10 According to a twenty first aspect of the present invention, we provide the use of a native Fve polypeptide, or an Fve polypeptide, nucleic acid, vector, DNA vaccine, host cell, transgenic organism, or a pharmaceutical composition as set out above to stimulate production of IL-2, IL-10, TGF-β, IFN-γ or TNF-α in CD3⁺ cells.

Preferably, production of IL-4 is not stimulated in the CD3⁺ cells.

15 According to a twenty second aspect of the present invention, we provide the use of a native Fve polypeptide, or an Fve polypeptide, nucleic acid, vector, DNA vaccine, host cell, transgenic organism, or a pharmaceutical composition as set out above as an adjuvant for a vaccine.

20 According to a twenty third aspect of the present invention, we provide the use of a native Fve polypeptide, or an Fve polypeptide, nucleic acid, vector, DNA vaccine, host cell, transgenic organism, or a pharmaceutical composition as set out above in a method of treatment or prophylaxis of a disease.

According to a twenty fourth aspect of the present invention, we provide the use of a native Fve polypeptide, or an Fve polypeptide, nucleic acid, vector or host cell as set out above for the preparation of a pharmaceutical composition for the treatment of a disease.

According to a twenty fifth aspect of the present invention, we provide a method of treating an individual suffering from a disease or preventing the occurrence of a disease in an individual, the method comprising administering to the individual a therapeutically or prophylactically effective amount of a native Fve polypeptide, or an Fve polypeptide, 5 nucleic acid, vector, DNA vaccine, host cell, transgenic organism, or a pharmaceutical composition as set out above.

Preferably, the use or method is such that disease comprises an atopic disease or allergy.

Preferably, the allergy is selected from the group consisting of: allergic asthma, a 10 seasonal respiratory allergy, a perennial respiratory allergy, allergic rhinitis, hayfever, nonallergic rhinitis, vasomotor rhinitis, irritant rhinitis, an allergy against grass pollen, weed pollen, tree pollen or animal danders, an allergy associated with allergic asthma and a food allergy.

Preferably, the allergy is to a house dust mite from Family Glyphagidae, preferably 15 *Blomia tropicalis* or from Family Pyroglyphidae, preferably *Dermatophagoides pteronyssinus* or *Dermatophagoides farinae*, or to fungi or fungal spores, preferably *Aspergillus fumigatus*.

In an alternative embodiment, the disease comprises a cancer.

According to a twenty seventh aspect of the present invention, we provide the use 20 of a DNA vaccine as described, in a method of treatment or prevention of a cancer, or in a method of suppressing tumour progression.

Preferably, the cancer comprises a T cell lymphoma, melanoma, lung cancer, colon cancer, breast cancer or prostate cancer.

According to a twenty eighth aspect of the present invention, we provide a method of identifying a molecule capable of binding to Fve, the method comprising exposing a native Fve polypeptide, or an Fve polypeptide or nucleic acid, vector, host cell or transgenic organism according as set out above to a candidate molecule and detecting whether the candidate molecule binds to the native Fve polypeptide, or an Fve polypeptide or nucleic acid, vector, host cell or transgenic organism.

According to a twenty ninth aspect of the present invention, we provide a method of identifying an agonist or antagonist of an Fve polypeptide, the method comprising: (a) providing a cell or organism; (b) exposing the cell or organism to a native Fve polypeptide, or an Fve polypeptide or nucleic acid, vector, host cell or transgenic organism as set out above; (c) exposing the cell to a candidate molecule; and (d) detecting an Fve mediated effect.

Preferably, the Fve mediated effect is selected from the biological activities set out above.

Preferably, the method further comprises isolating or synthesising a selected or identified molecule.

According to a thirtieth aspect of the present invention, we provide a molecule identified or selected using such a method.

According to a thirty first aspect of the present invention, we provide a native Fve polypeptide, or an Fve polypeptide in crystalline form.

Preferably, the crystal has the structural coordinates shown in **Appendix C**.

According to a thirty second aspect of the present invention, we provide a model for at least part of Fve made using such a crystal.

According to a thirty third aspect of the present invention, we provide a method of screening for a receptor capable of binding to Fve, or designing a ligand capable of modulating the interaction between Fve and an Fve receptor, comprising the use of such a model.

5 According to a thirty fourth aspect of the present invention, we provide a computer readable medium having stored thereon the structure of such a crystal or such a model.

According to a thirty fifth aspect of the present invention, we provide a ligand identified by the method set out above.

10 According to a thirty sixth aspect of the present invention, we provide a use of such a molecule or such a ligand for the treatment or prevention of a disease in an individual.

According to a thirty seventh aspect of the present invention, we provide a pharmaceutical composition comprising such a molecule or such a ligand and optionally a pharmaceutically acceptable carrier, diluent, excipient or adjuvant or any combination thereof.

15 According to a thirty eighth aspect of the present invention, we provide a method of treating and/or preventing a disease comprising administering such a molecule or such a ligand and/or such a pharmaceutical composition to a mammalian patient.

According to a thirty ninth aspect of the present invention, we provide a method of amplifying a sub-population of cells, the method comprising: (a) obtaining a population of 20 cells from an individual; (b) amplifying CD3⁺ CD8⁺ and CD18^{+ bright} T cells by exposing the population of cells to a native Fve polypeptide, or an Fve polypeptide or nucleic acid, vector, host cell or transgenic organism as set out above.

Preferably, the method further comprises the step of: (c) isolating the CD3⁺ CD8⁺ and CD18^{+ bright} T cells.

According to a fortieth aspect of the present invention, we provide a method of treating an individual suffering from a disease or preventing the occurrence of a disease in an individual, the method comprising amplifying a CD3⁺ CD8⁺ and CD18^{+ bright} T cell by such a method, and administering the amplified CD3⁺ CD8⁺ and CD18^{+ bright} T cell to an 5 individual.

According to a forty first aspect of the present invention, we provide a combination comprising a first component comprising an immunomodulator and a second component comprising at least a portion of an allergen, a viral antigen or a tumour associated antigen.

Preferably, the first component is separate from the second component.

10 Alternatively, or in addition, the first component may be associated with the second component. Preferably, the combination comprises a fusion protein.

The first component may comprise a native Fve polypeptide, or a polypeptide as set out above. The second component may comprise an allergen selected from the group consisting of: a mite allergen, an mite allergen from Family *Glycyphagidae* or Family 15 *Pyroglyphidae*, a group 1 allergen (Der p 1, Der f 1, Blo t 1, Eur m1, Lep d 1), a group 2 allergen (Der p 2, Der f 2, Blo t 2, Eur m 2, Lep d 2), a group 5 allergen (Blo t 5, Der p 5, Der f 5, Eur m 5, Lep d 5), a group 15 allergen (Der p 15, Der f 15, Blot 15, Eur m 15, Lep d 15), a tree pollen allergen, Bet v 1 and Bet v 2 from birch tree; grass pollen allergen, Phl p 1 and Phl p 2 from timothy grass; weed pollen allergen, antigen E from ragweed; major 20 feline antigen, Fel d; major fungal allergen, Asp f1, Asp f2, and Asp f3 from *Aspergillus fumigatus*.

In preferred embodiments, the second component comprises a viral antigen selected from the group consisting of: E6 and E7 from HPV; core Ag and E2 from HCV; core and surface antigens from HBV; LMP-1, LMP-2, EBNA-2, EBNA-3 from EBV; and 25 Tax from HTLV-1. Alternatively, or in addition, the second component may comprise a tumour-associated antigen selected from the group consisting of: MAGE-1, MAGE-2, MAGE-3, BAGE, GAGE, PRAME, SSX-2, Tyrosinase, MART-1, NY-ESO-1, gp100,

TRP-1, TRP-2, A2 melanotope, BCR/ABL, Proteinase-3/Myeloblastin, HER2/neu, CEA, P1A, HK2, PAPA, PSA, PSCA, PSMA, pg75, MUM-1, MUC-1, BTA, GnT-V, β -catenin, CDK4, and P15.

- We further disclose an immunomodulator-antigen conjugate, preferably an
5 immunomodulator-allergen conjugate, an immunomodulator-tumour associated antigen conjugate or a immunomodulator-viral antigen conjugate, in which the immunomodulator preferably comprises an Fve polypeptide.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1. Analysis of purified native Fve by SDS-PAGE and gel filtration chromatography. (a). The native Fve protein purified by cation and anion exchange chromatography is analyzed by Tricine SDS-PAGE. Fve protein gave a single band with an apparent molecular mass of 12.7 kDa. Lane M, molecular mass markers; lane 1, purified native Fve protein. (b) Elution profile of calibration proteins by Superdex 75 chromatography. Peaks, 1. bovine serum albumin (67 kDa); 2. ovalbumin (43 kDa); 3. 15 chymotrypsinogen A (25 kDa); 4. ribonuclease A (13.7 kDa). (c) Purified native Fve formed homodimer at 25.5 kDa.

Figure 2 shows a profile of cytokines and iNOS produced by mouse splenocytes upon stimulation with Fve protein. Mouse spleen cells from Balb/cJ mice are stimulated with 20 μ g of Fve. The mRNAs of cytokines are analyzed by RT-PCR after culturing for 48 hours. A: A non-stimulated culture as negative controls, B: A culture stimulated with 20 μ g of Fve.

Figure 3 shows a profile of human cytokines, transcriptional factors, adhesion molecule and anti-apoptotic protein produced by human PBMC upon stimulation with Fve protein. Human PBMC from healthy donor are stimulated with 20 μ g of Fve. The mRNA expression is analyzed by RT-PCR after culturing for 48 hours. A: A non-stimulated culture as negative control, B: A culture stimulated with 20 μ g of Fve.

Figure 4. A schematic representation showing the principle of overlap extension PCR for the generation of single amino acid residue substitution (A) and deletion mutagenesis of DNA (B).

Figure 5. A schematic representation of the strategy used to generate mutants. On 5 the basis of the structures predicted by PHD prediction program, eleven deletion mutants and three point mutants of Fve plasmid DNA are generated by PCR-based mutagenesis.

Figure 6. SDS-PAGE analysis of recombinant Fve mutant proteins.

Figure 7. *In vitro* proliferation assay of mouse splenocytes. Mouse splenocytes from Balb/cJ is stimulated with 2.5 μ g/ml, 5 μ g/ml, 10 μ g/ml, and 20 μ g/ml, respectively, 10 with 13 of Fve mutant proteins for 48 hours. Recombinant GST-Fve is positive control. GST is negative control.

Figure 8. Lymphoproliferation activity of human PBMC at 48 hours. Human PBMC from a healthy donor is stimulated with 2.5 μ g/ml, 5 μ g/ml, 10 μ g /ml, and 20 μ g /ml, respectively, with eleven of Fve mutant proteins for 48 hours. Recombinant GST-Fve and 15 native Fve are positive control. GST and Blo t 5 are negative control.

Figure 9. Recombinant GST-Fve (wild type) and GST-FveT29 mutant protein showed strong lymphoproliferative activity. Human PBMC from healthy donor are cultured with: (A) no antigen, (B) GST, (C) wild type GST-Fve, (D) GST-FveT29, each protein is used at 20 μ g /ml. The percentage of CD3 $^{+}$ T lymphocytes is analyzed at day 5 by 20 using flow cytometry.

Figure 10. Increased production of TNF- α , IFN- γ , IL-2 but not IL-4 in CD3 $^{+}$ T lymphocytes after stimulation with native Fve protein. The production of (A) IL-4; (B) IL-2; (C) IFN- γ and (D) TNF- α by human PBMC after stimulation with 20 μ g /ml of native Fve protein for three days. The data are analyzed by flow cytometry.

Figure 11. Recombinant wild type GST-Fve and mutant GST-FveT29A, but not mutant GST-FveG28A, maintained IFN- γ cytokine production activity. Human PBMC from healthy donor are cultured with 20 μ g of GST (1); GST-Fve (2); GST-FveR27A (3); GST-FveG28A (4); GST-FveT29A (5). IFN- γ cytokine by T cells is detected at day 3 by staining with anti-CD3 PerCP and anti-IFN- γ FITC specific monoclonal antibody. IFN- γ secretion by small granular lymphocytes and large granular lymphocytes are shown in (a) and (b), respectively. The total amount of IFN- γ production by T cells is the sum of (a) and (b).

Figure 12. Recombinant wild type GST-Fve and mutant GST-FveT29A, but not mutant GST-FveG28A, maintained TNF- α production activity. Human PBMC from healthy donor are cultured with 20 μ g of GST (1); GST-Fve (2); GST-FveR27A (3); GST-FveG28A (4); GST-FveT29A (5). IFN- γ cytokine by T cells is detected at day 3 by staining with anti-CD3 PerCP and anti- TNF- α FITC specific monoclonal antibody. TNF- α secretion by small granular lymphocytes and large granular lymphocytes are shown in (a) and (b), respectively. The total amount of TNF- α production by T cells is the sum of (a) and (b).

Figure 13. Schematic representation of the experimental design of the *in vivo* study Balb/cJ mice are immunized with Der p 2 in aluminum hydroxide at day 0 and boosted at day 21 by intraperitoneal injection. Treatment with Der p 2 alone or Der p 2 plus Fve is started at day 28 by given 6 subcuteneous injections over 12 days. Mice are challenged with Der p 2 at day 42.

Figure 14. Enhanced anti-Der p 2 IgG2a by adjuvanticity of Fve protein. IgG2a response in mice that are subcutaneously injected six times with Der p 2 alone (close circle), or Der p 2 plus Fve (close square) twenty-eight days after the initial sensitization with Der p 2 in alum. Mice received third intraperitoneal injection with Der p 2 in alum at day 42. Results are shown as mean titers and error bars indicate the standard deviations from the mean titers.

Figure 15. Fve could reduce wheal and erythematic flare formation on skin prick test-positive human subject. Both the left and right hands of the house dust mite allergen sensitized human subject are challenged with saline, histamine, Der p 2, and mixture of Der p 2 and Fve at the separated sites on hands. The diameter sizes of wheal (A) and 5 erythematic flare (B) are measured after 10 minutes incubation time

Figure 16. A schematic representation of the seven fusion proteins of Bt5-Fve (wild type), Bt5-FveR27A, Bt5-FveT29A, Dp2-FveR27A, Dp2-FveT29A, Bt5-Dp2-FveR27A, and Bt5-Dp2-FveT29A.

Figure 17. Expression and purification of recombinant fusion protein Bt5-Fve, Bt5-10 FveR27A, and GST-Dp2-FveR27A. Lane 1 and 10 are protein marker. Lane 2 to 9 are GST; Blo t 5; Fve; Bt5-Fve; Bt5-FveR27A; Der p 2; Fve; and GST-Bt5, respectively.

Figure 18. Functional characterization of recombinant fusion proteins of Fve and allergen. The morphology of human lymphocytes upon stimulation with three different fusion proteins for three days. All photographs are taken at a magnification of $\times 10$ and $\times 40$ 15 with a confocal microscope. 1(a) Control: Non-stimulated (10×10 magnification); 1(b) Control: Non-stimulated (40×10 magnification); 2(a): $20 \mu\text{g}$ of GST 10×10 ; 2(b): $20 \mu\text{g}$ of GST 40×10 ; 3(a): $20 \mu\text{g}$ of Blo t 5 10×10 ; 3(b): $20 \mu\text{g}$ of Blo t 5 40×10 ; 4(a): $20 \mu\text{g}$ of native Fve 10×10 ; 4(b): $20 \mu\text{g}$ of native Fve 40×10 ; 5(a): $20 \mu\text{g}$ of Bt5-Fve 10×10 ; 5(b): $20 \mu\text{g}$ of Bt5-Fve 40×10 ; 6(a): $40 \mu\text{g}$ of Bt5-Fve 10×10 ; 6(b): $40 \mu\text{g}$ of Bt5-Fve 40×10 ; 7(a) $40 \mu\text{g}$ of Bt5-FveR27A 10×10 ; 7(b): $40 \mu\text{g}$ of Bt5-FveR27A 40×10 ; 8(a): $20 \mu\text{g}$ of Der p 2 10×10 ; 8(b): $20 \mu\text{g}$ of Der p 2 40×10 ; 9(a): $40 \mu\text{g}$ of GST-Dp2-FveR27A 10×10 ; 9(b): $40 \mu\text{g}$ of GST-Dp2-FveR27A 40×10 . Human lymphocytes maintained aggregation ability upon stimulation with Bt5-Fve (5a, 5b, 6a, 6b) and Bt5-FveR27A (7a, 7b) for 3 days. Native Fve (4a, 4b) is a positive control. Non-stimulated cells (1a, 1b), GST (2a, 2b), Blo t 5 (3a, 3b), 25 and Der p 2 (8a, 8b) are negative controls. The aggregation ability of GST-Dp2-FveR27A is not apparent at day 3 (9a, 9b).

Figure 19. Cell number comparison of human PBMC after 7 days cultured with tested antigens. Human PBMC are cultured with different doses of recombinant allergen and Fve fusion proteins. Non-stimulated cells and cells stimulated with either 20 μ g of Blo t 5; 20 μ g of Fve; 20 μ g of Bt5-Fve; 40 μ g of Bt5-Fve; 20 μ g of Bt5-FveR27A; and 40 μ g of Bt5-FveR27A are shown in Figure 19A. Cells stimulated with 20 μ g of Der p 2; 20 μ g of GST-Dp2-FveR27A; and 40 μ g of GST-Dp2-FveR27A are shown in Figure 19B. The cells are collected and counted at day 7.

Figure 20. The lymphoproliferation activity of human lymphocytes upon stimulation with recombinant fusion protein Bt5-Fve for 72 hours. Human PBMC from a healthy donor is co-cultured with 5 μ g /ml, 10 μ g /ml, 20 μ g /ml, and 40 μ g /ml, respectively, with fusion protein Bt5-Fve (BFwt). Recombinant GST and Blo t 5 are used as negative controls. Fve is used a positive control.

Figure 21. Bt5Fve fusion protein maintained CD8 T cells polarization activity. Human PBMC are isolated from healthy donar and stimulated with 20 μ g of GST (b), 20 μ g of Blo t 5 allergen (c), 20 μ g of Fve (d), 20 μ g of Bt5Fve (e), 40 μ g of Bt5Fve (f), 20 μ g of Bt5FveR27 (g), and 40 μ g of Bt5FveR27 (h) for 5 days. Cells without any stimulation served as negative control (a). Cultured cells are stained with CD3-PerCP and CD8-FITC monoclonal antibodies and analyzed with FACSCalibur cytometry.

Figure 22. Fve and allergen-Fve fusion protein are able to induce T helper type 1 and T regulatory immune responses. (A). Fve induced IFN- γ and IL-10 production. Human PBMC from seven individuals are cultured with 20 μ g of Fve. The production of IFN- γ , IL-4 and IL-10 is assayed by ELISA at day 3. (B). Comparable levels of IFN- γ production are induced by Fve and allergen – Fve fusion protein. Human PBMC are stimulated with Fve, Blot5, Blot5-Fve (wild type) and Blot5-FveR27A (mutant), respectively. The production of IL-4 and IFN- γ is detected by ELISA at day 3 and day 7.

Figure 23. Competitive inhibition assay. Varying concentrations of inhibitors are used to inhibit the binding of human IgE to GST-Blot5 bound to the Elisa plate. The

concentration of different inhibitors ranged from 0.01ng to 10000ng/ml. Results are obtained from serum of a representative allergic subject with high IgE reactivity to house dust mite allergens. GST: Glutathione S-transferase. GF: GST-Fve. GFB: GST-Fve-Blot5. GBF: GST-Blot5-Fve. BF: Blot5-Fve. B: Blot 5.

- 5 Figure 24. Human PBMC stimulated with native Fve protein for five days showed a significant increase in CD16⁺ and CD56⁺ cells. The CD3⁺ cells and CD16⁺ + CD56⁺ cells are analyzed by FACScan after staining with anti-CD3 FITC, anti-CD16 PE and anti-CD56 PE conjugated mouse anti-human specific monoclonal antibody. Cells stimulated with (a) no antigen; (b). 5 μ g of Der p 2 house dust mite allergen as negative control; (c).
- 10 10 5 μ g of PHA; (d). 5 μ g of Fve; (e). 25 μ g of Fve.

- Figure 25. Human PBMC stimulated with Fve protein for ten days showed a significant increase in CD8⁺ cells. The proportion of CD4⁺ and CD8⁺ T cells are analyzed by FACScan after staining with anti-CD4 FITC and anti-CD8 PE conjugated mouse anti-human specific monoclonal antibody. Cells are stimulated with (a). no antigen; (b). 5 μ g of Der p 2 house dust mite allergen as negative control; (c). 5 μ g of PHA; (d). 5 μ g of Fve; (e). 25 μ g of Fve.

- 15 Figure 26. Expanded human CD3⁺CD18^{+Bright} T cells subset in human PBMC after stimulation with Fve protein for five days. Human PBMC from healthy donor are cultured alone (a and c) or with 20 μ g of native Fve protein (b and d) for 5 days. Cells are then analyzed by flow cytometry after staining with anti-CD3 PerCP, anti-CD8 PE and anti-CD18 FITC.

- 20 25 Figure 27. Expanded CD3⁺CD8^{+Bright}CD18^{+Bright} T cells in human PBMC after cultured with Fve protein for five days. Human PBMC from healthy donor are cultured alone (a and c) or with 20 μ g of native Fve protein (b and d) for five days. Cells are analyzed by flow cytometry after staining with anti-CD3 PerCP, anti-CD8 PE and anti-CD18 FITC.

Figure 28. Proportion of *in vivo* BrdU incorporated natural killer (NK) cells from spleen of C57BL/6J naïve mice (a), or mouse received three consecutive subcutaneous injections with 10 μ g of Fve (b), 50 μ g of Fve (c), 250 μ g of Fve (d). Splenocytes are stained with anti-Pan NK PE and anti-BrdU FITC monoclonal antibodies and then analyzed with flow cytometry.

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Figure 29. Proportion of *in vivo* BrdU incorporated CD8 $^{+}$ T cells from spleen of C57BL/6J naïve mice (a), or mouse received three consecutive subcutaneous injections with 10 μ g of Fve (b), 50 μ g of Fve (c), 250 μ g of Fve (d). Splenocytes are stained with anti-CD8 PE and anti-BrdU FITC monoclonal antibodies and then analyzed with flow cytometry.

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Figure 30. Proportion of *in vivo* BrdU incorporated CD4 $^{+}$ T cells from spleen of C57BL/6J naïve mice (a), or mouse received three consecutive subcutaneous injections with 10 μ g of Fve (b), 50 μ g of Fve (c), 250 μ g of Fve (d). Splenocytes are stained with anti-CD4 PE and anti-BrdU FITC monoclonal antibodies and then analyzed with flow cytometry.

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Figure 31. Proportion of *in vivo* BrdU incorporated CD19 $^{+}$ B cells from spleen of C57BL/6J naïve mice (a), or mouse received three consecutive subcutaneous injections with 10 μ g of Fve (b), 50 μ g of Fve (c), 250 μ g of Fve (d). Splenocytes are stained with anti-CD19 PE and anti-BrdU FITC monoclonal antibodies and then analyzed with flow cytometry.

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Figure 32. Proportion of *in vivo* BrdU incorporated CD8 $^{+}$ T cells from lymph nodes of C57BL/6J naïve mice (a), or mouse received three consecutive subcutaneous injections with 10 μ g of Fve (b), 50 μ g of Fve (c), 250 μ g of Fve (d). Lymph nodes are stained with anti-CD8 PE and anti-BrdU FITC monoclonal antibodies and then analyzed with flow cytometry.

Figure 33. Proportion of CD4⁺ and CD8⁺ T cell subsets from mouse peripheral blood mononuclear cells of Balb/cJ naïve mouse (a), or mouse received seven consecutive subcutaneous injections with 125µg of Fve. Panels (b), (c), (d) represent results for three respective individual mouse. Mouse peripheral blood mononuclear cells are collected in a tube with anti-coagulant. Cells are stained with anti-CD8 PE and anti-CD4 FITC monoclonal antibodies and then analyzed by flow cytometry.

Figure 34. Schematic representative of two mammalian eukaryotic expression vectors. (A) pCI-neo can constitutively express high level of recombinant protein in mammalian cells (Picture adopted from Promega, USA). (B) pDisplay can display recombinant protein to the surface of mammalian cells (Picture adopted from Invitrogen life technologies, USA).

Figure 35. Growth suppression of EL4 solid tumor. C57BL mice are inoculated with 8×10^6 EL4 cells have reduced tumor growing rate in the group treated with pCIneo-fve plasmid DNA and Fve protein (Square curve). The control group received pCIneo DNA vector alone and 1xPBS (Diamond curve). EL4 tumor formation is observed at day 3. 100µg of pCIneo-fve DNA is intramuscularly injected into the tibialis muscle at days 0 and 7. 20µg of Fve protein is given by subcutaneous injection at days 5, 7, 9, 11, 13, 15, and 18, respectively.

Figure 36. C57BL/6J mice with EL4 solid tumor have extended mean survival time following treatment with the native Fve protein. Eight weeks old female C57BL mice are inoculated with EL4 tumor in the dorsal back. Tumor formation is observed 3 days after inoculation. Red line: 100µg of pCIneo-fve plasmid DNA is intramuscularly injected at the tribilis muscle at days 0 and 7. Mice are received 20µg of native Fve protein treatment by subcutaneous injection surrounding the tumor site at days 5, 7, 9, 11, 13, 15, and 18, respectively. Blue line: Mice received 100µg of pCIneo vector alone and 1xPBS as control group.

Figure 37. C57BL/6J mice with B16-F1 melanoma have extended mean survival time following treatment with native Fve protein. Mice are inoculated with B16-F1 tumor cells in the dorsal back. Tumor formation is observed at day 3. Red line: 200 μ g of pCIneo-fve plasmid DNA is intramuscularly injected at the tribilis muscle at days -30 and day -1. 5 50 μ g of Fve protein is given by subcutaneous injection surrounding the tumor site at days 4, 7, 9, and 12, respectively. Blue line: Mice received 200 μ g of pCIneo vector and 1xPBS as control group.

Figure 38. B16-Fve transfectant has longer survival rate as comparing with B16-vec transfectant. Two groups of C56BL/6J female mice are inoculated either with 5 \times 10⁴ of 10 B16-Fve (Red line) or 5 \times 10⁴ of B16-vec (Blue line) transfectants in the right flank. Transfectant melanoma solid tumor is established at days 5-7. The fatal rates of mice are recorded and presented as survival curve.

Figure 39. Combined DNA vaccination and Fve gene-transduced melanoma cells synergizes the extension of life span in solid tumor-established mice. C57BL/6J mice are 15 separated into three groups and each group consisted of ten mice. Mice are inoculated with 5 \times 10⁴ of B16-F1 tumor transfectants in the dorsal back. Tumor formation is observed at day 5-7. 100 μ g of pCIneo-fve plasmid DNA is intramuscularly injected at the right and left tribilis muscle of C57BL/6J at day -77, day -35 and day -21. Mice are subcutaneously injected with 5 \times 10⁴ of B16-Fve transfectants (Red line) and B16-vec transfectant (Green 20 Line) at day 0, respectively. 100 μ g of pCIneo plasmid DNA is operated as same experimental procedure and mice are subcutaneously injected with 5 \times 10⁴ of B16-vec transfectants as negative control (Blue line).

Figure 40. Strategy of oral primed with Fve protein and intramuscular boosted with 25 plasmid DNA could extend the survival rate of mice with lung metastasis. Two groups of five C57BL/6J mice are given with 10mg/ml of Fve protein in the drinking water at day -35, -28 and -21, and each water providing is maintained consecutively for one week. Mice are intravenously injected with 2 \times 10⁴ of B16-F1 (wild type) melanoma cells at day 0. One week after, mice are intramuscularly injected with 100 μ g of pCIneo-fve plasmid DNA into

the right and left tribilis muscle, respectively. The mixture of 5×10^4 of B16-Fve cells lysate plus 10 μ g of Fve protein (Red line) or 10 μ g of Fve protein alone (Green line) are subcutaneously injected to mice at the following three weeks. Negative control group of mice received same amount of 1xPBS in the drinking water, intravenously injected with 5 2×10^4 of B16-F1 melanoma cells, followed by intramuscularly injected with plasmid DNA vector pCIneo, and finally subcutaneously injected with B16-vec cells lysate plus 1xPBS (Blue line).

Figure 41. Two representative crystals of Fve. Tetragonal crystal is grown in 2% PEG 400, 2.0 M Ammonium Sulfate; 0.1 M Tris-HCl pH 8.5. The crystal dimensions are 10 approximately 1 mm \times 0.9 mm \times 0.5 mm.

Figure 42. 1° oscillation image of Fve crystal. The edge of the image corresponds to a resolution of 1.4 \AA . Image displayed with Mosflm/Scala.

Figure 43, 44A, 44B, 44C, 45A and 45B show structures of Fve.

SEQUENCES

15 Appendix A shows the nucleic acid and/or amino acid sequences of the deletion mutants Fve D6-18, Fve D19-33, Fve D34-46, Fve D47-60, Fve D61-72, Fve D73-84, Fve D85-97, Fve D98-106, Fve D107-115, Fve D61-97, Fve p55-100.

20 Appendix A also shows the nucleic acid and/or amino acid sequences of the substitution mutants Fve R27A, Fve G28A, Fve T29A, as well as the fusion proteins Blo t 5-Fve (two-in-one chimeric wild type), Blo t 5-Fve R27A (two-in-one chimeric mutant), Blo t 5-Fve T29A (two-in-one chimeric mutant), Der p 2-Fve R27A (two-in-one chimeric mutant), Der p 2-Fve T29A(two-in-one chimeric mutant), Blo t 5-Der p 2-Fve R27A(three-in-one chimeric mutant).

Appendix A also shows the nucleic acid and/or amino acid sequences of the Fusion Proteins of Viral Antigen and Fve, HPV E7-FveT29A and HCV Core23-FveT29A, as well as the nucleic acid and/or amino acid sequences of the Fusion Proteins of Tumor-Associated Antigen and Fve, MAGE3-FveT29A, MART1-FveT29A and CEA-FveT29A.

5 Appendix A also shows the sequences of the primers Fd6-18F (36 mer), Fd6-18R (36 mer), Fd19-33F(36 mer), Fd19-33R(36 mer), Fd34-46F(36 mer), Fd34-46R(36 mer), Fd47-60F(36 mer), Fd47-60R(36 mer), Fd61-72F(36 mer), Fd61-72R(36 mer), Fd73-84F(36 mer), Fd73-84R(36 mer), Fd85-97F(36 mer), Fd85-97R(36 mer), Fd98-106F (36 mer), Fd98-106R (36 mer), Fd107-115R(39 mer), d(61-97)-F(36mer), d(61-97)-R(36mer),
10 [Fv55-100]-F(48mer), [Fv55-100]-R(42mer), F(R27A)-F (27 mer), F(R27A)-R (27 mer), F(G28A)-F (27 mer), F(G28A)-R (27 mer), F(T29A)-F (27 mer), F(T29A)-R (27 mer), Bt5Fv-F (36mer), Bt5Fv-R (36mer), Dp2Fv-F (36mer), Dp2Fv-R (36mer), Bt5Dp2-F(36mer), Bt5Dp2-R(36mer).

15 Appendix B shows the sequences of fragments of Fve, which comprise all or part of the RGT motif.

Appendix C shows the crystal coordinates of Fve protein.

The methods and compositions described here may suitably employ any one or more of the sequences shown in the Appendices.

DETAILED DESCRIPTION

20 We have identified an immunoregulatory protein, designated as native Fve, from *Flammulina velutipes*. The cDNA encoding Fve protein has been isolated and biologically active recombinant Fve has been successfully produced in *E.coli*.

Our studies show that native Fve is capable of inducing high levels of expression of IFN- γ , TNF- α and ICAM-I gene expression in activated human T -and NK cells. It also

up-regulates transcription factors IRF-I and NF- κ B (c-Rel), but down-regulates IL-4. In allergic murine model, mice treated with Der p 2, a major house dust mite allergen from *Dermatophagoides pteronyssinus*, plus native Fve show a significant boost of Der p 2-specific IgG2a production. Native Fve also reduces wheel and erythematic flare formation 5 on Der p 2 skin prick test-positive human subject. We also find that fragments, homologues, variants derivatives of native Fve disclosed here (termed "Fve polypeptides") as well as nucleic acids encoding these, also have such activities.

Furthermore, we show in the Examples that Fve polypeptide and native Fve polypeptide is a potent adjuvant that can be codelivered with specific allergens for 10 desensitization of allergic disorders such as asthma, rhinitis and atopic dermatitis. In addition, Fve selectively induces polarization of NK (natural killer) cells and cytotoxic CD8 $^{+}$ T cells *in vitro* and *in vivo*. We therefore disclose anti-cancer therapies and methods which employ these immunostimulatory or immunomodulatory effects. We disclose *in vivo* animal studies which show that this protein prolongs survival rate in solid tumor- 15 transplanted mice.

Fve and its polypeptides may therefore be used for any application where up-regulation of a immune response is desired or necessary. Fve polypeptides may in particular be used in therapy, for example for the treatment of diseases such as infections, cancer, etc.

20 We further disclose a combination of Fve polypeptide or native Fve, with an allergen, for example in the form of a fusion protein. Such a combination is able to counteract an established allergic reaction. Combinations of Fve polypeptide or native Fve with a tumour associated protein or viral oncogenic protein may be used to prevent or treat cancer, or specifically for preventing tumour progression.

25 We disclose immunotherapeutic methods and reagents for allergy and virus infections, which take advantage of these immunomodulatory effects of native Fve and Fve polypeptides. We also disclose methods of treatment or prevention of a cancer,

tumour, neoplasm or cancerous cell, by use of the Fve polypeptides and nucleic acids described here. DNA vaccines, expression vectors, host cells and transgenic organisms comprising such Fve nucleic acids, or a fragment, homologue, variant or derivative thereof, may also be used for such a purpose. In general, any use of a Fve polypeptide 5 described here may employ a nucleic acid encoding such, or a DNA vaccine, expression vector, host cell and transgenic organism comprising such, and the disclosure should be read accordingly.

We also show that native Fve stimulates gene expression of human IFN- γ , TNF- α , IL-1 β , IL-2, IRF-1, c-Rel, Bcl-X_L, ICAM-1 and iNOS. In addition, we show that Fve up-10 regulates a novel subset of CD8 $^+$ T cells (CD3 $^+$ CD8 $^+$ CD18 $^{+bright}$), and induces NK cell and CD8 $^+$ T cell proliferation *in vivo*. Animal studies show that Fve prolongs survival rate of tumor-inoculated mice treated with Fve gene and protein. We disclose methods and reagents for cancer therapy using the Fve gene, protein and products, for example in the form of cell-based vaccines for cancers.

15 Fve may be used *in vitro* to stimulate the proliferation of CD3 $^+$ CD8 $^{+bright}$ CD18 $^{+bright}$ populations, and the amplified populations may then be administered to the individual in need of treatment. Thus, while it is possible to stimulate CD3 $^+$ CD8 $^{+bright}$ CD18 $^{+bright}$ populations in the context of the body of the animal, it will be apparent that such amplification is also possible *in vitro*. We therefore disclose the use of Fve 20 polypeptides to stimulate such cells *in vitro*. Such amplified populations may then be infused into or otherwise administered to the individual in need of treatment. The starting cell population may come from another individual, but preferably it is derived from the same individual who requires treatment.

We also disclose a crystal of FIP, which has been crystallised for the first time. 25 Such a crystal may be used for modelling, or designing ligands which may interact with Fve. The crystal or model may be stored on a computer, or on a computer readable medium, and manipulated using methods known to the skilled person. A computer readable medium comprising a data representation of the crystal is therefore provided.

The practice of the present invention will employ, unless otherwise indicated, conventional techniques of chemistry, molecular biology, microbiology, recombinant DNA and immunology, which are within the capabilities of a person of ordinary skill in the art. Such techniques are explained in the literature. See, for example, J. Sambrook, E. 5 F. Fritsch, and T. Maniatis, 1989, *Molecular Cloning: A Laboratory Manual*, Second Edition, Books 1-3, Cold Spring Harbor Laboratory Press; Ausubel, F. M. et al. (1995 and periodic supplements; *Current Protocols in Molecular Biology*, ch. 9, 13, and 16, John Wiley & Sons, New York, N.Y.); B. Roe, J. Crabtree, and A. Kahn, 1996, *DNA Isolation and Sequencing: Essential Techniques*, John Wiley & Sons; J. M. Polak and James O'D. 10 McGee, 1990, *In Situ Hybridization: Principles and Practice*; Oxford University Press; M. J. Gait (Editor), 1984, *Oligonucleotide Synthesis: A Practical Approach*, Irl Press; and, D. M. J. Lilley and J. E. Dahlberg, 1992, *Methods of Enzymology: DNA Structure Part A: Synthesis and Physical Analysis of DNA Methods in Enzymology*, Academic Press. Each of these general texts is herein incorporated by reference.

15 **NATIVE FVE**

The terms "native Fve polypeptide" or "native Fve protein", as used in this document, should be taken to refer to the immunoregulatory protein Fve from *Flammulina velutipes*, preferably in isolated form. The term "wild type Fve" should be understood to be synonymous with "native" Fve; furthermore, the term "nFve" is sometimes used to 20 refer to native Fve.

Preferably, "native" Fve has an amino acid sequence set out as as GenBank accession numbers: S69147 immunomodulatory protein FIP-fve - golden needle mushroom gi|7438667|pir||S69147[7438667] and P80412 IMMUNOMODULATORY PROTEIN FIP-FVE gi|729544|sp|P80412|FVE_FLAVE[729544]. A polypeptide and 25 nucleic acid sequence of "native" or "wild type" Fve is also shown in Appendix A, and the term "native FIP" preferably refers to a polypeptide or nucleic acid, as the case may be, having such sequence. Methods of isolating the "native" Fve gene and protein from *Flammulina velutipes* are known in the art, and are also set out in the Examples.

A “native” Fve may comprise a methionine residue at the N terminus; however, a native Fve may include versions which lack the initial methionine. The nucleic acid sequence which encodes such a native Fve may therefore comprise or not comprise an initial ATG codon.

5 As noted above, we have identified certain previously unknown properties of native Fve, including immunomodulatory and stimulatory properties, and one aspect of the invention is directed to such new uses of native Fve nucleic acid and native Fve polypeptide. These are disclosed in further detail below.

It should be understood, therefore, that the invention preferably does not include
10 wild-type or native Fve protein; however, it does encompass the uses of this in immunomodulation, enhancing immune response and in allergy and cancer treatment. Furthermore, we disclose a fusion protein comprising glutathione S transferase (GST) and native Fve; such a fusion protein is shown in the Examples to have the beneficial properties of native Fve itself. The sequence of GST-Fve is shown in **Appendix A**.
15 Therefore, the invention includes this GST-Fve fusion protein (also referred to as rGST-Fve and GST-Fve (wild type)), and nucleic acids encoding it.

We further disclose a nucleic acid sequence encoding native Fve, termed here a “native Fve nucleic acid sequence”. The Examples describe the cloning and isolation of a cDNA encoding native Fve protein. The sequence of this is set out as “Fve (Wild type)” in
20 **Appendix A**. Preferably such a sequence is in isolated form.

FVE POLYPEPTIDES

Additionally, we have identified various fragments, homologues, variants and derivatives of “native Fve”, which are previously unknown. Such fragments, homologues, variants and derivatives are referred to here as “Fve polypeptides” (as contrasted with
25 “native Fve polypeptides”). We disclose such Fve polypeptides, and their uses.

It will be apparent that the terms "Fve" and "Fve polypeptide", as they are used in this document, preferably exclude the wild type or native Fve protein or gene encoding this, and includes only molecules derived from native Fve, being fragments, homologues, variants and derivatives of native Fve (i.e., Fve polypeptides).

- 5 The Fve polypeptides are preferably at least as biologically active as native Fve. However, they may have 10%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, 90%, 95% or more of the biological activity of native Fve, for example as assayed by any of the tests set out below. As used herein "biologically active" refers to a sequence having a similar structural function (but not necessarily to the same degree), and/or similar regulatory function (but
10 not necessarily to the same degree), and/or similar biochemical function (but not necessarily to the same degree) of the naturally occurring sequence.

- "Fve polypeptides" preferably comprise at least one biological activity of native Fve. By "biological activity" in relation to Fve, we refer to at least one of the following activities: up-regulation of expression of Th1 cytokines, preferably IFN- γ and TNF- α ,
15 down-regulation of expression of Th2 cytokines, preferably IL-4 and IL-13, hemagglutination activity, cell aggregation activity, lymphocyte aggregation activity, lymphoproliferation activity, up-regulation of expression of IL-2, IFN- γ , TNF- α , but not IL-4 in CD3 $^{+}$ T cells, interaction with T and NK cells, adjuvant activity, stimulation of CD3 $^{+}$ CD16 $^{+}$ CD56 $^{+}$ natural killer (NK) T cells, and up-regulation of expression of
20 allergen specific IgG2a antibody. Further biological activities preferably comprised by Fve polypeptides as described here include prevention of systemic anaphylactic reactions and/or decreased footpad edema, preferably as assayed using the Arthus reaction (Ko et al, 1995). In particular, Fve polypeptides preferably comprise at least some of useful properties, preferably medically or therapeutically useful properties, of native Fve.
- 25 Assays for each of these activities are set out in the Examples, and preferably, whether a Fve polypeptide comprises a "biological activity" of Fve is to be assessed according to the relevant assay set out in the Examples.

Preferably, Fve polypeptides comprise at least one or more of the biological activities for the relevant use, preferably use as an immunomodulator, or for upregulating immune response. Preferably, they comprise at least one or more of the biological activities which enable use as a cancer therapy or allergy therapy.

5 Preferably, Fve polypeptides comprise two or more biological activities of native Fve, preferably substantially all the biological activities of native Fve.

We show in the Examples that the sequence RGT at positions 27-29 of the native Fve polypeptide sequence plays a crucial role in the biological activity of native Fve. In particular, the RGT is shown to mediate the ability of native Fve to cause lymphocyte aggregation and adhesion. This sequence is also shown to mediate lymphoproliferation, 10 and stimulation of IL-2, IFN- γ and TNF- γ secretion in T cells, preferably CD3 $^+$ T cells.

Accordingly, in preferred embodiments, the Fve polypeptides comprise at least one, two or all three of the RGT residues (or a functional variant such as RGD) at or about a position corresponding to position 28 of the native Fve polypeptide. By functional 15 variant of RGT, we mean any change in the residues of RGT (or a sequence surrounding it) which does not substantially abolish its function, preferably its function in mediating the activities set out above. Preferably, the Fve polypeptide comprises between 2 to 50, more preferably between 2 to 40, more preferably between 2 to 30, most preferably between 2 to 20 residues of amino acid sequence flanking the glycine residue 20 corresponding to position 28 of native Fve. More preferably, the Fve polypeptide comprises the sequence RGT or the sequence RGD.

However, we show that mutations of R at position 27, as well as mutations of T at position 29, have advantageous effects, in that they independently increase activity of a Fve polypeptide comprising either or both of these mutations. Furthermore, each of the 25 mutations, or in combination, have the potential to increase the solubility of the Fve polypeptide comprising it or them. One, each or both of R27 and T29 may therefore be independently mutated advantageously, by substitution or deletion.

In preferred embodiments, the or each of R27 and T29 are mutated by substitution. The R27 and / or T29 may be substituted by any other residue, but preferably a neutral residue such as G or A. We therefore disclose Fve polypeptides in which R at position 27 is changed to another residue, for example, Fve polypeptides in which R27 is mutated to 5 A, i.e., a Fve polypeptide comprising R27A. We therefore disclose Fve polypeptides in which T at position 29 is changed to another residue, for example, Fve polypeptides in which T29 is mutated to A, i.e., a Fve polypeptide comprising T29A.

Combinations are also possible; hence we disclose Fve polypeptides in which R at position 27 and T at position 29 are independently changed to one or more other residues. 10 For example, we disclose Fve polypeptides in which R27 is mutated to A, and T29 is mutated to A, i.e., a Fve polypeptide comprising R27A and T29A. As noted above, the polypeptide may comprise between 2 to 50, 40, 30 or preferably 20 residues of amino acid flanking the glycine residue at position 28 of native Fve.

Fve polypeptides may comprise fragments of native Fve. For example, Fve D6-18, 15 Fve D19-33, Fve D34-46, Fve D47-60, Fve D61-72, Fve D73-84, Fve D85-97, Fve D98-106, Fve D107-115, Fve D61-97, and Fvep55-100. Fusion proteins comprising these deletion fragments and GST are also disclosed. Fve polypeptides may comprise substitutions, including FveR27A, FveG28A and FveT29A. Further examples of Fve 20 polypeptides are shown in Appendix B, each of which includes at least a portion of the RGT sequence (preferably the whole of the RGT sequence) discussed above. Preferably, the length of such a fragment is 9 amino acid residues or more, e.g., fragment numbers 34-403.

Fve polypeptides may comprise fusion proteins, particularly fusion proteins 25 between an allergen and a Fve polypeptide as defined here. Such allergen-immunomodulator combinations include Blo t 5-Fve(two-in-one chimeric wild type), Blo t 5-FveR27A (two-in-one chimeric mutant), Blo t 5-FveT29A (two-in-one chimeric mutant), Der p 2-FveR27A (two-in-one chimeric mutant), Der p 2-FveT29A (two-in-one chimeric mutant) and Blo t 5-Der p 2-FveR27A (three-in-one chimeric mutant).

Fragments, homologues, variants and derivatives of each of these Fve polypeptides are also included.

The Fve polypeptides may be made by biochemical methods, for example, protein digestion of native Fve, or preferably by recombinant DNA methods as known in the art.

- 5 Accordingly, it will be understood that Fve polypeptides specifically include recombinant Fve polypeptides. For example, we disclose in the Examples successful production in *E.coli* of biologically active recombinant Fve polypeptide.

The Fve polypeptides disclosed also include homologous sequences obtained from any source, for example related viral/bacterial proteins, cellular homologues and synthetic peptides, as well as variants or derivatives thereof. Thus polypeptides also include those encoding homologues of Fve from other species including other microorganisms. 10 Furthermore, homologues from higher animals such as mammals (e.g. mice, rats or rabbits), especially primates, more especially humans are also included.

Homologues

- 15 In the context of this document, a "homologous" sequence is taken to include an amino acid sequence which is at least 15, 20, 25, 30, 40, 50, 60, 70, 80 or 90% identical, preferably at least 95 or 98% identical at the amino acid level over at least 10, 15, 20, 25, 30, 40, 50, 60, 70, 80, 90, 100, 110 or 114 amino acids with the sequence of native Fve shown as "Fve (Wild type)" in Appendix A. In particular, homology should typically be 20 considered with respect to those regions of the sequence known to be essential for protein function rather than non-essential neighbouring sequences. This is especially important when considering homologous sequences from distantly related organisms.

25 Although homology can also be considered in terms of similarity (i.e. amino acid residues having similar chemical properties/functions), in the context of the present document it is preferred to express homology in terms of sequence identity.

Homology comparisons can be conducted by eye, or more usually, with the aid of readily available sequence comparison programs. These publicly and commercially available computer programs can calculate % homology between two or more sequences.

- % homology may be calculated over contiguous sequences, i.e. one sequence is
- 5 aligned with the other sequence and each amino acid in one sequence directly compared with the corresponding amino acid in the other sequence, one residue at a time. This is called an "ungapped" alignment. Typically, such ungapped alignments are performed only over a relatively short number of residues (for example less than 50 contiguous amino acids).
- 10 Although this is a very simple and consistent method, it fails to take into consideration that, for example, in an otherwise identical pair of sequences, one insertion or deletion will cause the following amino acid residues to be put out of alignment, thus potentially resulting in a large reduction in % homology when a global alignment is performed. Consequently, most sequence comparison methods are designed to produce
- 15 optimal alignments that take into consideration possible insertions and deletions without penalising unduly the overall homology score. This is achieved by inserting "gaps" in the sequence alignment to try to maximise local homology.

However, these more complex methods assign "gap penalties" to each gap that occurs in the alignment so that, for the same number of identical amino acids, a sequence

20 alignment with as few gaps as possible - reflecting higher relatedness between the two compared sequences - will achieve a higher score than one with many gaps. "Affine gap costs" are typically used that charge a relatively high cost for the existence of a gap and a smaller penalty for each subsequent residue in the gap. This is the most commonly used gap scoring system. High gap penalties will of course produce optimised alignments with

25 fewer gaps. Most alignment programs allow the gap penalties to be modified. However, it is preferred to use the default values when using such software for sequence comparisons. For example when using the GCG Wisconsin Bestfit package (see below) the default gap penalty for amino acid sequences is -12 for a gap and -4 for each extension.

Calculation of maximum % homology therefore firstly requires the production of an optimal alignment, taking into consideration gap penalties. A suitable computer program for carrying out such an alignment is the GCG Wisconsin Bestfit package (University of Wisconsin, U.S.A; Devereux *et al.*, 1984, Nucleic Acids Research 12:387).

- 5 Examples of other software than can perform sequence comparisons include, but are not limited to, the BLAST package (see Ausubel *et al.*, 1999 *ibid* – Chapter 18), FASTA (Atschul *et al.*, 1990, J. Mol. Biol., 403–410) and the GENWORKS suite of comparison tools. Both BLAST and FASTA are available for offline and online searching (see Ausubel *et al.*, 1999 *ibid*, pages 7-58 to 7-60). However it is preferred to use the GCG
- 10 Bestfit program.

Although the final % homology can be measured in terms of identity, the alignment process itself is typically not based on an all-or-nothing pair comparison. Instead, a scaled similarity score matrix is generally used that assigns scores to each pairwise comparison based on chemical similarity or evolutionary distance. An example of such a matrix 15 commonly used is the BLOSUM62 matrix - the default matrix for the BLAST suite of programs. GCG Wisconsin programs generally use either the public default values or a custom symbol comparison table if supplied (see user manual for further details). It is preferred to use the public default values for the GCG package, or in the case of other software, the default matrix, such as BLOSUM62.

- 20 Advantageously, the BLAST algorithm is employed, with parameters set to default values. The BLAST algorithm is described in detail at http://www.ncbi.nih.gov/BLAST/blast_help.html, which is incorporated herein by reference. The search parameters are defined as follows, can be advantageously set to the defined default parameters.

- 25 Advantageously, “substantial identity” when assessed by BLAST equates to sequences which match with an EXPECT value of at least about 7, preferably at least about 9 and most preferably 10 or more. The default threshold for EXPECT in BLAST searching is usually 10.

BLAST (Basic Local Alignment Search Tool) is the heuristic search algorithm employed by the programs blastp, blastn, blastx, tblastn, and tblastx; these programs ascribe significance to their findings using the statistical methods of Karlin and Altschul (Karlin and Altschul 1990, *Proc. Natl. Acad. Sci. USA* 87:2264-68; Karlin and Altschul, 5 1993, *Proc. Natl. Acad. Sci. USA* 90:5873-7; see http://www.ncbi.nih.gov/BLAST/blast_help.html) with a few enhancements. The BLAST programs are tailored for sequence similarity searching, for example to identify homologues to a query sequence. For a discussion of basic issues in similarity searching of sequence databases, see Altschul *et al* (1994) *Nature Genetics* 6:119-129.

10 The five BLAST programs available at <http://www.ncbi.nlm.nih.gov> perform the following tasks: **blastp** - compares an amino acid query sequence against a protein sequence database; **blastn** - compares a nucleotide query sequence against a nucleotide sequence database; **blastx** - compares the six-frame conceptual translation products of a nucleotide query sequence (both strands) against a protein sequence database; **tblastn** - 15 compares a protein query sequence against a nucleotide sequence database dynamically translated in all six reading frames (both strands); **tblastx** - compares the six-frame translations of a nucleotide query sequence against the six-frame translations of a nucleotide sequence database.

BLAST uses the following search parameters:

20 **HISTOGRAM** - Display a histogram of scores for each search; default is yes. (See parameter H in the BLAST Manual).

DESCRIPTIONS - Restricts the number of short descriptions of matching sequences reported to the number specified; default limit is 100 descriptions. (See parameter V in the manual page).

25 **EXPECT** - The statistical significance threshold for reporting matches against database sequences; the default value is 10, such that 10 matches are expected to be found

merely by chance, according to the stochastic model of Karlin and Altschul (1990). If the statistical significance ascribed to a match is greater than the EXPECT threshold, the match will not be reported. Lower EXPECT thresholds are more stringent, leading to fewer chance matches being reported. Fractional values are acceptable. (See parameter E 5 in the BLAST Manual).

CUTOFF - Cutoff score for reporting high-scoring segment pairs. The default value is calculated from the EXPECT value (see above). HSPs are reported for a database sequence only if the statistical significance ascribed to them is at least as high as would be ascribed to a lone HSP having a score equal to the CUTOFF value. Higher CUTOFF 10 values are more stringent, leading to fewer chance matches being reported. (See parameter S in the BLAST Manual). Typically, significance thresholds can be more intuitively managed using EXPECT.

ALIGNMENTS - Restricts database sequences to the number specified for which high-scoring segment pairs (HSPs) are reported; the default limit is 50. If more database 15 sequences than this happen to satisfy the statistical significance threshold for reporting (see EXPECT and CUTOFF below), only the matches ascribed the greatest statistical significance are reported. (See parameter B in the BLAST Manual).

MATRIX - Specify an alternate scoring matrix for BLASTP, BLASTX, TBLASTN and TBLASTX. The default matrix is BLOSUM62 (Henikoff & Henikoff, 1992). The 20 valid alternative choices include: PAM40, PAM120, PAM250 and IDENTITY. No alternate scoring matrices are available for BLASTN; specifying the MATRIX directive in BLASTN requests returns an error response.

STRAND - Restrict a TBLASTN search to just the top or bottom strand of the database sequences; or restrict a BLASTN, BLASTX or TBLASTX search to just reading 25 frames on the top or bottom strand of the query sequence.

FILTER - Mask off segments of the query sequence that have low compositional complexity, as determined by the SEG program of Wootton & Federhen (1993) Computers and Chemistry 17:149-163, or segments consisting of short-periodicity internal repeats, as determined by the XNU program of Claverie & States (1993) Computers and Chemistry 5 17:191-201, or, for BLASTN, by the DUST program of Tatusov and Lipman (see <http://www.ncbi.nlm.nih.gov>). Filtering can eliminate statistically significant but biologically uninteresting reports from the blast output (e.g., hits against common acidic-, basic- or proline-rich regions), leaving the more biologically interesting regions of the query sequence available for specific matching against database sequences.

10 Low complexity sequence found by a filter program is substituted using the letter "N" in nucleotide sequence (e.g., "NNNNNNNNNNNNNN") and the letter "X" in protein sequences (e.g., "XXXXXXXXXX").

Filtering is only applied to the query sequence (or its translation products), not to database sequences. Default filtering is DUST for BLASTN, SEG for other programs.

15 It is not unusual for nothing at all to be masked by SEG, XNU, or both, when applied to sequences in SWISS-PROT, so filtering should not be expected to always yield an effect. Furthermore, in some cases, sequences are masked in their entirety, indicating that the statistical significance of any matches reported against the unfiltered query sequence should be suspect.

20 NCBI-gi - Causes NCBI gi identifiers to be shown in the output, in addition to the accession and/or locus name.

Most preferably, sequence comparisons are conducted using the simple BLAST search algorithm provided at <http://www.ncbi.nlm.nih.gov/BLAST>. In some embodiments, no gap penalties are used when determining sequence identity.

Once the software has produced an optimal alignment, it is possible to calculate % homology, preferably % sequence identity. The software typically does this as part of the sequence comparison and generates a numerical result.

Variants and Derivatives

5 The terms "variant" or "derivative" in relation to the amino acid sequences disclosed here includes any substitution of, variation of, modification of, replacement of, deletion of or addition of one (or more) amino acids from or to the sequence providing the resultant amino acid sequence retains substantially the same activity as the unmodified sequence. Preferably, the modified sequence has at least one biological activity as the
10 unmodified sequence, preferably all the biological activities of the unmodified sequence. Preferably, the "variant" or "derivative" has at least one biological activity of native Fve, as described above.

Polypeptides having the amino acid sequence shown in the description and Examples, or fragments or homologues thereof may be modified for use in the methods
15 and compositions described here. Typically, modifications are made that maintain the biological activity of the sequence. Amino acid substitutions may be made, for example from 1, 2 or 3 to 10, 20 or 30 substitutions provided that the modified sequence retains the biological activity of the unmodified sequence. Alternatively, modifications may be made to deliberately inactivate one or more functional domains of the polypeptides described
20 here. Functional domains of native Fve include the α helix at the N terminus, any of the six β helices, as well as the "loop-like" structures at the N and C termini. Preferably, the functional domain of native Fve comprises the N-terminus helix and the loop/strand, which are essential for protein dimerization.

Amino acid substitutions may include the use of non-naturally occurring
25 analogues, for example to increase blood plasma half-life of a therapeutically administered polypeptide.

Conservative substitutions may be made, for example according to the Table below. Amino acids in the same block in the second column and preferably in the same line in the third column may be substituted for each other:

ALIPHATIC	Non-polar	G A P
		I L V
	Polar - uncharged	C S T M
		N Q
	Polar - charged	D E
		K R
AROMATIC		H F W Y

Polypeptides also include fragments of the full length sequence of native Fve, or
 5 any of the Fve polypeptides disclosed here. Preferably fragments comprise at least one epitope. Methods of identifying epitopes are well known in the art. Fragments will typically comprise at least 6 amino acids, more preferably at least 10, 20, 30, 50 or 100 amino acids.

Fve polypeptides, fragments, homologues, variants and derivatives, are typically
 10 made by recombinant means, for example as described below in the Examples. However they may also be made by synthetic means using techniques well known to skilled persons such as solid phase synthesis. The proteins may also be produced as fusion proteins, for example to aid in extraction and purification. Examples of fusion protein partners include glutathione-S-transferase (GST), 6xHis, GAL4 (DNA binding and/or transcriptional
 15 activation domains) and β-galactosidase. It may also be convenient to include a proteolytic cleavage site between the fusion protein partner and the protein sequence of interest to allow removal of fusion protein sequences. Preferably the fusion protein will not hinder the function of the protein of interest sequence. Proteins may also be obtained by purification of cell extracts from animal cells.

20 The Fve polypeptides, variants, homologues, fragments and derivatives disclosed here may be in a substantially isolated form. It will be understood that such polypeptides

may be mixed with carriers or diluents which will not interfere with the intended purpose of the protein and still be regarded as substantially isolated. A Fve variant, homologue, fragment or derivative may also be in a substantially purified form, in which case it will generally comprise the protein in a preparation in which more than 90%, e.g. 95%, 98% or 5 99% of the protein in the preparation is a protein.

The Fve polypeptides, variants, homologues, fragments and derivatives disclosed here may be labelled with a revealing label. The revealing label may be any suitable label which allows the polypeptide, etc to be detected. Suitable labels include radioisotopes, e.g. ¹²⁵I, enzymes, antibodies, polynucleotides and linkers such as biotin. Labelled polypeptides

10 may be used in diagnostic procedures such as immunoassays to determine the amount of a polypeptide in a sample. Polypeptides or labelled polypeptides may also be used in serological or cell-mediated immune assays for the detection of immune reactivity to said polypeptides in animals and humans using standard protocols.

A Fve polypeptide, variant, homologue, fragment or derivative disclosed here, 15 optionally labelled, may also be fixed to a solid phase, for example the surface of an immunoassay well or dipstick. Such labelled and/or immobilised polypeptides may be packaged into kits in a suitable container along with suitable reagents, controls, instructions and the like. Such polypeptides and kits may be used in methods of detection of antibodies to the polypeptides or their allelic or species variants by immunoassay.

20 Immunoassay methods are well known in the art and will generally comprise: (a) providing a polypeptide comprising an epitope bindable by an antibody against said protein; (b) incubating a biological sample with said polypeptide under conditions which allow for the formation of an antibody-antigen complex; and (c) determining whether antibody-antigen complex comprising said polypeptide is formed.

25 The Fve polypeptides, variants, homologues, fragments and derivatives disclosed here may be used in *in vitro* or *in vivo* cell culture systems to study the role of their corresponding genes and homologues thereof in cell function, including their function in

disease. For example, truncated or modified polypeptides may be introduced into a cell to disrupt the normal functions which occur in the cell. The polypeptides may be introduced into the cell by *in situ* expression of the polypeptide from a recombinant expression vector (see below). The expression vector optionally carries an inducible promoter to control the 5 expression of the polypeptide.

The use of appropriate host cells, such as insect cells or mammalian cells, is expected to provide for such post-translational modifications (e.g. myristylation, glycosylation, truncation, lapidation and tyrosine, serine or threonine phosphorylation) as may be needed to confer optimal biological activity on recombinant expression products.

10 Such cell culture systems in which the Fve polypeptides, variants, homologues, fragments and derivatives disclosed here are expressed may be used in assay systems to identify candidate substances which interfere with or enhance the functions of the polypeptides in the cell.

IMMUNOMODULATOR-ANTIGEN COMBINATIONS AND CONJUGATES

15 We show throughout this document (for the first time) that Fve has immunomodulatory properties, and in particular can act to potentiate an immune response. The adjuvant property of Fve may be exploited by administering Fve polypeptide or nucleic acid (or a fragment, homologue, variant or derivative thereof, or a host cell or vector comprising such) as described below, along with a molecule to which an immune 20 response is desired.

The Fve polypeptide, etc may be administered to an individual either in combination, sequentially or simultaneously or in succession with the molecule to which an immune response is desired. We therefore provide for the first time a combination of a Fve polypeptide, etc with an antigenic molecule.

25 Where the Fve polypeptide, etc and the molecule are administered in combination, this may be achieved by administering a mixture of the Fve polypeptide, etc and the

molecule. We therefore provide a simple combination of the Fve polypeptide, etc and the molecule, preferably as a kit. The kit may comprise the Fve polypeptide, etc and the molecule to which an immune response is desired in separate containers, and may optionally comprise instructions to administer these simultaneously, sequentially, etc.

- 5 The molecule to which an immune response is desired may comprise an allergen. These are set out in further detail in the following section.

The molecule to which an immune response is desired may comprise a tumour associated antigen. In preferred embodiments, the tumour associated antigen comprises MAGE-1, MAGE-2, MAGE-3, BAGE, GAGE, PRAME, SSX-2, Tyrosinase, MART-1, 10 NY-ESO-1, gp100, TRP-1, TRP-2, A2 melanotope, BCR/ABL, Proteinase-3/Myeloblastin, HER2/neu, CEA, P1A, HK2, PAPA, PSA, PSCA, PSMA, pg75, MUM-1, MUC-1, BTA, GnT-V, β -catenin, CDK4, or P15. Nucleic acid and amino acid sequences of these antigens are known in the art, and the skilled person will know how to produce tumour associated antigens, including those set out above. We therefore disclose 15 combinations, preferably in the form of kits, comprising an Fve polypeptide or nucleic acid (or a fragment, homologue, variant or derivative thereof, or a host cell or vector comprising such), together with a tumour associated antigen, for example as set out above.

The molecule to which an immune response is desired may comprise a viral antigen. In preferred embodiments, the viral antigen comprises a protein from an 20 oncogenic virus; such viruses are known in the art. Preferably the oncogenic viral antigen comprises E6 and E7 from HPV; core Ag and E2 from HCV; core and surface antigens from HBV; LMP-1, LMP-2, EBNA-2, EBNA-3 from EBV; or Tax from HTLV-1.

In a further embodiment, the viral antigen comprises an antigen, preferably a protein, more preferably an antigenic protein or fragment thereof from an infectious virus. 25 Such immunomodulator-viral antigen conjugates may be used to treat or prevent a viral infectious disease, i.e., the cognate disease. For example, an immunomodulator-HSV antigen conjugate, for example, a Fve polypeptide-HSV antigen conjugate, may be used to

treat or prevent Herpes Simplex Virus infection. Other preferred viral antigens include those from Adenovirus, Parainfluenza 3 virus, Human Immunodeficiency Virus (HIV), Herpes simplex virus, HSV, Respiratory syncytial virus, RSV, and Influenza A, Flu A. These viruses, and the diseases they cause, are well known in the art, and methods for 5 making and purifying antigens from such viruses are also well known. For example, US Patent Number 4,313,927 (Fridlender) discloses detailed protocols for preparation of rubella and Cytomegalovirus (CMV) antigen.

Nucleic acid and amino acid sequences of these viral antigens are known in the art, and the skilled person will know how to produce viral antigen antigens, including these set 10 out above. We therefore disclose combinations, preferably in the form of kits, comprising an Fve polypeptide or nucleic acid (or a fragment, homologue, variant or derivative thereof, or a host cell or vector comprising such), together with a viral antigen, for example as set out above.

In preferred embodiments, we provide administration of the Fve polypeptide, etc 15 and the molecule to which an immune response is desired, in which there is some degree of association between the Fve polypeptide, etc and the molecule in question.

We therefore disclose for the first time an agent which comprises an immunomodulator coupled, fused, mixed, combined, or otherwise joined to an allergen. Such a construct is referred to as a "immunomodulator-allergen conjugate" in this 20 document. In particular, we disclose the use of Fve adjuvanted allergen vaccines, as explained in further detail in Examples 13 and 14.

The coupling, etc between the immunomodulator and the allergen may be permanent or transient, and may involve covalent or non-covalent interactions (including ionic interactions, hydrophobic forces, Van der Waals interactions, etc). The exact mode of 25 coupling is not important, so long as the immunomodulator-allergen conjugate. Accordingly, where reference is made to "comprising", "conjugation", "coupling", etc,

these references should be taken to include any form of interaction between the immunomodulator and the allergen.

Thus, the immunomodulator may be a polypeptide which is provided as a fusion protein with the allergen, for example as shown in Example 13 for F've/Allergen. An expression vector may be constructed by standard recombinant DNA technology to include a nucleotide sequence capable of expressing a immunomodulator, such that a fusion protein is expressed comprising the allergen of interest fused to the immunomodulator. The expression vector is transfected or transformed into a suitable host for large scale production of fusion protein, by means known in the art. Purification of the fusion protein may also be carried out by known means. Alternatively, or in addition, and as discussed above, the allergen may be physically associated with the immunomodulator, and attached to it by chemical conjugation. Thus, Example 14 below describes the use of allergen physically conjugated to F've.

In preferred embodiments, the immunomodulator-allergen conjugate is capable of at least one of the following, preferably two or more, more preferably all: increase the number of human PBMC, to stimulate the proliferation of human lymphocytes, to polarize human CD8⁺ T cells, and to increase the production of IFN-γ (Th1 response) and IL-10 (Tr response). Preferably, the immunomodulator-allergen conjugate is capable of inducing both Th1 and Tr immune responses. Preferably, the Th1 response inhibits the development of Th2 cells via IFN-γ, more preferably it is capable of inducing a life-long (or substantially long lasting) protective Th1 memory immune response. Allergen specific Tr cells may in turn dampen the anti-allergic Th1 immune response, ensuring a well-balanced protective but nonpathological Th1 response. Allergen-F've fusion proteins meet these criteria since they induce cytokine IL-10, and these are therefore preferred.

Where the conjugate comprises F've, the F've portion of the conjugate may comprise the whole molecule, or fragments of it. It may for example comprise the native F've, or any F've polypeptide as disclosed above. The allergen portion may comprise any allergen, whether proteinaceous or not. Advantageously, proteinaceous allergens are conjugated to

the immunomodulator portion by means of covalent bonds, for example, amide bonds (for example, as a fusion protein).

The allergen may comprise for example the whole or a portion of Blo t 5 or Der p 2 allergen. In highly preferred embodiments, the immunomodulator-allergen conjugate 5 comprises Bt5-Fve, Bt5-FveR27 or GST-Dp2-FveR27. Examples of other allergens suitable for use in the immunomodulator-allergen conjugate described here are provided below.

Furthermore, protein-protein conjugation also provides a convenient and alternative choice to develop allergen vaccine. Any suitable means of conjugation, for 10 example, chemical conjugation may be used to couple the immunomodulator and the allergen. Cross-linkers, for example, heterobifunctional cross linkers are known in the art, and may be used. Furthermore, other conjugation agents, for example, poly-lactic acid (PLA) and polyethylene glycol (PEG) may also be employed.

ALLERGENS

15 In general, the allergen from which an immunomodulator-allergen conjugate may be constructed may come from any source, for example, a source known to induce allergenic responses in humans. For example, the allergen may comprise a tree pollen allergen, a grass pollen allergen, a weed pollen allergen, a feline antigen, or a fungal allergen. Thus, the allergen may comprise a tree pollen allergen, for example Bet v 1 and 20 Bet v 2 from birch tree. It may comprise a grass pollen allergen, for example, Phl p 1 and Phl p 2 from timothy grass. It may comprise a weed pollen allergen, for example, antigen E from ragweed. It may comprise a major feline antigen, for example, Fel d 1. It may comprise a major fungal allergen, for example, Asp f1, Asp f2, and Asp f3 from *Aspergillus fumigatus*.

25 In preferred embodiments, the allergen comprises a dust mite allergen, preferably a house dust mite allergen. In particular, the allergen is preferably derived from a mite from

Family Glycyphagidae or Family Pyroglyphidae. Dust mites of Family Glycyphagidae include those in the genera Aeroglyphus, Austroglycyphagus, Blomia, Ctenoglyphus, Glycyphagus, Gohieria, Lepidoglyphus. Dust mites of Family Pyroglyphidae include those in the genera Dermatophagoïdes, Euroglyphus, Pyroglyphus. In preferred embodiments,
5 the allergen is preferably an allergen from a species in any of these genera.

In highly preferred embodiments, the allergen is a group 1 allergen (Der p 1, Der f 1, Blo t 1, Eur m1, Lep d 1), a group 2 allergen (Der p 2, Der f 2, Blo t 2, Eur m 2, Lep d 2), a group 5 allergen (Blo t 5, Der p 5, Der f 5, Eur m 5, Lep d 5) or a group 15 allergen (Der p 15, Der f 15, Blot 15, Eur m 15, Lep d 15) from dust mite. Nucleic acid and amino acid sequences of these allergens are known in the art, and the skilled person will know how to produce allergen-immunomodulator conjugates from any of these allergens using such sequences.
10

OTHER IMMUNOMODULATOR CONJUGATES

Immunomodulator-Tumour Associated Antigen Conjugates

15 We also disclose for the first time an agent which comprises an immunomodulator coupled, fused, mixed, combined, or otherwise joined to an tumour associated antigen. Such a construct is referred to as a "immunomodulator-tumour associated antigen conjugate" in this document.

As the term is used here, "tumour associated antigen" generally includes a cancer
20 protein or a cancer antigen, i.e., a protein which is preferentially expressed in a tumour cell or a transformed cell, compared to a "normal" non-cancerous cell.

In highly preferred embodiments, the tumour associated antigen may comprise MAGE-1, MAGE-2, MAGE-3, BAGE, GAGE, PRAME, SSX-2, Tyrosinase, MART-1, NY-ESO-1, gp100, TRP-1, TRP-2, A2 melanotope, BCR/ABL, Proteinase-
25 3/Myeloblastin, HER2/neu, CEA, P1A, HK2, PAPA, PSA, PSCA, PSMA, pg75, MUM-1, MUC-1, BTA, GnT-V, β -catenin, CDK4, or P15. Nucleic acid and amino acid sequences

of these antigens are known in the art, and the skilled person will know how to produce tumour associated antigen-immunomodulator conjugates from any of these allergens using such sequences.

We present in Appendix A the sequences of MAGE3-FveT29A, MART1-

- 5 FveT29A and CEA-FveT29A, which are preferred Immunomodulator-Tumour Associated Antigen Conjugates suitable for use in the methods and compositions described here.

Immunomodulator-Viral Antigen Conjugates

We further disclose an agent comprising an immunomodulator coupled, etc to a viral antigen. In highly preferred embodiments, the viral antigen comprises a protein from an oncogenic virus; such viruses are known in the art. Preferably the oncogenic viral antigen comprises E6 and E7 from HPV; core Ag and E2 from HCV; core and surface antigens from HBV; LMP-1, LMP-2, EBNA-2, EBNA-3 from EBV; or Tax from HTLV-1. Nucleic acid and amino acid sequences of these viral antigens are known in the art, and the skilled person will know how to produce viral antigen-immunomodulator conjugates from any of these allergens using such sequences.

We also provide an agent (for example a polypeptide) comprising a first portion comprising at least a portion of Fve and a second portion comprising at least a portion of a viral antigen, preferably coupled together. The viral antigen may be selected from the group consisting of antigens from Adenovirus, Parainfluenza 3 virus, Herpes simplex virus, HSV, Respiratory syncytial virus, RSV, and Influenza A, Flu A.

The viral antigen may comprise any portion of the native viral antigen, for example, a portion of the HCV core antigen. We have established that a deletion of the HCV core antigen, particularly a deletion of 23 amino acids from residues 141 to 163 of the core antigen leads to an increase in efficiency of protein production. Accordingly, we provide an agent comprising an immunomodulator coupled, etc to a viral antigen, which viral antigen comprises such a deleted core antigen (here referred to as "Core23"), e.g., the fusion protein HCV Core23-FveT29A.

In particular, we find that the polypeptides HCV Core23-FveT29A and HPV E7-FveT29A (the sequences of which are shown in **Appendix A**) are particularly useful as Immunomodulator-Viral Antigen conjugates.

The coupling, etc between the immunomodulator and the tumour associated
5 antigen, and the viral antigen, may be as described above for the immunomodulator-allergen conjugate.

FVE NUCLEIC ACIDS

We provide for a nucleic acid encoding a Fve polypeptide, which we refer to as a “Fve nucleic acid”. We also provide nucleic acids encoding variants, homologues,
10 derivatives and fragments of native Fve, as well as fragments, homologues, derivatives and variants of Fve nucleic acids.

Preferably, the Fve nucleic acid is derived from a natural or native Fve sequence, for example, the nucleic sequence shown as “Fve (Wild type)” in **Appendix A**. In a preferred embodiment, the Fve nucleic acid is a recombinant fragment of native Fve
15 nucleic acid, or any fragment, homologue, variant or derivative thereof. Fragments, homologues, variants and derivatives of each of the above sequences are also included.

“Fve nucleic acids” preferably encode polypeptides which have at least one biological activity of native Fve, as described above. Preferably, Fve nucleic acids encode polypeptides which comprise two or more biological activities of native Fve, preferably
20 substantially all the biological activities of native Fve.

In preferred embodiments, the Fve nucleic acids encode polypeptides which comprise at least one, two or all three of the RGT residues (or a functional variant as defined above, such as RGD) at or about a position corresponding to position 28 of the native Fve polypeptide. In particular, the Fve nucleic acid may comprise the sequence
25 CGTGGTACC. Alternatively, the Fve nucleic acid may comprise the sequence

CGTGGTGAT or the sequence CGTGGTGAC. The Fve nucleic acid may comprise a nucleotide sequence which encodes the same amino acids as a result of the redundancy of the genetic code.

The Fve nucleic acid may comprise a sequence comprising three codons, with a first 5 codon selected from the group consisting of: CGT, CGC, CGA, CGG, AGA and AGG, a second codon selected from the group consisting of: GGT, GGC, GGA and GGG, and a third codon selected from the group consisting of: ACT, ACC, ACA and ACG. Alternatively, the third codon may be selected from the group consisting of: GAT and GAC,

10 Preferably, the Fve polypeptide comprises between 2 to 60 residues of nucleic acid sequence flanking the codon for the glycine residue corresponding to position 28 of native Fve.

In preferred embodiments, Fve nucleic acids may comprise nucleic acids encoding fragments of native Fve. For example, Fve nucleic acids may comprise the nucleic acid 15 sequences depicted in Appendix A as Fve D6-18, Fve D19-33, Fve D34-46, Fve D47-60, Fve D61-72, Fve D73-84, Fve D85-97, Fve D98-106, Fve D107-115, Fve D61-97, and Fvep55-100. Nucleic acids encoding fusion proteins comprising these deletion fragments and GST are also disclosed. Fve nucleic acids may comprise those encoding substitutions, including FveR27A, FveG28A and FveT29A. Fve nucleic acids include those which 20 encode the polypeptide sequences shown in Appendix A.

We also disclose Fve nucleic acids which encode Fve polypeptides comprising fusion proteins, particularly fusion proteins between an allergen and a Fve polypeptide as defined here. We disclose in particular nucleic acid sequences of Blo t 5-Fve(two-in-one chimeric wild type), Blo t 5-FveR27A (two-in-one chimeric mutant), Blo t 5-FveT29A 25 (two-in-one chimeric mutant), Der p 2-FveR27A (two-in-one chimeric mutant), Der p 2-FveT29A (two-in-one chimeric mutant) and Blo t 5-Der p 2-FveR27A (three-in-one chimeric mutant), and shown in Appendix A.

As used here in this document, the terms "polynucleotide", "nucleotide", and nucleic acid are intended to be synonymous with each other. "Polynucleotide" generally refers to any polyribonucleotide or polydeoxribonucleotide, which may be unmodified RNA or DNA or modified RNA or DNA. "Polynucleotides" include, without limitation 5 single- and double-stranded DNA, DNA that is a mixture of single- and double-stranded regions, single- and double-stranded RNA, and RNA that is mixture of single- and double-stranded regions, hybrid molecules comprising DNA and RNA that may be single-stranded or, more typically, double-stranded or a mixture of single- and double-stranded regions. In addition, "polynucleotide" refers to triple-stranded regions comprising RNA or DNA or 10 both RNA and DNA. The term polynucleotide also includes DNAs or RNAs containing one or more modified bases and DNAs or RNAs with backbones modified for stability or for other reasons. "Modified" bases include, for example, tritylated bases and unusual bases such as inosine. A variety of modifications has been made to DNA and RNA; thus, "polynucleotide" embraces chemically, enzymatically or metabolically modified forms of 15 polynucleotides as typically found in nature, as well as the chemical forms of DNA and RNA characteristic of viruses and cells. "Polynucleotide" also embraces relatively short polynucleotides, often referred to as oligonucleotides.

It will be understood by a skilled person that numerous different polynucleotides and nucleic acids can encode the same polypeptide as a result of the degeneracy of the 20 genetic code. In addition, it is to be understood that skilled persons may, using routine techniques, make nucleotide substitutions that do not affect the polypeptide sequence encoded by the polynucleotides described here to reflect the codon usage of any particular host organism in which the polypeptides are to be expressed.

Fve nucleic acids, variants, fragments, derivatives and homologues may comprise 25 DNA or RNA. They may be single-stranded or double-stranded. They may also be polynucleotides which include within them synthetic or modified nucleotides. A number of different types of modification to oligonucleotides are known in the art. These include methylphosphonate and phosphorothioate backbones, addition of acridine or polylysine chains at the 3' and/or 5' ends of the molecule. For the purposes of this document, it is to

be understood that the polynucleotides may be modified by any method available in the art. Such modifications may be carried out in order to enhance the *in vivo* activity or life span of polynucleotides of interest.

The terms "variant", "homologue" or "derivative" in relation to a nucleotide sequence include any substitution of, variation of, modification of, replacement of, deletion of or addition of one (or more) nucleic acid from or to the sequence. Preferably said variant, homologues or derivatives code for a polypeptide having biological activity.

As indicated above, with respect to sequence homology, preferably there is at least 50 or 75%, more preferably at least 85%, more preferably at least 90% homology to the sequences shown in the sequence listing herein. More preferably there is at least 95%, more preferably at least 98%, homology. Nucleotide homology comparisons may be conducted as described above. A preferred sequence comparison program is the GCG Wisconsin Bestfit program described above. The default scoring matrix has a match value of 10 for each identical nucleotide and -9 for each mismatch. The default gap creation penalty is -50 and the default gap extension penalty is -3 for each nucleotide.

We further describe nucleotide sequences that are capable of hybridising selectively to the sequences presented herein, or any variant, fragment or derivative thereof, or to the complement of any of the above. Nucleotide sequences are preferably at least 15 nucleotides in length, more preferably at least 20, 30, 40 or 50 nucleotides in length.

The term "hybridization" as used herein shall include "the process by which a strand of nucleic acid joins with a complementary strand through base pairing" as well as the process of amplification as carried out in polymerase chain reaction technologies.

Polynucleotides capable of selectively hybridising to the nucleotide sequences presented herein, or to their complement, will be generally at least 70%, preferably at least 80 or 90% and more preferably at least 95% or 98% homologous to the corresponding

nucleotide sequences presented herein over a region of at least 20, preferably at least 25 or 30, for instance at least 40, 60 or 100 or more contiguous nucleotides.

The term "selectively hybridizable" means that the polynucleotide used as a probe is used under conditions where a target polynucleotide is found to hybridize to the probe at a level significantly above background. The background hybridization may occur because of other polynucleotides present, for example, in the cDNA or genomic DNA library being screened. In this event, background implies a level of signal generated by interaction between the probe and a non-specific DNA member of the library which is less than 10 fold, preferably less than 100 fold as intense as the specific interaction observed with the target DNA. The intensity of interaction may be measured, for example, by radiolabelling the probe, e.g. with ^{32}P .

Hybridization conditions are based on the melting temperature (T_m) of the nucleic acid binding complex, as taught in Berger and Kimmel (1987, Guide to Molecular Cloning Techniques, Methods in Enzymology, Vol 152, Academic Press, San Diego CA), and confer a defined "stringency" as explained below.

Maximum stringency typically occurs at about $T_m - 5^\circ\text{C}$ (5°C below the T_m of the probe); high stringency at about 5°C to 10°C below T_m ; intermediate stringency at about 10°C to 20°C below T_m ; and low stringency at about 20°C to 25°C below T_m . As will be understood by those of skill in the art, a maximum stringency hybridization can be used to identify or detect identical polynucleotide sequences while an intermediate (or low) stringency hybridization can be used to identify or detect similar or related polynucleotide sequences.

In a preferred aspect, we provide nucleotide sequences that can hybridise to the Fve nucleic acids, fragments, variants, homologues or derivatives disclosed here under stringent conditions (e.g. 65°C and 0.1xSSC {1xSSC = 0.15 M NaCl, 0.015 M $\text{Na}_3\text{Citrate}$ pH 7.0}).

Where the polynucleotide is double-stranded, both strands of the duplex, either individually or in combination, are encompassed by the methods and compositions described here. Where the polynucleotide is single-stranded, it is to be understood that the complementary sequence of that polynucleotide is also included.

5 Polynucleotides which are not 100% homologous to the Fve sequences disclosed here but which are also included can be obtained in a number of ways. Other variants of the sequences may be obtained for example by probing DNA libraries made from a range of individuals, for example individuals from different populations. For example, Fve homologues may be identified from other individuals, or other species. Further 10 recombinant Fve nucleic acids and polypeptides may be produced by identifying corresponding positions in the homologues, and synthesising or producing the molecule as described elsewhere in this document. Furthermore, the collagen region, neck region and carbohydrate binding domain in such homologues may be identified, for example, by sequence gazing or computer assisted comparisons, and selected for combination into or 15 production of a recombinant Fve which has one or more biological activities of native Fve.

In addition, other viral/bacterial, or cellular homologues of Fve particularly cellular homologues found in mammalian cells (e.g. rat, mouse, bovine and primate cells), may be obtained and such homologues and fragments thereof in general will be capable of selectively hybridising to Fve. Such homologues may be used to design non-human Fve 20 nucleic acids, fragments, variants and homologues. Mutagenesis may be carried out by means known in the art to produce further variety.

Sequences of Fve homologues may be obtained by probing cDNA libraries made from or genomic DNA libraries from other animal or non-animal species, particularly microbial or fungal species, and probing such libraries with probes comprising all or part 25 of any of the Fve nucleic acids, fragments, variants and homologues, or other fragments of Fve under conditions of medium to high stringency.

Similar considerations apply to obtaining species homologues and allelic variants of the polypeptide or nucleotide sequences disclosed here.

Variants and strain/species homologues may also be obtained using degenerate PCR which will use primers designed to target sequences within the variants and homologues encoding conserved amino acid sequences within the sequences of the Fve nucleic acids. Conserved sequences can be predicted, for example, by aligning the amino acid sequences from several variants/homologues. Sequence alignments can be performed using computer software known in the art. For example the GCG Wisconsin PileUp program is widely used.

The primers used in degenerate PCR will contain one or more degenerate positions and will be used at stringency conditions lower than those used for cloning sequences with single sequence primers against known sequences. It will be appreciated by the skilled person that overall nucleotide homology between sequences from distantly related organisms is likely to be very low and thus in these situations degenerate PCR may be the method of choice rather than screening libraries with labelled fragments the Fve sequences.

In addition, homologous sequences may be identified by searching nucleotide and/or protein databases using search algorithms such as the BLAST suite of programs.

Alternatively, such polynucleotides may be obtained by site directed mutagenesis of characterised sequences, for example, Fve nucleic acids, or variants, homologues, derivatives or fragments thereof. This may be useful where for example silent codon changes are required to sequences to optimise codon preferences for a particular host cell in which the polynucleotide sequences are being expressed. Other sequence changes may be desired in order to introduce restriction enzyme recognition sites, or to alter the property or function of the polypeptides encoded by the polynucleotides.

The polynucleotides described here may be used to produce a primer, e.g. a PCR primer, a primer for an alternative amplification reaction, a probe e.g. labelled with a revealing label by conventional means using radioactive or non-radioactive labels, or the polynucleotides may be cloned into vectors. Such primers, probes and other fragments will
5 be at least 8, 9, 10, or 15, preferably at least 20, for example at least 25, 30 or 40 nucleotides in length, and are also encompassed by the term "polynucleotides" as used herein.

Polynucleotides such as a DNA polynucleotides and probes may be produced recombinantly, synthetically, or by any means available to those of skill in the art. They
10 may also be cloned by standard techniques.

In general, primers will be produced by synthetic means, involving a step wise manufacture of the desired nucleic acid sequence one nucleotide at a time. Techniques for accomplishing this using automated techniques are readily available in the art.

Longer polynucleotides will generally be produced using recombinant means, for
15 example using a PCR (polymerase chain reaction) cloning techniques. This will involve making a pair of primers (e.g. of about 15 to 30 nucleotides) flanking a region of the lipid targeting sequence which it is desired to clone, bringing the primers into contact with mRNA or cDNA obtained from an animal or human cell, performing a polymerase chain reaction under conditions which bring about amplification of the desired region, isolating
20 the amplified fragment (e.g. by purifying the reaction mixture on an agarose gel) and recovering the amplified DNA. The primers may be designed to contain suitable restriction enzyme recognition sites so that the amplified DNA can be cloned into a suitable cloning vector

Polynucleotides or primers may carry a revealing label. Suitable labels include
25 radioisotopes such as ^{32}P or ^{35}S , enzyme labels, or other protein labels such as biotin. Such labels may be added to polynucleotides or primers and may be detected using by techniques known *per se*. Polynucleotides or primers or fragments thereof labelled or

unlabeled may be used by a person skilled in the art in nucleic acid-based tests for detecting or sequencing polynucleotides in the human or animal body.

Such tests for detecting generally comprise bringing a biological sample containing DNA or RNA into contact with a probe comprising a polynucleotide or primer under hybridising conditions and detecting any duplex formed between the probe and nucleic acid in the sample. Such detection may be achieved using techniques such as PCR or by immobilising the probe on a solid support, removing nucleic acid in the sample which is not hybridised to the probe, and then detecting nucleic acid which has hybridised to the probe. Alternatively, the sample nucleic acid may be immobilised on a solid support, and the amount of probe bound to such a support can be detected. Suitable assay methods of this and other formats can be found in for example WO89/03891 and WO90/13667.

Tests for sequencing nucleotides, for example, the Fve nucleic acids, involve bringing a biological sample containing target DNA or RNA into contact with a probe comprising a polynucleotide or primer under hybridising conditions and determining the sequence by, for example the Sanger dideoxy chain termination method (see Sambrook *et al.*).

Such a method generally comprises elongating, in the presence of suitable reagents, the primer by synthesis of a strand complementary to the target DNA or RNA and selectively terminating the elongation reaction at one or more of an A, C, G or T/U residue; allowing strand elongation and termination reaction to occur; separating out according to size the elongated products to determine the sequence of the nucleotides at which selective termination has occurred. Suitable reagents include a DNA polymerase enzyme, the deoxynucleotides dATP, dCTP, dGTP and dTTP, a buffer and ATP. Dideoxynucleotides are used for selective termination.

PROTEIN EXPRESSION AND PURIFICATION

Host cells comprising polynucleotides may be used to express polypeptides, such as Fve polypeptides, fragments, homologues, variants or derivatives thereof. Host cells may be cultured under suitable conditions which allow expression of the proteins.

- 5 Expression of the polypeptides may be constitutive such that they are continually produced, or inducible, requiring a stimulus to initiate expression. In the case of inducible expression, protein production can be initiated when required by, for example, addition of an inducer substance to the culture medium, for example dexamethasone or IPTG.

10 Polypeptides can be extracted from host cells by a variety of techniques known in the art, including enzymatic, chemical and/or osmotic lysis and physical disruption.

Polypeptides may also be produced recombinantly in an *in vitro* cell-free system, such as the TnTTM (Promega) rabbit reticulocyte system.

FVE NUCLEIC ACID MOLECULES

We disclose a nucleic molecule that: a) has a strand that encodes an Fve polypeptide disclosed here, b) has a strand that is complementary with a strand as described in a) above; or c) has a strand that hybridises with a molecule as described in a) or b) above.

20 Unless the context indicates otherwise, such nucleic acid molecules, which are included within the term "Fve nucleic acid molecule" may have one or more of the following characteristics:

- 1) They may be DNA or RNA (including variants of naturally occurring DNA or RNA structures, which have non-naturally occurring bases and/or non-naturally occurring backbones).

- 2) They may be single-stranded or double-stranded (or in some cases higher stranded, e.g. triple- stranded).
- 3) They may be provided in recombinant form i.e. covalently linked to a heterologous 5' and/or 3' flanking sequence to provide a chimeric molecule (e.g. a vector)
- 5 that does not occur in nature.
- 4) They may be provided with or without 5' and/or 3' flanking sequences that normally occur in nature.
- 5) They may be provided in substantially pure form, e.g. by using probes to isolate cloned molecules having a desired target sequence or by using chemical synthesis
- 10 techniques. Thus they may be provided in a form that is substantially free from contaminating proteins and/or from other nucleic acids.
- 6) They may be provided with introns (e.g. as a full-length gene) or without introns (e.g. as DNA).
- 7) They may be provided in linear or non-linear (e.g. circular) form.
- 15 These five molecules include not only molecules with classical DNA or RNA structures, but also variants with modified (non-phosphodiester) backbones - e.g. morpholino derivatives and peptide nucleic acids (PNAs), which contain an N-(2-aminoethyl)glycine-based pseudopeptide backbone. (See Nielsen, P.E., Annual Review of Biophysics & Biomolecular Structure, 24:167-83 (1995)). Nucleic acid variants
- 20 with modified backbones can have increased stability relative to unmodified nucleic acids and are particularly useful where hybridisation is desired over a relatively long period (e.g. in antisense therapy).

Nucleic acid molecules and uses thereof are discussed in further detail below:

a) Coding nucleic acid molecules

The Fve polypeptides can be coded for by a large variety of nucleic acid molecules, taking into account the well-known degeneracy of the genetic code. All of these coding nucleic acid molecules are within the scope of the present document.

5 The Fve nucleic acids may be administered to an individual and used to express polypeptides disclosed here. Thus, they may be used for the same treatments as the Fve polypeptides.

10 The Fve nucleic acid molecules may be provided in the form of vectors, although this is not essential. Preferred vectors for use in treatment include replication-deficient adenoviruses, retroviruses and adeno-associated viruses.

15 Fve nucleic acid molecules may be administered to a patient by physical methods. These methods include topical application of the nucleic acid in an appropriate vehicle, for example in solution in a pharmaceutically acceptable excipient, such as phosphate buffered saline (PBS). They also include particle bombardment (which is sometimes known as "gene gun" technology and is described in US Patent No. 5371015). Here inert particles, such as gold beads coated with a nucleic acid, can be accelerated at speeds sufficient to enable them to penetrate cells. They can be used for example to penetrate the skin of a patient and may be administered by means of discharge under high pressure from a projecting device. Other physical methods of administering the Fve nucleic acid directly to 20 a recipient include ultrasound, electrical stimulation (including iontophoresis) and microseeding (see e.g. US Patent No. 5697901). Alternatively, the Fve nucleic acid molecules may simply be injected at appropriate site (e.g. muscle). They may be incorporated in or on a carrier (which may be a lipid-based carrier, such as a liposome).

25 Fve nucleic acid molecules may be introduced into host cells (optionally in the form of vectors) to enable the expression of polypeptides. Alternatively, cell-free expression systems may be used. By using an appropriate expression system the Fve polypeptides can be produced in a desired form. For example, the Fve polypeptides can be

produced by micro-organisms such as bacteria or yeast, by cultured insect cells (which may be baculovirus-infected), by mammalian cells (such as CHO cells) or by transgenic animals that, for instance, secrete the Fve proteins in milk (see e.g. international patent application WO88/00239). Where glycosylation is desired, eukaryotic (e.g. mammalian or 5 insect) expression systems are preferred.

Whatever means is used to obtain expression, transcriptional and translational control sequences will normally be present and will be operatively linked to a sequence encoding a polypeptide to be expressed. These control sequences may be heterologous to the sequence encoding the Fve polypeptide or may be found associated with it *in vivo*.

10 Promoter, operator and /or enhancer sequences may, for example, be provided, as may polyadenylation sites, splice sites, stop and start codons, upstream and downstream regulatory regions, etc. If desired, a constitutive promoter may be provided. Alternatively, a regulatable promoter may be provided to enable transcription to be controlled by administration of a regulator. The promoter (if present) may be tissue-specific or non 15 tissue-specific.

Polypeptides comprising N-terminal methionine may be produced using certain expression systems, whilst in others the mature polypeptide may lack this residue. Fve polypeptides may initially be expressed so as to include signal sequences. Different signal sequences may be provided for different expression systems. Alternatively, signal 20 sequences may be absent, if not needed.

Once expressed, Fve polypeptides may be purified by a wide variety of techniques. Purification techniques may be used under reducing conditions (in order prevent disulphide bond formation) or non-reducing conditions. Available purification techniques include, for example, electrophoretic techniques, such as SDS PAGE (see e.g. Hunkapiller 25 *et al, Methods Enzymol.* 91:227 (1983), which discloses "Isolation of microgram quantities of proteins from polyacrylamide gels for amino acid sequence analysis."); affinity techniques (e.g. immunoaffinity chromatography); HPLC; gel filtration; ion-exchange

chromatography; isoelectric focussing; etc. If desired, combinations of different purification steps may be used and/or individual purification steps may be repeated.

In summary, techniques for cloning, expressing and purifying polypeptides are well known to the skilled person. Various such techniques are disclosed in standard text-books, 5 such as in Sambrook *et al* [*Molecular Cloning* 2nd Edition, Cold Spring Harbor Laboratory Press (1989)]; in Old & Primrose [*Principles of Gene Manipulation* 5th Edition, Blackwell Scientific Publications (1994)]; and in Stryer [*Biochemistry* 4th Edition, W H Freeman and Company (1995)].

b) *Complementary nucleic acid molecules*

We also describe nucleic acid strands complementary thereto, whether or not the 10 coding and complementary strands are associated in a duplex. Thus, for example, mRNA and cDNA molecules are included.

c) *Hybridising nucleic acid molecules*

Nucleic acid molecules that can hybridise to one or more of the F've nucleic acid 15 molecules discussed above are also disclosed. Such nucleic acid molecules are referred to herein as "hybridising" nucleic acid molecules. Desirably hybridising molecules are at least 10 nucleotides in length and preferably are at least 20, at least 50, at least 100, or at least 200 nucleotides in length.

A hybridising nucleic acid molecule may have a high degree of sequence identity 20 along its length with a nucleic acid molecule within the scope of b) or a) above (e.g. at least 50%, at least 75% or at least 90% sequence identity), although this is not essential. The greater the degree of sequence identity that a given single stranded nucleic acid molecule has with a strand of a nucleic acid molecule, the greater the likelihood that it will hybridise to the complement of said strand.

Most preferably, hybridising nucleic acid molecules hybridise to either DNA strand of a Fve nucleic acid, for example a sequence shown in Appendix A, or to an RNA equivalent thereof, or to a strand that is complementary to either of the aforesaid strands.

Hybridising nucleic acid molecules can be useful as probes or primers, for
5 example.

Probes can be used to purify and/or to identify Fve nucleic acids. They may be used in diagnosis. For example, probes may be used to determine whether or not an organism such as a fungus has a wild-type gene encoding a Fve polypeptide described here, or whether or not one or more deletions, insertions and/or replacements of bases relative to 10 the wild-type sequence are present. It may therefore be used to identify organisms that do not express Fve polypeptides or that express Fve polypeptides having reduced activity (including inactive polypeptides).

Primers are useful in synthesising nucleic acids or parts thereof based upon a template to which a probe hybridises. They can be used in techniques such as PCR to 15 provide large numbers of nucleic acid molecules.

Hybridising molecules also include antisense strands. These hybridise with "sense" strands so as to inhibit transcription and /or translation. An antisense strand can be synthesised based upon knowledge of a sense strand and base pairing rules. It may be exactly complementary with a sense strand, although it should be noted that exact 20 complementarity is not always essential. It may also be produced by genetic engineering, whereby a part of a DNA molecule is provided in an antisense orientation relative to a promoter and is then used to transcribe RNA molecules. Large numbers of antisense molecules can be provided (e.g. by cloning, by transcription, by PCR, by reverse PCR, etc.

Hybridising molecules include ribozymes. Ribozymes can also be used to regulate 25 expression by binding to and cleaving RNA molecules that include particular target sequences recognised by the ribozymes. Ribozymes can be regarded as special types of

antisense molecule. They are discussed, for example, by Haselhoff and Gerlach (Nature (1988) 334:585 – 91).

Antisense molecules may be DNA or RNA molecules. They may be used in antisense therapy to prevent or reduce undesired expression or activity. Antisense 5 molecules may be administered directly to a patient (e.g. by injection). Alternatively, they may be synthesised *in situ* via a vector that has been administered to a patient.

In addition to the uses described above, the Fve nucleic acid molecules disclosed here (of whatever nature) may be used in screening. Screening can be done to identify 10 moieties that bind to said nucleic acid molecules (e.g. to identify hybridising molecules). It can also be done to identify moieties that affect transcription or translation from said nucleic acid molecules.

It can be used to analyse expression, including analysing expression levels or expression patterns (e.g. by analysing mRNA or cDNA), etc. It can be used to identify 15 particular nucleic acid molecules in a sample. This is useful for identifying biological material from a given source (e.g. from a human or non-human animal). For example, a reference nucleic acid molecule (or part of it) can be digested with restriction enzymes and the resultant nucleic acid fragments can be run on a gel. This can provide a restriction fragment pattern or “fingerprint” that can be compared with a sample. If the comparison provides a match that is unlikely to have occurred by chance, a conclusion can be reached 20 that the sample and the reference molecule are likely to have originated from a common source. By performing statistical analysis a specific degree of confidence that such a conclusion is correct can be provided.

We also describe a library having a Fve nucleic acid molecule described here, as well as an array comprising such an Fve nucleic acid molecule (which may be a library). 25 Preferably the array is a regular array. The array may have a predetermined pattern. It may have a grid-like pattern. The discussion provided herein in respect of libraries and arrays

comprising a polypeptide described here applies *mutatis mutandis* to libraries and arrays comprising the corresponding nucleic acid molecule.

One or more Fve nucleic acid molecules may be immobilised upon a surface (e.g. the surface of a bead or a chip). The surface may, for example, be silicon surface, glass, 5 quartz, a membrane, etc. Techniques for immobilising nucleic acid molecules upon a surface are known and are disclosed, for example, in EP-A-0487104, WO96/04404, WO90/02205, WO96/12014, WO98/44151. In some cases they may include a step of nucleic acid amplification, which may involve PCR. Immobilisation is not however essential. For example nucleic acids may be provided in wells or other containment means 10 (e.g. in a fluid environment).

The Fve nucleic acids may be used in various ways. For example, sequence information can be used in predicting structure and/or function, in homology or identity studies, etc.

VECTORS

As indicated above the nucleic acid molecules described here may be provided in 15 the form of vectors.

Vectors comprising such nucleic acid include plasmids, phasmids, cosmids, viruses (including bacteriophages), YACs, PACs, etc. They will usually include an origin of 20 replication and may include one or more selectable markers e.g. drug resistance markers and/or markers enabling growth on a particular medium. A vector may include a marker that is inactivated when a nucleic acid molecule, such as the ones described here, is inserted into the vector. Here a further marker may be provided that is different from the marker that is inactivated (e.g. it encodes a different type of drug resistance).

Vectors may include one or more regions necessary for transcription of RNA 25 encoding a polypeptide. Such vectors are often referred to as expression vectors. They will

usually contain a promoter and may contain additional regulatory regions – e.g. operator sequences, enhancer sequences, etc. Translation can be provided by a host cell or by a cell free expression system.

- Vectors need not be used for expression. They may be provided for maintaining a
5 given nucleic acid sequence, for replicating that sequence, for manipulating, it or for transferring it between different locations (e.g. between different organisms).

Large nucleic acid molecules may be incorporated into high capacity vectors (e.g. cosmids, phasmids, YACs or PACs). Smaller nucleic acid molecules may be incorporated into a wide variety of vectors.

- 10 Fve polynucleotides, for example those described here, can be incorporated into a recombinant replicable vector. The vector may be used to replicate the nucleic acid in a compatible host cell. Thus in a further embodiment, we provide a method of making polynucleotides by introducing a polynucleotide into a replicable vector, introducing the vector into a compatible host cell, and growing the host cell under conditions which bring about replication of the vector. The vector may be recovered from the host cell. Suitable host cells include bacteria such as *E. coli*, yeast, mammalian cell lines and other eukaryotic cell lines, for example insect Sf9 cells.
15

- 20 Preferably, a polynucleotide in a vector is operably linked to a control sequence that is capable of providing for the expression of the coding sequence by the host cell, i.e. the vector is an expression vector. The term “operably linked” means that the components described are in a relationship permitting them to function in their intended manner. A regulatory sequence “operably linked” to a coding sequence is ligated in such a way that expression of the coding sequence is achieved under condition compatible with the control sequences.

The control sequences may be modified, for example by the addition of further transcriptional regulatory elements to make the level of transcription directed by the control sequences more responsive to transcriptional modulators.

- Vectors may be transformed or transfected into a suitable host cell as described
- 5 below to provide for expression of a protein. This process may comprise culturing a host cell transformed with an expression vector as described above under conditions to provide for expression by the vector of a coding sequence encoding the protein, and optionally recovering the expressed protein. Vectors will be chosen that are compatible with the host cell used.
- 10 The vectors may be for example, plasmid or virus vectors provided with an origin of replication, optionally a promoter for the expression of the said polynucleotide and optionally a regulator of the promoter. The vectors may contain one or more selectable marker genes, for example an ampicillin resistance gene in the case of a bacterial plasmid or a neomycin resistance gene for a mammalian vector. Vectors may be used, for example,
- 15 to transfect or transform a host cell.

Control sequences operably linked to sequences encoding the polypeptide include promoters/enhancers and other expression regulation signals. These control sequences may be selected to be compatible with the host cell for which the expression vector is designed to be used in. The term promoter is well-known in the art and encompasses nucleic acid regions ranging in size and complexity from minimal promoters to promoters including upstream elements and enhancers.

The promoter is typically selected from promoters which are functional in mammalian cells, although prokaryotic promoters and promoters functional in other eukaryotic cells, such as insect cells, may be used. The promoter is typically derived from 25 promoter sequences of viral or eukaryotic genes. For example, it may be a promoter derived from the genome of a cell in which expression is to occur. With respect to eukaryotic promoters, they may be promoters that function in a ubiquitous manner (such as

promoters of α -actin, β -actin, tubulin) or, alternatively, a tissue-specific manner (such as promoters of the genes for pyruvate kinase). They may also be promoters that respond to specific stimuli, for example promoters that bind steroid hormone receptors. Viral promoters may also be used, for example the Moloney murine leukaemia virus long terminal repeat (MMLV LTR) promoter, the rous sarcoma virus (RSV) LTR promoter or the human cytomegalovirus (CMV) IE promoter.

It may also be advantageous for the promoters to be inducible so that the levels of expression of the heterologous gene can be regulated during the life-time of the cell. Inducible means that the levels of expression obtained using the promoter can be regulated.

In addition, any of these promoters may be modified by the addition of further regulatory sequences, for example enhancer sequences. Chimeric promoters may also be used comprising sequence elements from two or more different promoters described above.

Polynucleotides may also be inserted into the vectors described above in an antisense orientation to provide for the production of antisense RNA. Antisense RNA or other antisense polynucleotides may also be produced by synthetic means. Such antisense polynucleotides may be used in a method of controlling the levels of RNAs transcribed from genes comprising any one of the polynucleotides described here.

20 HOST CELLS

Vectors and polynucleotides or nucleic acids comprising or encoding Fve nucleic acids, fragments, homologues, variants or derivatives thereof may be introduced into host cells for the purpose of replicating the vectors/polynucleotides and/or expressing the polypeptides encoded by the polynucleotides. Although the polypeptides may be produced using prokaryotic cells as host cells, it is preferred to use eukaryotic cells, for example yeast, insect or mammalian cells, in particular mammalian cells.

Vectors/polynucleotides may be introduced into suitable host cells using a variety of techniques known in the art, such as transfection, transformation and electroporation. Where vectors/polynucleotides are to be administered to animals, several techniques are known in the art, for example infection with recombinant viral vectors such as 5 retroviruses, herpes simplex viruses and adenoviruses, direct injection of nucleic acids and biolistic transformation.

We therefore further disclose cells comprising F've nucleic acid molecules or vectors. These may for example be used for expression, as described herein.

A cell capable of expressing a F've polypeptide described here can be cultured and 10 used to provide the F've polypeptide, which can then be purified.

Alternatively, the cell may be used in therapy for the same purposes as the F've polypeptide. For example, cells may be provided from a patient (e.g. via a biopsy), transfected with a nucleic acid molecule or vector and, if desired, cultured *in vitro*, prior to being returned to the patient (e.g. by injection). The cells can then produce the F've 15 polypeptide *in vivo*. Preferably the cells comprise a regulatable promoter enabling transcription to be controlled via administration of one or more regulator molecules. If desired, the promoter may be tissue specific.

Expression is not however essential since the cells may be provided simply for maintaining a given nucleic acid sequence, for replicating the sequence, for manipulating 20 it, etc.

Such cells may be provided in any appropriate form. For example, they may be provided in isolated form, in culture, in stored form, etc. Storage may, for example, involve cryopreservation, buffering, sterile conditions, etc. Such cells may be provided by gene cloning techniques, by stem cell technology or by any other means. They may be part 25 of a tissue or an organ, which may itself be provided in any of the forms discussed above. The cell, tissue or organ may be stored and used later for implantation, if desired.

Techniques for providing tissues or organs, include stem cell technology, the provision of cells tissues or organs from transgenic animals, retroviral and non-retroviral techniques for introducing nucleic acids, etc.

In some case cells may be provided together with other material to aid the structure
5 or function or of an implant. For example scaffolds may be provided to hold cells in position, to provide mechanical strength, etc. These may be in the form of matrixes of biodegradable or non-biodegradable material. WO95/01810 describes various materials that can be used for this purpose.

ANIMALS

10 We also disclose transgenic animals, preferably non-human transgenic animals. Such animals may be useful for producing the particular Fve polypeptides described here (e.g. via secretion in milk, as described herein). Alternatively, they may be useful as test animals for analysing the effect(s) of such Fve polypeptides.

15 Techniques for producing transgenic animals are well known and are described e.g. in US patents 4870009 and 4873191. For example, a nucleic acid encoding a Fve polypeptide of interest may be microinjected into a pronucleus of a fertilised oocyte. The oocyte may then be allowed to develop in a pseudopregnant female foster animal. The animal resulting from development of the oocyte can be tested (e.g. with antibodies) to determine whether or not it expresses the particular polypeptide. Alternatively, it can be 20 tested with a probe to determine if it has a transgene (even if there is no expression).

A transgenic animal can be used as a founder animal, which may be bred from in order to produce further transgenic animals. Two transgenic animals may be crossed. For example, in some cases transgenic animals may be haploid for a given gene and it may be desired to try to provide a diploid offspring via crossing.

A transgenic animal may be cloned, e.g. by using the procedures set out in WO97/07668 and WO97/07699 (see also Nature 385:810-813 (1997)). Thus a quiescent cell can be provided and combined with an oocyte from which the nucleus has been removed combined. This can be achieved using electrical discharges. The resultant cell can
5 be allowed to develop in culture and can then be transferred to a pseudopregnant female.

ANALYTICAL TOOLS AND SYSTEMS

We disclose a moiety comprising a Fve polypeptide, a Fve nucleic acid, a vector comprising Fve, a cell expressing Fve, an Fve binding agent, a moiety identified/identifiable by a screen as described here, when used as an analytical tool or
10 when present in a system suitable for analysis, especially high throughput analysis.

Such an analytical tool or system is useful for a plethora of different purposes. These include diagnosis, forensic science, screening, the identification or characterisation of individuals or populations, preventative medicine, etc.

Libraries comprising such a Fve moiety may be used for the above purposes. A
15 library will generally comprise a plurality of heterologous moieties. Preferred libraries comprise at least 100, at least 10,000, at least 1,000,000, or at least 1,000,000,000 heterologous moieties. Desirably a moiety is provided at a predetermined position within a library. In some cases a plurality of moieties may be present within a library at predetermined positions. A predetermined position may be assigned spatial co-ordinates.
20 These may be stored or processed in a computer in order to assist in analysis.

We further disclose an array comprising such a Fve moiety (whether or not the array is also a library). Preferably the array is a regular array. The array may have a predetermined pattern. It may have a grid-like pattern. Preferred arrays comprise at least 100, at least 10,000, at least 1,000,000, or at least 1,000,000,000 components.

A library or array may include naturally occurring moieties, non-naturally occurring moieties, or a mixture of naturally occurring and non-naturally occurring moieties. The moieties may be provided in solution, on beads, on chips (see e.g. Fodor (1993) Nature 364:555-556), on bacteria (see e.g. US Patent 5223409), on spores (see e.g. US 5 Patent 5223409), on 'phage (see e.g. Scott and Smith (1990) Science 249:386-90 and US Patent 5223409), etc.

Such F^{ve} moieties may be immobilised upon a surface, if desired. For example, one or more nucleic acid molecules may be immobilised upon a surface (e.g. the surface of a bead or a chip). The surface may, for example, be silicon, glass, quartz, a membrane, etc. 10 Techniques for immobilising nucleic acid molecules upon a surface are known and are disclosed, for example, in EP-A-0487104, WO96/04404, WO90/02205, WO96/12014, WO98/44151. In some cases they may include a step of nucleic acid amplification, and may involve PCR.

Immobilisation is not however essential, even if moieties are to be used in high 15 throughput analysis. For example, they may be provided in wells, channels, grooves or other containment means.

Whether or not present in a library, an array or in immobilised or non-immobilised form, it is often desirable to locate the position of one or more moieties being analysed or being used in analysis. This can be done by assigning it spatial co-ordinates, which may be 20 provided, stored or processed or provided by a computer. In some cases the location may be determined by a sensor (e.g. a CCD device), which may be operatively linked with a computer.

DNA VACCINES

Any of the F^{ve} nucleic acids disclosed here may be administered to an individual 25 in the form of a DNA vaccine. DNA vaccines are known in the art, and are described in detail in, for example, WO03012117, WO03007986, etc.

The Fve may be administered to an individual in the form of a DNA vaccine. A DNA encoding the Fve, for example, a Fve nucleic acid as disclosed here, may be in any form, for example in the form of a cloned plasmid DNA or a synthetic oligonucleotide. The DNA may be delivered together with a cytokine, for example, IL-2, and / or other co-stimulatory molecules. The cytokines and / or co-stimulatory molecules may themselves be delivered in the form of plasmid or oligonucleotide DNA.

The response to a DNA vaccine has been shown to be increased by the presence of immunostimulatory DNA sequences (ISS). These can take the form of hexameric motifs containing methylated CpG, according to the formula: 5' purine-purine-CG-pyrimidine-pyrimidine-3'. The DNA vaccines may incorporate these or other ISSs, in the DNA encoding the Fve, in the DNA encoding the cytokine or other co-stimulatory molecules, or in both. A review of the advantages of DNA vaccination is provided by Tighe et al (1998, Immunology Today, 19(2), 89-97).

ANTIBODIES

We also provide monoclonal or polyclonal antibodies to polypeptides or fragments thereof. Thus, we further provide a process for the production of monoclonal or polyclonal antibodies to an Fve polypeptide, fragment, homologue, variant or derivative thereof

If polyclonal antibodies are desired, a selected mammal (e.g., mouse, rabbit, goat, horse, etc.) is immunised with an immunogenic polypeptide bearing an epitope(s) from a polypeptide. Serum from the immunised animal is collected and treated according to known procedures. If serum containing polyclonal antibodies to an epitope from a polypeptide contains antibodies to other antigens, the polyclonal antibodies can be purified by immunoaffinity chromatography. Techniques for producing and processing polyclonal antisera are known in the art. In order that such antibodies may be made, we also provide polypeptides or fragments thereof haptenised to another polypeptide for use as immunogens in animals or humans.

Monoclonal antibodies directed against epitopes in the polypeptides can also be readily produced by one skilled in the art. The general methodology for making monoclonal antibodies by hybridomas is well known. Immortal antibody-producing cell lines can be created by cell fusion, and also by other techniques such as direct 5 transformation of B lymphocytes with oncogenic DNA, or transfection with Epstein-Barr virus. Panels of monoclonal antibodies produced against epitopes in the polypeptides can be screened for various properties; i.e., for isotype and epitope affinity.

An alternative technique involves screening phage display libraries where, for example the phage express scFv fragments on the surface of their coat with a large variety 10 of complementarity determining regions (CDRs). This technique is well known in the art.

Antibodies, both monoclonal and polyclonal, which are directed against epitopes from polypeptides are particularly useful in diagnosis, and those which are neutralising are useful in passive immunotherapy. Monoclonal antibodies, in particular, may be used to raise anti-idiotype antibodies. Anti-idiotype antibodies are immunoglobulins which carry 15 an "internal image" of the antigen of the agent against which protection is desired.

Techniques for raising anti-idiotype antibodies are known in the art. These anti-idiotype antibodies may also be useful in therapy.

For the purposes of this document, the term "antibody", unless specified to the contrary, includes fragments of whole antibodies which retain their binding activity for a 20 target antigen. Such fragments include Fv, F(ab') and F(ab')₂ fragments, as well as single chain antibodies (scFv). Furthermore, the antibodies and fragments thereof may be humanised antibodies, for example as described in EP-A-239400.

Antibodies may be used in method of detecting polypeptides present in biological samples by a method which comprises: (a) providing an antibody; (b) incubating a 25 biological sample with said antibody under conditions which allow for the formation of an

antibody-antigen complex; and (c) determining whether antibody-antigen complex comprising said antibody is formed.

Suitable samples include extracts tissues such as brain, breast, ovary, lung, colon, pancreas, testes, liver, muscle and bone tissues or from neoplastic growths derived from
5 such tissues.

Antibodies may be bound to a solid support and/or packaged into kits in a suitable container along with suitable reagents, controls, instructions and the like.

ASSAYS

We disclose assays that are suitable for identifying substances which bind to Fve
10 polypeptides, or fragments, homologues, variants or derivatives thereof

In general, such binding assays involve exposing a Fve polypeptide, nucleic acid, or a fragment, homologue, variant or derivative thereof to a candidate molecule and detecting an interaction or binding between the Fve polypeptide, nucleic acid, or a fragment, homologue, variant or derivative thereof and the candidate molecule. The
15 binding assay may be conducted *in vitro*, or *in vivo*.

We disclose assays for identifying substances which are capable of potentiating the activities of Fve polypeptide. Activities of Fve have been described in detail above. Such compounds may be employed as agonists of Fve polypeptide, and may for example be co-administered to an individual to enhance any desired effect.

20 In general, an assay to identify such substances or compounds involves providing a cell or organism, exposing the cell or organism to a Fve polypeptide, nucleic acid, or a fragment, homologue, variant or derivative thereof, exposing the cell to a candidate molecule, and detecting an effect associated with Fve. Any Fve polypeptide mediated

effect or function, as disclosed in this document, particularly the Examples, may be detected.

- In particular, the Fve polypeptide mediated effect is preferably chosen from the group consisting of: up-regulation of expression of Th1 cytokines, preferably IFN- γ and
- 5 TNF- α , down-regulation of expression of Th2 cytokines, preferably IL-4 and IL-13, hemagglutination activity, cell aggregation activity, lymphocyte aggregation activity, lymphoproliferation activity, up-regulation of expression of IL-2, IFN- γ , TNF- α , but not IL-4 in CD3 $^{+}$ T cells, interaction with T and NK cells, adjuvant activity, stimulation of
- 10 CD3 $^{+}$ CD16 $^{+}$ CD56 $^{+}$ natural killer (NK) T cells, up-regulation of expression of allergen specific IgG2a antibody, prevention of systemic anaphylactic reactions and/or decreased footpad edema, preferably as assayed using the Arthus reaction (Ko et al, 1995).

In order to identify agonists, an additive or preferably synergistic effect is detected. Thus, while Fve polypeptide on its own is, for example, capable of reducing a level or number, or down-regulation of expression of a molecule, the assays identify molecules

15 which further reduce the level, number or further down-regulate the expression of a molecule. Thus, preferably, the candidate molecule in conjunction with the Fve polypeptide, nucleic acid, or a fragment, homologue, variant or derivative thereof, down-regulates the expression of, or reduces the level or number, by more than 10%, more than 20%, more than 30%, more than 40%, more than 50%, more than 60%, more than 70%,

20 more than 80%, more than 90%, or more compared to an Fve polypeptide on its own. Thus, for example, a candidate molecule suitable for use as an agonist is one which is capable of enhancing by 10% more the up-regulation of expression of Th1 cytokines, preferably IFN- γ and TNF- α , achieved by Fve polypeptide on its own.

Conversely, assays to identify antagonists involve the detection of a reduction in

25 Fve polypeptide mediated effect. Preferably, the down-regulation of expression or reduction in number or level achieved by Fve polypeptide is reduced in the presence of a suitable candidate molecule. Preferably, the reduction is at least 10%, preferably at least 20%, preferably at least 30%, preferably at least 40%, preferably at least 50%, preferably

at least 60%, preferably at least 70%, preferably at least 80%, preferably at least 90%, or more compared to an Fve polypeptide on its own. Thus, for example, a candidate molecule suitable for use as an antagonist is one which is capable of reducing by 10% more the up-regulation of expression of Th1 cytokines, preferably IFN- γ and TNF- α , achieved by Fve 5 polypeptide on its own.

As an illustration, if N1 is the expression of Th1 cytokines, in an untreated organism or cell, and N2 the expression in an organism or cell exposed to Fve polypeptide, nucleic acid, or a fragment, homologue, variant or derivative thereof, the expression of Th1 cytokines is increased by $R = (N2-N1)/N1 \times 100\%$. Agonists increase R, by a factor x, 10 where x is greater than 1 (e.g., x = 1, 1.1, 1.2, 1.3, 1.4, 1.5, 2, 3, 4, 5, 10, 20, 50, 100 etc); while antagonists decrease R, by a factor x, where x is less than 1 (e.g., x = 0.9, 0.9, 0.7, 0.6, 0.5, 0.4, 0.3, 0.2, 0.1 etc).

For example, an organism may be exposed to a Fve polypeptide, nucleic acid, or a fragment, homologue, variant or derivative thereof and a candidate molecule, and any of 15 the biological activities as set out above, or any combination, detected. Preferred candidate molecules are those which provide an additive or synergistic effect in combination with Fve.

Also disclosed are assays to identify antagonists of Fve polypeptide. Such assays involve detecting a reduced effect on exposure of a cell or organism to an Fve polypeptide, 20 nucleic acid, or a fragment, homologue, variant or derivative thereof in conjunction with a candidate molecule.

In a preferred embodiment, the assays are conducted on whole organisms rather than cells. Preferably, the organism is one which suffers from a disease as disclosed in this document, or exhibits one or more symptoms of such a disease.

CANDIDATE MOLECULES

Suitable candidate molecules for use in the above assays include peptides, especially of from about 5 to 30 or 10 to 25 amino acids in size. Peptides from panels of peptides comprising random sequences or sequences which have been varied consistently 5 to provide a maximally diverse panel of peptides may be used.

Suitable candidate molecules also include antibody products (for example, monoclonal and polyclonal antibodies, single chain antibodies, chimeric antibodies and CDR-grafted antibodies). Furthermore, combinatorial libraries, peptide and peptide mimetics, defined chemical entities, oligonucleotides, and natural product libraries may be 10 screened for activity. The candidate molecules may be used in an initial screen in batches of, for example 10 types of molecules per reaction, and the molecules of those batches which show enhancement or reduction of a Fve polypeptide mediated effect tested individually.

LIBRARIES

- 15 Libraries of candidate molecules, such as libraries of polypeptides or nucleic acids, may be employed in the methods and compositions described here. Such libraries are exposed a cell or organism in the presence of a Fve polypeptide, nucleic acid, or a fragment, homologue, variant or derivative thereof, and an Fve polypeptide mediated effect detected and assayed as described above.
- 20 Selection protocols for isolating desired members of large libraries are known in the art, as typified by phage display techniques. Such systems, in which diverse peptide sequences are displayed on the surface of filamentous bacteriophage (Scott and Smith (1990 *supra*), have proven useful for creating libraries of antibody fragments (and the nucleotide sequences that encoding them) for the *in vitro* selection and amplification of 25 specific antibody fragments that bind a target antigen. The nucleotide sequences encoding the V_H and V_L regions are linked to gene fragments which encode leader signals that direct

them to the periplasmic space of *E. coli* and as a result the resultant antibody fragments are displayed on the surface of the bacteriophage, typically as fusions to bacteriophage coat proteins (e.g., pIII or pVIII). Alternatively, antibody fragments are displayed externally on lambda phage capsids (phagebodies). An advantage of phage-based display systems is that, because they are biological systems, selected library members can be amplified simply by growing the phage containing the selected library member in bacterial cells. Furthermore, since the nucleotide sequence that encodes the polypeptide library member is contained on a phage or phagemid vector, sequencing, expression and subsequent genetic manipulation is relatively straightforward.

Methods for the construction of bacteriophage antibody display libraries and lambda phage expression libraries are well known in the art (McCafferty *et al.* (1990) *supra*; Kang *et al.* (1991) *Proc. Natl. Acad. Sci. U.S.A.*, 88: 4363; Clackson *et al.* (1991) *Nature*, 352: 624; Lowman *et al.* (1991) *Biochemistry*, 30: 10832; Burton *et al.* (1991) *Proc. Natl. Acad. Sci U.S.A.*, 88: 10134; Hoogenboom *et al.* (1991) *Nucleic Acids Res.*, 19: 4133; Chang *et al.* (1991) *J. Immunol.*, 147: 3610; Breitling *et al.* (1991) *Gene*, 104: 147; Marks *et al.* (1991) *supra*; Barbas *et al.* (1992) *supra*; Hawkins and Winter (1992) *J. Immunol.*, 22: 867; Marks *et al.*, 1992, *J. Biol. Chem.*, 267: 16007; Lerner *et al.* (1992) *Science*, 258: 1313, incorporated herein by reference). Such techniques may be modified if necessary for the expression generally of polypeptide libraries.

One particularly advantageous approach has been the use of scFv phage-libraries (Bird, R.E., *et al.* (1988) *Science* 242: 423-6, Huston *et al.*, 1988, *Proc. Natl. Acad. Sci U.S.A.*, 85: 5879-5883; Chaudhary *et al.* (1990) *Proc. Natl. Acad. Sci U.S.A.*, 87: 1066-1070; McCafferty *et al.* (1990) *supra*; Clackson *et al.* (1991) *supra*; Marks *et al.* (1991) *supra*; Chiswell *et al.* (1992) *Trends Biotech.*, 10: 80; Marks *et al.* (1992) *supra*). Various embodiments of scFv libraries displayed on bacteriophage coat proteins have been described. Refinements of phage display approaches are also known, for example as described in WO96/06213 and WO92/01047 (Medical Research Council *et al.*) and WO97/08320 (Morphosys, *supra*), which are incorporated herein by reference.

Alternative library selection technologies include bacteriophage lambda expression systems, which may be screened directly as bacteriophage plaques or as colonies of lysogens, both as previously described (Huse *et al.* (1989) *Science*, 246: 1275; Caton and Koprowski (1990) *Proc. Natl. Acad. Sci. U.S.A.*, 87; Mullinax *et al.* (1990) *Proc. Natl. Acad. Sci. U.S.A.*, 87: 8095; Persson *et al.* (1991) *Proc. Natl. Acad. Sci. U.S.A.*, 88: 2432) and are of use. These expression systems may be used to screen a large number of different members of a library, in the order of about 10^6 or even more. Other screening systems rely, for example, on direct chemical synthesis of library members. One early method involves the synthesis of peptides on a set of pins or rods, such as described in WO84/03564. A similar method involving peptide synthesis on beads, which forms a peptide library in which each bead is an individual library member, is described in U.S. Patent No. 4,631,211 and a related method is described in WO92/00091. A significant improvement of the bead-based methods involves tagging each bead with a unique identifier tag, such as an oligonucleotide, so as to facilitate identification of the amino acid sequence of each library member. These improved bead-based methods are described in WO93/06121.

Another chemical synthesis method involves the synthesis of arrays of peptides (or peptidomimetics) on a surface in a manner that places each distinct library member (e.g., unique peptide sequence) at a discrete, predefined location in the array. The identity of each library member is determined by its spatial location in the array. The locations in the array where binding interactions between a predetermined molecule (e.g., a receptor) and reactive library members occur is determined, thereby identifying the sequences of the reactive library members on the basis of spatial location. These methods are described in U.S. Patent No. 5,143,854; WO90/15070 and WO92/10092; Fodor *et al.* (1991) *Science*, 251: 767; Dower and Fodor (1991) *Ann. Rep. Med. Chem.*, 26: 271.

Other systems for generating libraries of polypeptides or nucleotides involve the use of cell-free enzymatic machinery for the *in vitro* synthesis of the library members. In one method, RNA molecules are selected by alternate rounds of selection against a target ligand and PCR amplification (Tuerk and Gold (1990) *Science*, 249: 505; Ellington and Szostak (1990) *Nature*, 346: 818). A similar technique may be used to identify DNA

sequences which bind a predetermined human transcription factor (Thiesen and Bach (1990) *Nucleic Acids Res.*, 18: 3203; Beaudry and Joyce (1992) *Science*, 257: 635; WO92/05258 and WO92/14843). In a similar way, *in vitro* translation can be used to synthesise polypeptides as a method for generating large libraries. These methods which 5 generally comprise stabilised polysome complexes, are described further in WO88/08453, WO90/05785, WO90/07003, WO91/02076, WO91/05058, and WO92/02536. Alternative display systems which are not phage-based, such as those disclosed in WO95/22625 and WO95/11922 (Affymax) use the polysomes to display polypeptides for selection. These and all the foregoing documents also are incorporated herein by reference.

10 COMBINATORIAL LIBRARIES

Libraries, in particular, libraries of candidate molecules, may suitably be in the form of combinatorial libraries (also known as combinatorial chemical libraries).

A “combinatorial library”, as the term is used in this document, is a collection of multiple species of chemical compounds that consist of randomly selected subunits. 15 Combinatorial libraries may be screened for molecules which are capable of potentiating, enhancing, reducing or minimising the a Fve polypeptide mediated effect when exposed to a cell or organism.

Various combinatorial libraries of chemical compounds are currently available, including libraries active against proteolytic and non-proteolytic enzymes, libraries of 20 agonists and antagonists of G-protein coupled receptors (GPCRs), libraries active against non-GPCR targets (e.g., integrins, ion channels, domain interactions, nuclear receptors, and transcription factors) and libraries of whole-cell oncology and anti-infective targets, among others. A comprehensive review of combinatorial libraries, in particular their construction and uses is provided in Dolle and Nelson (1999), *Journal of Combinatorial Chemistry*, Vol 1 No 4, 235-282. Reference is also made to *Combinatorial peptide library protocols* (edited by Shmuel Cabilly, Totowa, N.J.: Humana Press, c1998. *Methods in Molecular Biology*; v. 87).

- Further references describing chemical combinatorial libraries, their production and use include those available from the URL <http://www.netsci.org/Science/Combichem/>, including The Chemical Generation of Molecular Diversity. Michael R. Pavia, Sphinx Pharmaceuticals, A Division of Eli Lilly (Published July, 1995); Combinatorial Chemistry: 5 A Strategy for the Future - MDL Information Systems discusses the role its Project Library plays in managing diversity libraries (Published July, 1995); Solid Support Combinatorial Chemistry in Lead Discovery and SAR Optimization, Adnan M. M. Mjalli and Barry E. Toyonaga, Ontogen Corporation (Published July, 1995); Non-Peptidic Bradykinin Receptor Antagonists From a Structurally Directed Non-Peptide Library. Sarvajit 10 Chakravarty, Babu J. Mavunkel, Robin Andy, Donald J. Kyle*, Scios Nova Inc. (Published July, 1995); Combinatorial Chemistry Library Design using Pharmacophore Diversity Keith Davies and Clive Briant, Chemical Design Ltd. (Published July, 1995); A Database System for Combinatorial Synthesis Experiments - Craig James and David Weininger, Daylight Chemical Information Systems, Inc. (Published July, 1995); An 15 Information Management Architecture for Combinatorial Chemistry, Keith Davies and Catherine White, Chemical Design Ltd. (Published July, 1995); Novel Software Tools for Addressing Chemical Diversity, R. S. Pearlman, Laboratory for Molecular Graphics and Theoretical Modeling, College of Pharmacy, University of Texas (Published June/July, 1996); Opportunities for Computational Chemists Afforded by the New Strategies in Drug 20 Discovery: An Opinion, Yvonne Connolly Martin, Computer Assisted Molecular Design Project, Abbott Laboratories (Published June/July, 1996); Combinatorial Chemistry and Molecular Diversity Course at the University of Louisville: A Description, Arno F. Spatola, Department of Chemistry, University of Louisville (Published June/July, 1996); Chemically Generated Screening Libraries: Present and Future. Michael R. Pavia, Sphinx 25 Pharmaceuticals, A Division of Eli Lilly (Published June/July, 1996); Chemical Strategies For Introducing Carbohydrate Molecular Diversity Into The Drug Discovery Process.. Michael J. Sofia, Transcell Technologies Inc. (Published June/July, 1996); Data Management for Combinatorial Chemistry. Maryjo Zaborowski, Chiron Corporation and Sheila H. DeWitt, Parke-Davis Pharmaceutical Research, Division of Warner-Lambert 30 Company (Published November, 1995); and The Impact of High Throughput Organic Synthesis on R&D in Bio-Based Industries, John P. Devlin (Published March, 1996).

Techniques in combinatorial chemistry are gaining wide acceptance among modern methods for the generation of new pharmaceutical leads (Gallop, M. A. et al., 1994, J. Med. Chem. 37:1233-1251; Gordon, E. M. et al., 1994, J. Med. Chem. 37:1385-1401.). One combinatorial approach in use is based on a strategy involving the synthesis of 5 libraries containing a different structure on each particle of the solid phase support, interaction of the library with a soluble receptor, identification of the 'bead' which interacts with the macromolecular target, and determination of the structure carried by the identified 'bead' (Lam, K. S. et al., 1991, Nature 354:82-84). An alternative to this 10 approach is the sequential release of defined aliquots of the compounds from the solid support, with subsequent determination of activity in solution, identification of the particle from which the active compound was released, and elucidation of its structure by direct sequencing (Salmon, S. E. et al., 1993, Proc. Natl. Acad. Sci. USA 90:11708-11712), or by reading its code (Kerr, J. M. et al., 1993, J. Am. Chem. Soc. 115:2529-2531; Nikolaiev, V. et al., 1993, Pept. Res. 6:161-170; Ohlmeyer, M. H. J. et al., 1993, 15 Proc. Natl. Acad. Sci. USA 90:10922-10926).

Soluble random combinatorial libraries may be synthesized using a simple principle for the generation of equimolar mixtures of peptides which was first described by Furka (Furka, A. et al., 1988, Xth International Symposium on Medicinal Chemistry, Budapest 1988; Furka, A. et al., 1988, 14th International Congress of Biochemistry, 20 Prague 1988; Furka, A. et al., 1991, Int. J. Peptide Protein Res. 37:487-493). The construction of soluble libraries for iterative screening has also been described (Houghten, R. A. et al. 1991, Nature 354:84-86). K. S. Lam disclosed the novel and unexpectedly powerful technique of using insoluble random combinatorial libraries. Lam synthesized 25 random combinatorial libraries on solid phase supports, so that each support had a test compound of uniform molecular structure, and screened the libraries without prior removal of the test compounds from the support by solid phase binding protocols (Lam, K. S. et al., 1991, Nature 354:82-84).

Thus, a library of candidate molecules may be a synthetic combinatorial library (e.g., a combinatorial chemical library), a cellular extract, a bodily fluid (e.g., urine, blood,

tears, sweat, or saliva), or other mixture of synthetic or natural products (e.g., a library of small molecules or a fermentation mixture).

- A library of molecules may include, for example, amino acids, oligopeptides, polypeptides, proteins, or fragments of peptides or proteins; nucleic acids (e.g., antisense; 5 DNA; RNA; or peptide nucleic acids, PNA); aptamers; or carbohydrates or polysaccharides. Each member of the library can be singular or can be a part of a mixture (e.g., a compressed library). The library may contain purified compounds or can be "dirty" (i.e., containing a significant quantity of impurities). Commercially available libraries (e.g., from Affymetrix, ArQule, Neose Technologies, Sarco, Ciddco, Oxford Asymmetry, 10 Maybridge, Aldrich, Panlabs, Pharmacopoeia, Sigma, or Tripode) may also be used with the methods described here.

In addition to libraries as described above, special libraries called diversity files can be used to assess the specificity, reliability, or reproducibility of the new methods. Diversity files contain a large number of compounds (e.g., 1000 or more small molecules) 15 representative of many classes of compounds that could potentially result in nonspecific detection in an assay. Diversity files are commercially available or can also be assembled from individual compounds commercially available from the vendors listed above.

CANDIDATE SUBSTANCES

- Suitable candidate substances include peptides, especially of from about 5 to 30 or 20 10 to 25 amino acids in size, based on the sequence of the polypeptides described in the Examples, or variants of such peptides in which one or more residues have been substituted. Peptides from panels of peptides comprising random sequences or sequences which have been varied consistently to provide a maximally diverse panel of peptides may be used.
- 25 Suitable candidate substances also include antibody products (for example, monoclonal and polyclonal antibodies, single chain antibodies, chimeric antibodies and

CDR-grafted antibodies) which are specific for a polypeptide. Furthermore, combinatorial libraries, peptide and peptide mimetics, defined chemical entities, oligonucleotides, and natural product libraries may be screened for activity as inhibitors of binding of a polypeptide to the cell division cycle machinery, for example mitotic/meiotic apparatus (such as microtubules). The candidate substances may be used in an initial screen in batches of, for example 10 substances per reaction, and the substances of those batches which show inhibition tested individually. Candidate substances which show activity in *in vitro* screens such as those described below can then be tested in whole cell systems, such as mammalian cells which will be exposed to the inhibitor and tested for inhibition of any of the stages of the cell cycle.

POLYPEPTIDE BINDING ASSAYS

One type of assay for identifying substances that bind to a polypeptide involves contacting a polypeptide, which is immobilised on a solid support, with a non-immobilised candidate substance determining whether and/or to what extent the polypeptide and candidate substance bind to each other. Alternatively, the candidate substance may be immobilised and the polypeptide non-immobilised. This may be used to detect substances capable of binding to Fve polypeptides, or fragments, homologues, variants or derivatives thereof.

In a preferred assay method, the polypeptide is immobilised on beads such as agarose beads. Typically this is achieved by expressing the Fve polypeptide, or a fragment, homologue, variant or derivative thereof as a GST-fusion protein in bacteria, yeast or higher eukaryotic cell lines and purifying the GST-fusion protein from crude cell extracts using glutathione-agarose beads (Smith and Johnson, 1988). As a control, binding of the candidate substance, which is not a GST-fusion protein, to the immobilised polypeptide is determined in the absence of the polypeptide. The binding of the candidate substance to the immobilised polypeptide is then determined. This type of assay is known in the art as a GST pulldown assay. Again, the candidate substance may be immobilised and the polypeptide non-immobilised.

It is also possible to perform this type of assay using different affinity purification systems for immobilising one of the components, for example Ni-NTA agarose and histidine-tagged components.

- Binding of the Fve polypeptide, or a fragment, homologue, variant or derivative thereof to the candidate substance may be determined by a variety of methods well-known in the art. For example, the non-immobilised component may be labeled (with for example, a radioactive label, an epitope tag or an enzyme-antibody conjugate). Alternatively, binding may be determined by immunological detection techniques. For example, the reaction mixture can be Western blotted and the blot probed with an antibody that detects the non-immobilised component. ELISA techniques may also be used.

Candidate substances are typically added to a final concentration of from 1 to 1000 nmol/ml, more preferably from 1 to 100 nmol/ml. In the case of antibodies, the final concentration used is typically from 100 to 500 µg/ml, more preferably from 200 to 300 µg/ml.

15 FVE DISEASES

As disclosed elsewhere in this document, Fve polypeptides, nucleic acids, and fragments, homologues, variants and derivatives thereof, host cells, vectors, DNA vaccines, etc, are suitable for treating or preventing various diseases (here referred to as "Fve diseases"). They may be administered in an amount in the range of 1 microgram to 20 1 gramme to an average human patient or individual to be vaccinated. It is preferred to use a smaller dose in the range of 1 microgram to 1 milligram for each administration, however.

The Fve polypeptides, etc may be administered together, either simultaneously or separately with compounds such as cytokines and / or growth factors, such as interleukin-2 (IL-2), Interleukin 12 (IL-12), GM-CSF or the like in order to strengthen the immune response. The Fve polypeptides, etc can be used in a vaccine or a therapeutic

composition either alone or in combination with other materials, for example, in the form of a lipopeptide conjugate which is known to induce a high-affinity cytotoxic T cell responses (Deres, 1989, Nature 342).

In particular, Fve diseases include allergies and cancer, described in further detail
5 below.

Cancer

Fve polypeptides, nucleic acids, and fragments, homologues, variants and derivatives thereof, are suitable for treating or preventing cancer.

The terms "cancer" and "cancerous" refer to or describe the physiological
10 condition in mammals that is typically characterized by unregulated cell growth. Examples of cancer include but are not limited to, carcinoma, lymphoma, blastoma, sarcoma, and leukemia. More particular examples of such cancers include squamous cell cancer, small-cell lung cancer, non-small cell lung cancer, gastric cancer, pancreatic cancer, glial cell tumors such as glioblastoma and neurofibromatosis, cervical cancer, ovarian cancer, liver
15 cancer, bladder cancer, hepatoma, breast cancer, colon cancer, colorectal cancer, endometrial carcinoma, salivary gland carcinoma, kidney cancer, renal cancer, prostate cancer, vulval cancer, thyroid cancer, hepatic carcinoma and various types of head and neck cancer. Further examples are solid tumor cancer including colon cancer, breast cancer, lung cancer and prostate cancer, hematopoietic malignancies including leukemias
20 and lymphomas, Hodgkin's disease, aplastic anemia, skin cancer and familiar adenomatous polyposis. Further examples include brain neoplasms, colorectal neoplasms, breast neoplasms, cervix neoplasms, eye neoplasms, liver neoplasms, lung neoplasms, pancreatic neoplasms, ovarian neoplasms, prostatic neoplasms, skin neoplasms, testicular neoplasms, neoplasms, bone neoplasms, yellow fevertrophoblastic neoplasms, fallopian
25 tube neoplasms, rectal neoplasms, colonic neoplasms, kidney neoplasms, stomach neoplasms, and parathyroid neoplasms. Breast cancer, prostate cancer, pancreatic cancer, colorectal cancer, lung cancer, malignant melanoma, leukaemia, lymphoma, ovarian cancer, cervical cancer and biliary tract carcinoma are also included.

In preferred embodiments, Fve polypeptide, nucleic acid, and fragments, homologues, variants and derivatives thereof are used to treat T cell lymphoma, melanoma or lung cancer.

- The Fve polypeptides and nucleic acids, etc, as described here, may also be used in
- 5 combination with anticancer agents such as endostatin and angiostatin or cytotoxic agents or chemotherapeutic agent. For example, drugs such as such as adriamycin, daunomycin, cis-platinum, etoposide, taxol, taxotere and alkaloids, such as vincristine, and antimetabolites such as methotrexate. The term "cytotoxic agent" as used herein refers to a substance that inhibits or prevents the function of cells and/or causes destruction of cells.
- 10 The term is intended to include radioactive isotopes (e.g. I, Y, Pr), chemotherapeutic agents, and toxins such as enzymatically active toxins of bacterial, fungal, plant or animal origin, or fragments thereof.

Also, the term includes oncogene product/tyrosine kinase inhibitors, such as the bicyclic ansamycins disclosed in WO 94/22867; 1,2-bis(aryl amino) benzoic acid derivatives disclosed in EP 600832; 6,7-diamino-phthalazin-1-one derivatives disclosed in EP 600831; 4,5-bis(aryl amino)-phthalimide derivatives as disclosed in EP 516598; or peptides which inhibit binding of a tyrosine kinase to a SH2-containing substrate protein (see WO 94/07913, for example). A "chemotherapeutic agent" is a chemical compound useful in the treatment of cancer. Examples of chemotherapeutic agents include Adriamycin, Doxorubicin, 5-Fluorouracil (5-FU), Cytosine arabinoside (Ara-C), Cyclophosphamide, Thiotepa, Busulfan, Cytoxin, Taxol, Methotrexate, Cisplatin, Melphalan, Vinblastine, Bleomycin, Etoposide, Ifosfamide, Mitomycin C, Mitoxantrone, Vincristine, VP-16, Vinorelbine, Carboplatin, Teniposide, Daunomycin, Carminomycin, Aminopterin, Dactinomycin, Mitomycins, Nicotinamide, Esperamicins (see U.S. Pat. No. 4,675,187), Melphalan and other related nitrogen mustards, and endocrine therapies (such as diethylstilbestrol (DES), Tamoxifen, LHRH antagonizing drugs, progestins, anti-progestins etc).

Allergies

Existing treatments for allergies typically involve the long-term use of steroids to depress the immune system. There are undesirable side effects with long-term steroid therapy. We demonstrate that Fve polypeptide, nucleic acid, or a fragment, homologue, variant or derivative thereof (as well as DNA vaccines, host cells and transgenic organisms comprising any of these) may be used to alleviate the symptoms of allergy, or to treat allergy. The term "allergy" as used here, refers to any allergic reactions such as allergic contact hypersensitivity.

In general, the allergy may be to an allergen from any source, for example, a source known to induce allergenic responses in humans. For example, the allergy may be to a tree pollen allergen, a grass pollen allergen, a weed pollen allergen, a feline antigen, or a fungal allergen. Thus, the allergy may be to a tree pollen allergen, for example Bet v 1 and Bet v 2 from birch tree. The allergy may be to a grass pollen allergen, for example, Phl p 1 and Phl p 2 from timothy grass. It may be to a weed pollen allergen, for example, antigen E

from ragweed. It may be to an animal allergen, for example, a canine or feline antigen. Specifically, it may be to a major feline antigen, for example, Fel d 1. The allergy may be to a fungal allergen, for example a major fungal allergen, for example, Asp f1, Asp f2, and Asp f3 from *Aspergillus fumigatus*.

5 In preferred embodiments, the allergy is to a dust mite allergen, preferably a house dust mite allergen. In particular, the allergen is preferably derived from a mite from Family Glycyphagidae or Family Pyroglyphidae. Dust mites of Family Glycyphagidae include those in the genera *Aeroglyphus*, *Austroglycyphagus*, *Blomia*, *Ctenoglyphus*,
10 *Glycyphagus*, *Gohieria*, *Lepidoglyphus*. Dust mites of Family Pyroglyphidae include those in the genera *Dermatophagoides*, *Euroglyphus*, *Pyroglyphus*. In preferred embodiments, the allergy is preferably to an allergen from a species in any of these genera.

15 In highly preferred embodiments, the allergy is to an allergen which is a group 1 allergen (Der p 1, Der f 1, Blo t 1, Eur m1, Lep d 1), a group 2 allergen (Der p 2, Der f 2, Blo t 2, Eur m 2, Lep d 2), a group 5 allergen (Blo t 5, Der p 5, Der f 5, Eur m 5, Lep d 5) or a group 15 allergen (Der p 15, Der f 15, Blot 15, Eur m 15, Lep d 15) from dust mite.

20 Allergies suitable for treatment with Fve polypeptide, nucleic acid, or a fragment, homologue, variant or derivative thereof may therefore include a seasonal respiratory allergy, allergic rhinitis, hayfever, nonallergic rhinitis, vasomotor rhinitis, irritant rhinitis, an allergy against grass pollens, tree pollens or animal danders, an allergy associated with allergic asthma, and food allergies. In particular, and as described elsewhere, Fve polypeptide, nucleic acid, or a fragment, homologue, variant or derivative thereof may be used to treat allergies to house dust mite (*Dermatophagoides* spp), preferably *Dermatophagoides pteronyssinus* or *Dermatophagoides farinae*, or to fungi or fungal spores, preferably *Aspergillus fumigatus*. Preferably, the allergens are comprised in faeces
25 of *Dermatophagoides* spp.

Viral Infections

The immunomodulator-viral infectious antigen combinations, preferably conjugates, may be used to treat or prevent any of a number of viral infectious diseases. The virus concerned may be an RNA virus or a DNA virus. Preferably, the virus is an 5 integrating virus. Preferably, the virus is selected from a lentivirus and a herpesvirus. More preferably, the virus is an HIV virus or a HSV virus.

The methods described here can therefore be used to prevent the development and establishment of diseases caused by or associated with any of the above viruses, including 10 human immunodeficiency virus, such as HIV-1 and HIV-2, and herpesvirus, for example HSV-1, HSV-2, HSV-7 and HSV-8, as well as human cytomegalovirus, varicella-zoster virus, Epstein-Barr virus and human herpesvirus 6.in humans.

Examples of viruses which may be targeted using the methods and compositions described here are given in the tables below.

DNA VIRUSES			
Family	Genus or [Subfamily]	Example	Diseases
Herpesviridae	[Alphaherpesvirinae]	Herpes simplex virus type 1 (aka HHV-1) Herpes simplex virus type 2 (aka HHV-2) Varicella zoster virus (aka HHV-3)	Encephalitis, cold sores, gingivostomatitis Genital herpes, encephalitis Chickenpox, shingles
	[Gammaherpesvirinae]	Epstein Barr virus (aka HHV-4) Kaposi's sarcoma associated herpesvirus, KSHV (aka Human herpesvirus 8)	Mononucleosis, hepatitis, tumors (BL, NPC) ?Probably: tumors, inc. Kaposi's sarcoma (KS) and some B cell lymphomas
	[Betaherpesvirinae]	Human cytomegalovirus (aka HHV-5) Human herpesvirus 6 Human herpesvirus 7 Human adenoviruses Human papillomaviruses JC, BK viruses	Mononucleosis, hepatitis, pneumonitis, congenital Roseola (aka E. subitum), pneumonitis Some cases of roseola?
Adenoviridae	Mastadenovirus	50 serotypes (species); respiratory infections	
Papovaviridae	Papillomavirus	80 species; warts and tumors	
Hepadnaviridae	Polyomavirus	Mild usually; JC causes PML in AIDS	
Poxviridae	Orthopoxvirus	Hepatitis B virus (HBV) Hepatitis C virus (HCV) Vaccinia virus Monkeypox virus	Hepatitis (chronic), cirrhosis, liver tumors Hepatitis (chronic), cirrhosis, liver tumors Smallpox vaccine virus Smallpox-like disease; a rare zoonosis (recent outbreak in Congo; 92 cases from 2/96 - 2/97)
Parvoviridae	Parapoxvirus	Orf virus	Skin lesions ("pocks")
	Erythrovirus	B19 parvovirus	E. infectiosum (aka Fifth disease), aplastic crisis, fetal loss
Circoviridae	Dependovirus	Adeno-associated virus	Useful for gene therapy; integrates into chromosome

	Circovirus	TT virus (TTV)	Linked to hepatitis of unknown etiology
RNA VIRUSES			
Family	Genus or [Subfamily]	Example	Diseases
Picornaviridae	Enterovirus	Polioviruses Echoviruses Coxsackieviruses Hepatovirus Rhinovirus Calicivirus	3 types; Aseptic meningitis, paralytic poliomyelitis 30 types; Aseptic meningitis, rashes 30 types; Aseptic meningitis, myopericarditis Acute hepatitis (fecal-oral spread) 115 types; Common cold Gastrointestinal illness
Caliciviridae		Human rhinoviruses Norwalk virus	4 types; Common cold, bronchiolitis, pneumonia
Paramyxoviridae	Paramyxovirus	Parainfluenza viruses Rubulavirus Morbillivirus	Mumps: parotitis, aseptic meningitis (rare: orchitis, encephalitis) Measles: fever, rash (rare: encephalitis, SSPE)
		Pneumovirus	Common cold (adults), bronchiolitis, pneumonia (infants)
Orthomyxoviridae	Influenzavirus A	Respiratory syncytial virus Influenza virus A	Flu: fever, myalgia, malaise, cough, pneumonia
	Influenzavirus B	Influenza virus B	Flu: fever, myalgia, malaise, cough, pneumonia
Rhabdoviridae	Lyssavirus	Rabies virus	Rabies: long incubation, then CNS disease, death
Filoviridae	Filovirus	Ebola and Marburg viruses	Hemorrhagic fever, death
Bornaviridae	Bornavirus	Borna disease virus	Uncertain; linked to schizophrenia-like disease in some animals
Retroviridae	Deltaretrovirus	Human T-lymphotropic virus type-1 Human foamy viruses Human immunodeficiency virus type-1 and -2	Adult T-cell leukemia (ATL), tropical spastic paraparesis (TSP) No disease known AIDS, CNS disease
Togaviridae	Rubivirus	Rubella virus	Mild exanthem; congenital fetal defects
	Alphavirus	Equine encephalitis viruses (WEE, EEE, VEE)	Mosquito-born, encephalitis
Flaviviridae	Flavivirus	Yellow fever virus Dengue virus St. Louis Encephalitis virus Hepatitis C virus Hepatitis G virus	Mosquito-born; fever, hepatitis (yellow fever!) Mosquito-born; hemorrhagic fever Mosquito-born; encephalitis Hepatitis (often chronic), liver cancer Hepatitis???
Reoviridae	Rotavirus	Human rotaviruses	Numerous serotypes; Diarrhea
	Coltivirus	Colorado Tick Fever virus	Tick-born; fever
Bunyaviridae	Orthoreovirus	Human reoviruses	Minimal disease
	Hantavirus	Pulmonary Syndrome Hantavirus Hantaan virus	Rodent spread; pulmonary illness (can be lethal, "Four Corners" outbreak) Rodent spread; hemorrhagic fever with renal syndrome
	Phlebovirus	Rift Valley Fever virus	Mosquito-born; hemorrhagic fever
	Nairovirus	Crimean-Congo Hemorrhagic Fever virus	Mosquito-born; hemorrhagic fever
Arenaviridae	Arenavirus	Lymphocytic Choriomeningitis virus Lassa virus	Rodent-born; fever, aseptic meningitis
Coronaviridae	Deltavirus	Hepatitis Delta virus	Rodent-born; severe hemorrhagic fever (BL4 agents; also: Machupo, Junin)
Astroviridae	Coronavirus	Human coronaviruses	Requires HBV to grow; hepatitis, liver cancer
	Astrovirus	Human astroviruses	Mild common cold-like illness Gastroenteritis
Unclassified	"Hepatitis E-like viruses"	Hepatitis E virus	Hepatitis (acute); fecal-oral spread

Human Immunodeficiency Virus-1 (HIV-1)

The combinations and conjugates described here, including Fve polypeptide combinations and conjugates, may be used to treat or prevent Human Immunodeficiency Virus (HIV) infection. The methods described here can therefore be used to prevent the development and establishment of diseases caused by or associated with human immunodeficiency virus, such as HIV-1 and HIV-2.

Human Immunodeficiency Virus (HIV) is a retrovirus which infects cells of the immune system, most importantly CD4⁺ T lymphocytes. CD4⁺ T lymphocytes are important, not only in terms of their direct role in immune function, but also in stimulating normal function in other components of the immune system, including CD8⁺ T-lymphocytes. These HIV infected cells have their function disturbed by several mechanisms and/or are rapidly killed by viral replication. The end result of chronic HIV infection is gradual depletion of CD4⁺ T lymphocytes, reduced immune capacity, and ultimately the development of AIDS, leading to death.

The regulation of HIV gene expression is accomplished by a combination of both cellular and viral factors. HIV gene expression is regulated at both the transcriptional and post-transcriptional levels. The HIV genes can be divided into the early genes and the late genes. The early genes, Tat, Rev, and Nef, are expressed in a Rev-independent manner. The mRNAs encoding the late genes, Gag, Pol, Env, Vpr, Vpu, and Vif require Rev to be cytoplasmically localized and expressed. HIV transcription is mediated by a single promoter in the 5' LTR. Expression from the 5' LTR generates a 9-kb primary transcript that has the potential to encode all nine HIV genes. The primary transcript is roughly 600 bases shorter than the provirus. The primary transcript can be spliced into one of more than 30 mRNA species or packaged without further modification into virion particles (to serve as the viral RNA genome).

Any of the HIV proteins disclosed here may be used as a viral infectious antigen for productions of conjugates and combinations as described above.

Herpes Virus

The combinations and conjugates described here, including Fve polypeptide combinations and conjugates, may be used to treat or prevent Herpesvirus infection. The methods described here can therefore be used to prevent the development and 5 establishment of diseases caused by or associated with herpesvirus, for example HSV-1, HSV-2, HSV-7 and HSV-8.

Particular examples of herpesvirus include: herpes simplex virus 1 ("HSV-1"), herpes simplex virus 2 ("HSV-2"), human cytomegalovirus ("HCMV"), varicella-zoster virus ("VZV"), Epstein-Barr virus ("EBV"), human herpesvirus 6 ("HHV6"), herpes simplex virus 7 ("HSV-7") and herpes simplex virus 8 ("HSV-8").
10

Herpesviruses have also been isolated from horses, cattle, pigs (pseudorabies virus ("PSV") and porcine cytomegalovirus), chickens (infectious laryngotracheitis), chimpanzees, birds (Marck's disease herpesvirus 1 and 2), turkeys and fish (see "Herpesviridae: A Brief Introduction", Virology, Second Edition, edited by B. N. Fields,
15 Chapter 64, 1787 (1990)).

Herpes simplex viral ("HSV") infection is generally a recurrent viral infection characterized by the appearance on the skin or mucous membranes of single or multiple clusters of small vesicles, filled with clear fluid, on slightly raised inflammatory bases. The herpes simplex virus is a relatively large-sized virus. HSV-2 commonly causes herpes labialis. HSV-2 is usually, though not always, recoverable from genital lesions. Ordinarily, 20 HSV-2 is transmitted venereally.

Diseases caused by varicella-zoster virus (human herpesvirus 3) include varicella (chickenpox) and zoster (shingles). Cytomegalovirus (human herpesvirus 5) is responsible for cytomegalic inclusion disease in infants. There is presently no specific treatment for 25 treating patients infected with cytomegalovirus. Epstein-Barr virus (human herpesvirus 4) is the causative agent of infectious mononucleosis and has been associated with Burkitt's lymphoma and nasopharyngeal carcinoma. Animal herpesviruses which may pose a

problem for humans include B virus (herpesvirus of Old World Monkeys) and Marmoset herpesvirus (herpesvirus of New World Monkeys).

Herpes simplex virus 1 (HSV-1) is a human pathogen capable of becoming latent in nerve cells. Like all the other members of *Herpesviridae* it has a complex architecture and double-stranded linear DNA genome which encodes for variety of viral proteins including DNA pol. and TK.

HSV gene expression proceeds in a sequential and strictly regulated manner and can be divided into at least three phases, termed immediate-early (IE or α), early (β) and late (γ). The cascade of HSV-1 gene expression starts from IE genes, which are expressed immediately after lytic infection begins. The IE proteins regulate the expression of later classes of genes (early and late) as well as their own expression. The product of IE175k (ICP4) gene is critical for HSV-1 gene regulation and ts mutants in this gene are blocked at IE stage of infection.

The IE genes themselves are activated by a virion structural protein VP16 (expressed late in the replicative cycle and incorporated into HSV particle). All 5 IE genes of HSV-1 (IE110k - 2 copies/HSV genome, IE175 - 2 copies/HSV genome, IE68k, IE63k and IE12k) have at least one copy of a conserved promoter/enhancer sequence - TAATGARAT. This sequence is recognized by the transactivation complex which consists of; Oct-1, HCF and VP16. The GARAT element is required for efficient transactivation by VP16. This mechanism of gene activation is unique for HSV and despite Oct-1 being a common transcription factor, the Oct-1/HCF/VP16 complex activates specifically only HSV IE genes.

Any of the herpesvirus proteins disclosed here may be used as a viral infectious antigen for productions of conjugates and combinations as described above.

CYTOKINES

In a further embodiment, the Fve polypeptide, nucleic acid, fragment, homologue, variant or derivative thereof is used to modulate cytokine levels in an individual. Preferably, the level of inflammatory cytokines is down-regulated. Examples of 5 inflammatory cytokines include Granulocyte-Macrophage-Colony stimulating factor (GM-CSF), as well as any cytokine that mediates migration of alveolar macrophages into the lung and act to increase cell proliferation.

The term "cytokine" may be used to refer to any of a number of soluble molecules (e.g., glycoproteins) released by cells of the immune system, which act nonenzymatically 10 through specific receptors to regulate immune responses. Cytokines resemble hormones in that they act at low concentrations bound with high affinity to a specific receptor. Preferably, the term "cytokine" refers to a diverse group of soluble proteins and peptides which act as humoral regulators at nano- to picomolar concentrations and which, either under normal or pathological conditions, modulate the functional activities of individual 15 cells and tissues.

Particular examples of cytokines which are suitable for use in the methods and compositions described include interleukins, lymphokine, interferon, Colony Stimulating Factors (CSFs) such as Granulocyte-Colony Stimulating Factor (G-CSF), Macrophage-Colony stimulating factor (M-CSF) and Granulocyte-Macrophage-Colony stimulating 20 factor (GM-CSF), GSF, Platelet-Activating Factors (PAF), Tumor Necrosis Factor (TNF).

Thus, interleukins such as IL1, IL2 and IL4, as well as interferons such as IFN- α , IFN- β and IFN- γ are included. Tumour necrosis factors TNF- α (cachetin), TNF- β (lymphotoxin) may also be suitably employed.

Preferred cytokines are those which are capable of recruiting immune responses, 25 for example, stimulation of dendritic cell or cytotoxic T cell activity, or which are capable

of recruiting macrophages to the target site. In a highly preferred embodiment, the cytokine comprises IL-2, GM-CSF or GSF.

CHEMICAL COUPLING

As noted above, the immunomodulator may be coupled to the allergen by a number 5 of methods. Crosslinkers are divided into homobifunctional crosslinkers, containing two identical reactive groups, or heterobifunctional crosslinkers, with two different reactive groups. Heterobifunctional crosslinkers allow sequential conjugations, minimizing polymerization.

Any of the homobifunctional or heterobifunctional crosslinkers presented in the 10 table below may be used to couple the allergen with the immunomodulator to produce an immunomodulator-allergen conjugate.

Homobifunctional

Reagent	Cat. No.	Modifi ed Group	Solubility	Comments	Refs
BMME	442635-Y	-SH	DMF, Acetone	Homobifunctional crosslinker useful for formation of conjugates via thiol groups.	Weston, P.D., et al. 1980. Biochem. Biophys Acta. 612, 40.
BSOCOES	203851-Y	-NH2	Water	Base cleavable crosslinker useful for studying receptors and mapping surface polypeptide antigens on lymphocytes.	Howard, A.D., et al. 1985. J. Biol. Chem. 260, 10833.
DSP	322133-Y	-NH2	Water	Thiol cleavable crosslinker used to immobilize proteins on supports containing amino groups.	Lee, W.T., and Conrad, D.H. 1985. J. Immunol. 134, 518.
DSS	322131-Y	-NH2	Water	Non-cleavable, membrane impermeable crosslinker widely used for conjugating radiolabeled ligands to cell surface receptors and for detecting conformational changes in membrane proteins.	D'Souza, S.E., et al. 1988. J. Biol. Chem. 263, 3943.
EGS	324550-Y	-NH2	DMSO	Hydroxylamine cleavable reagent for crosslinking and reversible immobilization of proteins through their primary amine groups. Useful for studying structure-function relationships.	Geisler, N., et al. 1992. Eur. J. Biochem. 206, 841. 14. Moenner, M., et al. 1986. Proc. Natl. Acad. Sci. USA 83, 5024.

EGS, Water Soluble	324551-Y	-NH2	Water	Water soluble version of EGS that reacts rapidly with dilute proteins at neutral pH. Crosslinked proteins are readily cleaved with hydroxylamine at pH 8.5 for 3-6 hours, 37°C.	Yanagi, T., et al. 1989. Agric. Biol. Chem.53, 525.
Glutaraldehyde	354400-Y	-OH	Water	Used for crosslinking proteins and polyhydroxy materials. Conjugates haptens to carrier proteins; also used as a tissue fixative.	Harlow, E., and Lane, D. 1988. Antibodies: A Laboratory Manual, Cold Spring Harbor Publications, N.Y., p. 349.
SATA	573100-Y	-NH2	DMSO	Introduces protected thiols via primary amines. When treated with hydroxylamine, yields a free sulhydryl group that can be conjugated to maleimide-modified proteins.	Duncan, R.J.S., et al. 1983. Anal. Biochem.132, 68.

Heterobifunctional

Reagent	Cat. No.	Modifi ed Group	Solubility	Comments	Refs
GMBS	442630-Y	-NH2, -SH	DMSO	Heterobifunctional crosslinker useful for preparing enzyme-antibody conjugates (e.g. -gal-IgG) and for immobilizing enzymes on solid supports.	Kitagwa, T., et al. 1983. J. Biochem.94, 1160.19. Rusin, K.M., et al. 1992. Biosens. Bioelectron.7, 367.
MBS	442625-Y 442626-Y	-NH2, -SH -NH2, -SH	DMSO, Water	Thiol cleavable, heterobifunctional reagent especially useful for preparing peptide-carrier conjugates and conjugating toxins to antibodies.	Green, N., et al. 1982. Cell.28, 477.
PMPI	528250-Y	-SH2, -OH	DMSO, DMF	Used in the preparation of alkaline phosphatase conjugates of estradiol, progesterone, serine-enriched peptides, and vitamin B12.	Aithal, H.N., et al. 1988. J. Immunol. Methods12, 63.
SMCC	573114-Y 573115-Y	-NH2, -SH -NH2, -SH	DMF, AN Acetonitrile Water	Heterobifunctional reagent for enzyme labeling of antibodies and antibody fragments. The cyclohexane bridge provides extra stability to the maleimide group. Ideal reagent for preserving enzyme activity and antibody specificity after coupling.	Annunziato, M.E., et al. 1993. Bioconjugate Chem.4, 212.
SPDP	573112-Y	-NH2, -SH	DMF, AN Acetonitrile	Introduces protected thiol groups to amine groups. Thiolated proteins can be coupled to a second molecule via an iodoacetamide or maleimide group, or to a second pyridyldisulfide	Caruelle, D., et al. 1988. Anal. Biochem.173, 328.

				containing molecule.	
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Each of these reagents may be obtained from a number of manufacturers, for example, from Calbiochem (catalogue number in column 2), or Piece Chemical Company.

PHARMACEUTICAL COMPOSITIONS

Fve polypeptides may be produced in large amounts at low cost in a bioactive form, allowing the development of Fve containing formulations by aerosolisation, nebulisation, intranasal or intratracheal administration.

While it is possible for the composition comprising the Fve polypeptide or nucleic acid to be administered alone, it is preferable to formulate the active ingredient as a pharmaceutical formulation. We therefore also disclose pharmaceutical compositions comprising Fve polypeptide or nucleic acid, or a fragment, homologue, variant or derivative thereof. Such pharmaceutical compositions are useful for delivery of Fve polypeptide, nucleic acid, fragment, homologue, variant or derivative thereof to an individual for the treatment or alleviation of symptoms as described.

The composition may include the Fve polypeptide, nucleic acid, fragment, homologue, variant or derivative thereof, a structurally related compound, or an acidic salt thereof. The pharmaceutical formulations comprise an effective amount of Fve polypeptide, nucleic acid, fragment, homologue, variant or derivative thereof, together with one or more pharmaceutically-acceptable carriers. An "effective amount" of an Fve polypeptide, nucleic acid fragment, homologue, variant or derivative thereof is the amount sufficient to alleviate at least one symptom of a disease as described.

The effective amount will vary depending upon the particular disease or syndrome to be treated or alleviated, as well as other factors including the age and weight of the patient, how advanced the disease etc state is, the general health of the patient, the severity of the symptoms, and whether the Fvc polypeptide, nucleic acid, fragment, homologue,

variant or derivative thereof is being administered alone or in combination with other therapies.

Suitable pharmaceutically acceptable carriers are well known in the art and vary with the desired form and mode of administration of the pharmaceutical formulation. For 5 example, they can include diluents or excipients such as fillers, binders, wetting agents, disintegrators, surface-active agents, lubricants and the like. Typically, the carrier is a solid, a liquid or a vaporizable carrier, or a combination thereof. Each carrier should be "acceptable" in the sense of being compatible with the other ingredients in the formulation and not injurious to the patient. The carrier should be biologically acceptable without 10 eliciting an adverse reaction (e.g. immune response) when administered to the host.

The pharmaceutical compositions disclosed here include those suitable for topical and oral administration, with topical formulations being preferred where the tissue affected is primarily the skin or epidermis (for example, psoriasis, eczema and other epidermal diseases). The topical formulations include those pharmaceutical forms in which the 15 composition is applied externally by direct contact with the skin surface to be treated. A conventional pharmaceutical form for topical application includes a soak, an ointment, a cream, a lotion, a paste, a gel, a stick, a spray, an aerosol, a bath oil, a solution and the like. Topical therapy is delivered by various vehicles, the choice of vehicle can be important and generally is related to whether an acute or chronic disease is to be treated. Other 20 formulations for topical application include shampoos, soaps, shake lotions, and the like, particularly those formulated to leave a residue on the underlying skin, such as the scalp (Arndt et al, in Dermatology In General Medicine 2:2838 (1993)).

In general, the concentration of the Fve polypeptide, nucleic acid, fragment, homologue, variant or derivative thereof composition in the topical formulation is in an 25 amount of about 0.5 to 50% by weight of the composition, preferably about 1 to 30%, more preferably about 2-20%, and most preferably about 5-10%. The concentration used can be in the upper portion of the range initially, as treatment continues, the concentration can be lowered or the application of the formulation may be less frequent. Topical

applications are often applied twice daily. However, once-daily application of a larger dose or more frequent applications of a smaller dose may be effective. The stratum corneum may act as a reservoir and allow gradual penetration of a drug into the viable skin layers over a prolonged period of time.

- 5 In a topical application, a sufficient amount of active ingredient must penetrate a patient's skin in order to obtain a desired pharmacological effect. It is generally understood that the absorption of drug into the skin is a function of the nature of the drug, the behaviour of the vehicle, and the skin. Three major variables account for differences in the rate of absorption or flux of different topical drugs or the same drug in different vehicles;
- 10 the concentration of drug in the vehicle, the partition coefficient of drug between the stratum corneum and the vehicle and the diffusion coefficient of drug in the stratum corneum. To be effective for treatment, a drug must cross the stratum corneum which is responsible for the barrier function of the skin. In general, a topical formulation which exerts a high *in vitro* skin penetration is effective *in vivo*. Ostrenga et al (J. Pharm. Sci., 15 60:1175-1179 (1971) demonstrated that *in vivo* efficacy of topically applied steroids was proportional to the steroid penetration rate into dermatomed human skin *in vitro*.

- A skin penetration enhancer which is dermatologically acceptable and compatible with the agent can be incorporated into the formulation to increase the penetration of the active compound(s) from the skin surface into epidermal keratinocytes. A skin enhancer which increases the absorption of the active compound(s) into the skin reduces the amount of agent needed for an effective treatment and provides for a longer lasting effect of the formulation. Skin penetration enhancers are well known in the art. For example, dimethyl sulfoxide (U.S. Pat. No. 3,711,602); oleic acid, 1,2-butanediol surfactant (Cooper, J. Pharm. Sci., 73:1153-1156 (1984)); a combination of ethanol and oleic acid or oleyl alcohol (EP 267,617), 2-ethyl-1,3-hexanediol (WO 87/03490); decyl methyl sulphoxide and Azone.RTM. (Hadgraft, Eur. J. Drug. Metab. Pharmacokinet, 21:165-173 (1996)); alcohols, sulphoxides, fatty acids, esters, Azone.RTM., pyrrolidones, urea and polyoles (Kalbitz et al, Pharmazie, 51:619-637 (1996));

Terpenes such as 1,8-cineole, menthone, limonene and nerolidol (Yamane, J. Pharmacy & Pharmacology, 47:978-989 (1995)); Azone.RTM. and Transcutol (Harrison et al, Pharmaceutical Res. 13:542-546 (1996)); and oleic acid, polyethylene glycol and propylene glycol (Singh et al, Pharmazie, 51:741-744 (1996)) are known to improve skin penetration of an active ingredient.

Levels of penetration of an agent or composition can be determined by techniques known to those of skill in the art. For example, radiolabeling of the active compound, followed by measurement of the amount of radiolabeled compound absorbed by the skin enables one of skill in the art to determine levels of the composition absorbed using any of several methods of determining skin penetration of the test compound. Publications relating to skin penetration studies include Reifenrath, W G and G S Hawkins. The Weaning Yorkshire Pig as an Animal Model for Measuring Percutaneous Penetration. In: Swine in Biomedical Research (M. E. Tumbleson, Ed.) Plenum, New York, 1986, and Hawkins, G. S. Methodology for the Execution of *In Vitro* Skin Penetration Determinations. In: Methods for Skin Absorption, B W Kemppainen and W G Reifenrath, Eds., CRC Press, Boca Raton, 1990, pp.67-80; and W. G. Reifenrath, Cosmetics & Toiletries, 110:3-9 (1995).

For some applications, it is preferable to administer a long acting form of agent or composition using formulations known in the arts, such as polymers. The agent can be incorporated into a dermal patch (Junginger, H. E., in Acta Pharmaceutica Nordica 4:117 (1992); Thacharodi et al, in Biomaterials 16:145-148 (1995); Niedner R., in Hautarzt 39:761-766 (1988)) or a bandage according to methods known in the arts, to increase the efficiency of delivery of the drug to the areas to be treated.

Optionally, the topical formulations can have additional excipients for example; preservatives such as methylparaben, benzyl alcohol, sorbic acid or quaternary ammonium compound; stabilizers such as EDTA, antioxidants such as butylated hydroxytoluene or butylated hydroxyanisole, and buffers such as citrate and phosphate.

The pharmaceutical composition can be administered in an oral formulation in the form of tablets, capsules or solutions. An effective amount of the oral formulation is administered to patients 1 to 3 times daily until the symptoms of the disease alleviated. The effective amount of agent depends on the age, weight and condition of a patient. In 5 general, the daily oral dose of agent is less than 1200 mg, and more than 100 mg. The preferred daily oral dose is about 300-600 mg. Oral formulations are conveniently presented in a unit dosage form and may be prepared by any method known in the art of pharmacy. The composition may be formulated together with a suitable pharmaceutically acceptable carrier into any desired dosage form. Typical unit dosage forms include tablets, 10 pills, powders, solutions, suspensions, emulsions, granules, capsules, suppositories. In general, the formulations are prepared by uniformly and intimately bringing into association the agent composition with liquid carriers or finely divided solid carriers or both, and as necessary, shaping the product. The active ingredient can be incorporated into a variety of basic materials in the form of a liquid, powder, tablets or capsules to give an 15 effective amount of active ingredient to treat the disease.

Other therapeutic agents suitable for use herein are any compatible drugs that are effective for the intended purpose, or drugs that are complementary to the agent formulation. The formulation utilized in a combination therapy may be administered simultaneously, or sequentially with other treatment, such that a combined effect is 20 achieved.

The invention is described further, for the purpose of illustration only, in the following examples.

EXAMPLES

In each of the Examples presented below, where an activity is described for a Fve 25 polypeptide comprising a GST (glutathione S transferase) portion (for example, as a GST-FIP fusion protein), we find that the polypeptide itself, without the GST portion, has

substantially the same activity. This is to be expected, as the GST domain does not have any relevant biological activity as far as FIP is concerned.

Example 1. Isolation and Purification of Native Fve Protein from Golden Needle Mushroom

5 *Methods and materials*

Two kilograms of the fruit bodies of *Flammulina velutipes* are homogenized with 2L ice-cold 5% acetic acid in the presence of 0.05 M 2-mercaptoethanol and 0.3 M sodium chloride. The proteins in the supernatant are precipitated by 95% saturated ammonium sulfate.

10 The precipitate is re-dissolved and dialyzed against 10 mM Tris-HCl pH 8.5 (buffer A) at 4°C for 48 hours with six to eight changes of dialysis buffer. The protein solution is applied to the Q Sepharose FF column (2.6 × 10 cm, Pharmacia) that has been previously equilibrated with buffer A. The unbound fraction is collected and dialyzed against 10 mM sodium acetate pH 5.0 (buffer B) at 4°C for 48 hours with six to eight
15 changes of dialysis buffer and then further purified by applying to the SP Sepharose FF column (2.6 × 10 cm, Pharmacia) that has been previously equilibrated with buffer B.

The protein is eluted with a gradient of 0-0.5 M NaCl in buffer B. Fractions containing Fve protein are collected and analyzed by a 7.5% Tris-Tricine SDS-PAGE.

Results

20 *High yield of native Fve protein is purified from Flammulina velutipes*

The native Fve protein has an apparent molecular weight of 12.7 kDa as determined by SDS-PAGE (Figure 1A). However, it appears to be a homodimer with a molecular weight of 25.5 kDa as determined by Superdex 75 (26 × 60 cm, Pharmacia) gel filtration chromatography (Figure 1B and 1C). The running buffer for gel filtration is 10
25 mM Tris-HCl pH 7.5, 0.2 M sodium chloride.

Fve protein is the major component in the crude extract from the mushroom fruit bodies. By removing the cap of the mushroom, we managed to reduce the amount of polysaccharides that cause undesirable interference in the process of protein purification.

The yield of native Fve protein is 40 mg from 1 kg wet-weight of starting material.

5 **Example 2. Measurement of gene expression profile at mRNA level after Fve stimulation**

Methods and Materials

Two subsets of effector Th cells have been defined on the basis of their distinct cytokine secretion patterns and immunomodulatory effects (Mosmann et al., 1989; Paul 10 and seder, 1994; Abbas et al., 1996). Th1 cells produce inflammatory cytokines, such as IFN- γ , TNF- α , IL-12, IL-15 and IL-18, and enhance cellular immunity mediated by macrophages. In contrast, Th2 cells produce a different group of cytokines, such as IL-4, IL-5, IL-6 and IL-13. The differentiation of precursor T cells into Th1 or Th2 cells has important biologic implication in terms of susceptibility or resistance to a particular 15 disease.

In order to characterize the cytokines expression pattern induced by Fve, human PBMC from healthy donor and splenocytes from 8 week-old BALB/cJ mice are collected and cultured with 20 μ g of native Fve . The mRNA is extracted at 48 hours using RNeasy Mini mRNA Purification Kit (QIAGÉN). First-strand cDNA is then generated from the 20 mRNA template using oligo-dT primers and MMLV reverse transcriptase (Promega).

PCR reactions are performed with Taq polymerase (Promega) with standard conditions and optimized annealing temperatures. The amplified products are analysed by electrophoresis in 1.5% agarose gel containing ethidium bromide (0.5 μ g /ml) and photographed with UV exposure. Messenger RNA for various cytokines and transcription 25 factors are measured. House keeping genes mRNA for hypoxanthine ribosyl-transferase (HPRT) and cyclophilin are used as internal controls.

Results

Enhanced expression of IFN- γ , TNF- α , IL-1 β , IL-2, IRF-1, c-Rel, Bcl-X_L, ICAM-1, and iNOS mRNA

Human PBMC and spleen cells from BALB/cJ mice are cultured with 20 μ g of Fve
5 and analyzed for cytokine mRNA expression at 48hr. The results indicated that there is an increase in IFN- γ , TNF- α , iNOS mRNA production by spleen cells cultured with Fve protein. Mouse IL-12 remains unchanged. This phenomenon occurred in a dose dependent manner.

Similar results are seen in human PBMC. The mRNA for human cytokines IL-1 β ,
10 IL-2, IFN- γ and TNF- α ; transcription factor IRF-1 and c-Rel; adhesion molecule ICAM-1 and anti-apoptotic protein Bcl-X_L is up regulated after Fve stimulation. Figure 2 and Figure 3 show the patterns of mRNA expression for transcription factors, cytokines and adhesion molecules of the splenocytes and PBMC stimulated by Fve.

Example 3. Generation of Fve Mutants By PCR-Based Mutagenesis

15 Materials and Methods

A cDNA encoding for the Fve protein is cloned into the BamHI and EcoRI site of pGEX-4T1. This DNA template is used to generate a panel of mutants by recombinant-PCR method (Figure 4). A schematic representation of the strategy used to generate mutants is shown in Figure 5.

20 As predicted by PHD prediction program, Fve contains one α -helix, six β -strands and two β -turns. Each of these predicted secondary structures is serially deleted by recombinant-PCR method. In addition, we also examined the potential function of the R27, G28, T29 residues, which resembles the cell aggregating RGD motif, located in the N-terminal β -turn of Fve protein by point mutation. Each of the amino acid residues of
25 RGT is substituted by alanine residue.

A partial list of fragments of Fve is shown in **Appendix B**.

Example 4. Production of the Fve-Derived Mutant Proteins

Materials and methods

Eleven deletion mutants and three point mutants of Fve DNA are generated. Each
5 of the polypeptides is expressed in TG1 *E.coli* cells as fusion protein with GST carrier
protein and purified by glutathione affinity column. All the mutants could express protein
except insoluble mutant D6-18, in which α -helix has been deleted.

Figure 6 shows the panel of the affinity purified mutant proteins on a SDS-PAGE.
These purified proteins are used for the cell aggregation, hemagglutination and
10 lymphocytes proliferation assay.

Example 5. Comparison of Hemagglutination Activity of Fve Mutants

Materials and methods

5ml of whole human blood obtained from a healthy volunteer is centrifuged at
2500Xg for 10min. The plasma is removed and 2ml of packed red blood cells are collected
15 from the bottom of the tube.

The red blood cells (RBC) are diluted 5X with 1xPBS buffer and centrifuged at
1200Xg for 10min. RBC pellet is resuspended in 1.5%(v/v) of 1xPBS. 50ul of 2x serial
dilutions (from 64 μ g /ml to 0.25 μ g /ml) of each Fve mutant protein is added into 50ul of
20 0.2% gelatin in 1xPBS (pH 7.4) and then mixed with 100ul of 1.5% RBC in each well of
the 96-well round bottom microtiter plates. Cells are incubated at room temperature and
examined for hemagglutination after 2 hours and over night, respectively (Table 1).

Example 6. Lymphocytes Aggregation Activity of Fve and Its Mutants

Materials and methods

Human peripheral blood mononuclear cells (PBMC) from a healthy donor are isolated and cells are then cultured with 20 μ g /ml of various Fve mutants in 24-well plates.

- 5 Cells aggregation is observed by inverted light microscopy after 24 hours (Table 1).

Results

Mutant GST-FveG28A lost the hemagglutination and lymphocytes aggregation activity

- Native Fve, GST-Fve (wild type) and two point mutants, GST-FveR27A and GST-FveT29A, show positive aggregation and hemagglutination activity. These properties are not seen in all the deletion mutants and a point mutant GST-FveG28A. PHA and ConA are used as positive controls; GST and Blo t 5 are used as negative controls. These results are summarized in Table 1.

- The Arg-Gly-Asp (RGD) tripeptide sequence is the most common molecular recognition site implicated in several immunological reactions. Normally RGD motif is located in the β -turn structure. According to the PHD prediction, residue 19 to 33 is a β -turn structure. Therefore, we propose that glycine residue of RGT (RGD-like motif) tripeptide sequence at position 28 plays an important role on lymphocyte aggregation/adhesion. The potentially interaction between Fve and the proteins of integrin family will be addressed.

	Cell aggregation	Hemagglutination
D19-33	-	-
D34-46	-	-
D47-60	-	-
D61-72	-	-
D73-84	-	-
D85-97	-	-
D98-106	-	-
D107-115	-	-
P55-100	-	-

D61-97	-	-
*R27A	+	+
**G28A	-	-
***T29A	+	+
rGST-Fve	+	+
nFve	+	+
GST	-	-
Biot 5	-	-
ConA	+	+
PHA	+	+

Table 1. Lymphocytes aggregation and RBC hemagglutination activities of Fve mutants

Example 7. Lymphoproliferation Activity of Fve Mutants

5 Materials and methods

Splenocytes from Balb/cJ mice and peripheral blood mononuclear cells (PBMC) from a healthy donor are stimulated with 2.5µg /ml, 5µg /ml, 10µg /ml or 20µg /ml respectively of Fve mutant proteins for 24 hours. Then 1 µCi [³H]-thymidine is added to the culture and further incubated for 18 hours. [³H]-thymidine incorporation is measured in triplicates by a β counter (Beckman).

10

Results

Figure 7 and 8 show the results of the proliferation assay for the panel of proteins tested. Deletion mutants D19-33, D73-84, P55-100, and mutant with single amino acid substitution G28A showed significant reduction in lymphoproliferation activity in mouse splenocytes, whereas, such reduction is less pronounced for the rest of the mutants tested (Figure 7).

15

Interestingly, some mutants such as D34-46, D47-60 and D61-72, which show negative hemagglutination and cell aggregation, retain similar lymphoproliferative

activity as the wild type protein. For the result of human PBMC, deletion mutant D61-72 and mutant with single amino acid substitution G28A show more than 50% reduction in lymphoproliferation activity (Figure 8). Taken together the proliferation results from mouse splenocytes and human PBMC demonstrate that glycine at position 28 plays an key role in lymphocyte proliferation.

5

Example 8. Recombinant GST-Fve (Wild Type) and GST-FveT29A (Mutant) Show Similar Proliferative Activity of CD3⁺ T Cells as the Native Fve

Materials and methods

Human peripheral blood mononuclear cells (PBMC) from a healthy donor are isolated according to the standard protocol (Coligan et al., 1998). The cells are then cultured with 20 μ g /ml of recombinant wild type GST-Fve and mutant GST-FveT29A for 5 days. Cells are stained with anti-CD3⁺ PerCP monoclonal antibody (Becton Dickinson), and analyzed by FACScan flow cytometry (Becton Dickinson).

10

Results

15

A histogram shows that 8% and 17% enrichment of T cells are detected after stimulation with recombinant wild type GST-Fve and mutant GST-FveT29A for 5 days (Figure 9). Results showed that both recombinant wild type GST-Fve and mutant GST-FveT29A showed comparable lymphoproliferative activity of T lymphocytes as well as the native Fve protein.

20

These data suggest that Fve-mediated T cell polarization and enrichment is detectable at day 5.

Example 9. Detection of IFN- γ and TNF- α by Intracellular Cytokine Staining After Stimulation with Recombinant GST-Fve Protein*Methods and Materials*

Intracellular cytokine staining is done by modification of a standard method from

5 PharMingen. Briefly, human PBMC are stimulated *in vitro* with 20 μ g of native Fve protein, GST, recombinant GST-Fve, GST-R27A, GST-G28A, or with GST-T29A. GlogiPlugTM (PharMingen) is added 48hr after the cultures are initiated, cells are collected 14 hr later and then stained for T cells surface marker (CD3) in FACS buffer containing GlogiPlugTM. Cells are then treated with Cytofix/Cytoperm (PharMingen) for 30min. Cells
10 are incubated with cytokine antibodies for 30min after washing with Perm Wash buffer (PharMingen). Finally, cells are washed with PBS containing 1% paraformaldehyde and then analyzed by FACSCalibur flow cytometry (BD Biosciences). CellQuest software (BD Biosciences) is used for data analysis.

Results

15 The results show that native Fve protein is able to stimulate production of IL-2, IFN- γ , TNF- α , but not IL-4 in CD3⁺ T cells (Figure 10). Similar results are seen for the recombinant wild type GST-Fve and two mutants GST-FveR27A, GST-FveT29A. Strikingly, recombinant mutant GST-FveG28A failed to stimulate the production of such cytokines (Figure 11 and 12).

20 The percentages of IFN- γ production induced by GST, GST-Fve, GST-FveR27A, GST-FveG28A, GST-FveT29A are 0.8%, 12.3%, 14.3%, 1.8%, 17.6%, respectively. In contrast, the percentages of TNF- α production which induced by GST, GST-Fve, GST-FveR27A, GST-FveG28A, GST-FveT29A are 1.2%, 21.5%, 18.7%, 1.5%, 14.4%, respectively (Table 2). This data provides further evidence that the glycine residue at
25 position 28 of Fve protein plays an important role in the biological function such as aggregation/adhesion, cytokines production, proliferation, and differentiation of lymphocytes. Further examination of the physiological role of RGT sequence in Fve protein by using blocking monoclonal antibodies and peptide inhibition assay are carried

out to confirm this function. The possibility of integrin-mediated T/NK-cell adhesion is also investigated.

In summary, mutants FveR27A and FveT29A show enhanced mitogenic activities as compared to that of wild type Fve. In addition, the solubility of both mutant proteins is significantly increased in comparison with that of wild type Fve. This improved solubility will greatly facilitate the large scale production of such recombinant protein.

Recombinant proteins	Intracellular IFN- γ	Intracellular TNF- α
GST	0.8%	1.2%
GST-FveWT	12.3%	21.5%
GST-FveR27A	14.3%	18.7%
GST-FveG28A	1.8%	1.5%
GST-FveT29A	17.6%	14.4%

Table 2: The percentage of intracellular cytokines production in CD3⁺ T lymphocytes during stimulation with three different Fve mutants with single amino acid substitution

10 Example 10. Applications of Fve in Allergy

The increasing prevalence of atopic diseases such as hayfever or allergic asthma is a major problem in most developing and developed countries. Accumulating evidence indicates that appropriate immunotherapy prevents the onset of new sensitization and the progress of allergic rhinitis to asthma.

15 The central role of allergen-specific Th2 cells in the regulation of allergic inflammation has been highlighted. Exploration of novel and effective treatment for atopic diseases is active area of allergy research. Induction of allergen-specific T regulatory immune response, suppression of the effects of IL-4, IL-5 and IL-13 cytokines, and redirecting/balancing Th2 immune response in allergy is an attractive and promising 20 approach to pursue (Akbari et al., 2002; Scanga and Le Gros, 2000; Zuany-Amorim et al., 2002).

Our *in vitro* and *in vivo* studies reveal that Fve interacts with T and NK cells.

Fve-activated T cells produce Th1-skewed cytokines in high levels, and suppress Th2 cytokines (IL-4 and IL-13) production. Thus these biological activities of Fve can be exploited to treat Th2-associated diseases such allergic asthma and rhinitis. The use of the 5 immunomodulatory properties of Fve to treat allergic diseases is novel because there are a number of differences between Fve approach and other existing methods such as hexameric motifs, called CpG motifs or DNA immunostimulatory sequences (ISS).

The function of ISS is act as a danger signal to stimulate non-specific innate immune response (Krieg 2000). It is known that ISS is recognized by the toll-like receptor 10 9 on B cells and CD123⁺ dendritic cells. It is unexpected that TLR9 is also involved in autoimmunity (Leadbetter et al., 2002; Krieg 2002; Vinuesa and Goodnow, 2002). Upon the detection of CpG motifs or ISS element, B cells are induced to proliferate and secrete immunoglobulin (Ig), and dendritic cells (DCs) secrete a wide array of cytokines, interferons and chemokines that promote T helper type 1 (Th1) cells. Both B and DCs up-15 regulate costimulatory molecules and have enhanced abilities to induce Th1 cell immune responses. In contrast, Fve is directly target on T and NK cells to involve in the acquire immunity.

Example 11. *In vivo* Study of the Adjuvant Effect of Fve Using a Murine Allergic Asthma Model

20 Immunotherapy with recombinant allergen in combination with certain immunomodulator enhancing Th1 but suppressing Th2 immune response is a novel approach to achieve higher efficacies in immunotherapy. Since Fve protein is an activator of Th1/Tc1 immune response, it may be used as such an immunomodulator to provide the adjuvant effects to enhance Th1-skewed immunity.

We investigated the adjuvant effects of Fve for allergy immunotherapy with a combination of a recombinant house dust mite major allergen, Der p 2 and Fve using an animal model.

Methods and Materials

- 5 A schematic representation of the experimental design is shown in Figure 13.

8 to 10 week old male BALB/cJ mice obtained from the Sembawang Laboratory Animal Center of Singapore are divided into two groups for each experiment. Mice are sensitized by intraperitoneal injection of 10 μ g of recombinant Der p 2 in aluminum hydroxide at day 0 and day 21. Twenty-eight days after the sensitization, each group of 10 mice is subcutaneously injected with 50 μ g of Der p 2 and 50 μ g of Der p 2 plus 40 μ g of Fve, respectively. A total of six injections are performed at every alternative day over a period of 12 days. Mice are then challenged with the third intraperitoneal injection of 10 μ g of Der p 2 plus aluminum hydroxide at day 42. Der p 2-specific IgG1 and IgG2a are determined weekly starting at day 14 by ELISA. Since IgG2a is the hallmark of Th1 15 immunity in mouse, titer of IgG2a is used a measure of Th1 immunity.

Results

Increase allergen-specific IgG2a production in the treatment group with combination of Fve and allergen

As shown in Figure 13, mice that are subcutaneously treated with 50 μ g of Der p 2 20 alone produced relatively lower titers of Der p 2-specific IgG2a, whereas mice treated with 50 μ g of Der p 2 plus 40 μ g of Fve showed a significant boost of Der p 2-specific IgG2a production (Figure 14).

Upon challenge with intraperitoneal immunization of Der p 2 in alum at day 42, the 25 Der p 2-specific IgG2a in Fve administered mice is further increased at day 49. It is interesting to note that the Fve-specific IgG1 and IgG2a remained low (data not shown). Similar results are observed in similar experiments performed with another house dust mite major allergen, Blo t 5, from *Bromia tropicalis* (data not shown).

Taken together, the data suggested that Fve protein may act as a potent adjuvant/immunomodulator to boost antigen-specific Th1-skewed immune response, therefore it may serve as a useful reagent to improve the efficacies of immunotherapeutic treatment of allergy in humans. The adjuvanticity and immunomodulatory property of Fve protein may be improved by biomolecular engineering.

While not wishing to be bound by theory, it is postulated that this molecule may activate NK cells and CD8⁺ T cells and thus result in production of IFN- γ . These may induce a strong cellular-mediated immune response and promote isotype switching to specific IgG2a predominantly.

10 **Example 12. Assessment of Erythema Flare and Wheal Diameter Formation Induced by Skin Prick Tests in Human Allergic Subject**

Materials and methods

The skin prick test is a convenient diagnostic method test for allergy in the clinics. The aim of this study is to evaluate the suppression effect of Fve protein to allergen 15 hypersensitivity. As an *in vivo* topical challenge method, the skin prick test is administered to a human subject with history of sensitization to house dust mite *Dermatophagoides pteronyssinus*.

25 $25\mu\text{g}/\text{ml}$ of purified recombinant Der p 2 allergen mixed with same concentration of native Fve protein or Der p 2 allergen alone, is applied to the skin of left and right hand of human subject for 10 minutes. Histamine is used as a positive control. The size of the wheel and erythematic flare diameter is measured manually.

Results

Fve reduce wheal and erythematic flare formation on Der p 2 skin prick test-positive human subject

25 The formation of wheal and erythematic flare could be detected in the challenged site of histamine, Der p 2, and Der p 2 combined with Fve. The diameter of the wheals in

both left and right hand induced by Der p 2 is 22mm and 24mm, respectively.

Interestingly, the mixture of Der p 2 and Fve reduces the wheal's diameter in both hands to 15mm and 18mm, respectively (Figure 15A). A similar reduction is also seen in the size of erythematic flare (Figure 15B, Table 3A and 3B).

5 The data indicates that there is a suppression of allergic reaction mediated by immunomodulatory effects of Fve protein. The results provide additional evidence that Fve could be used as an adjuvant for allergens immunotherapy.

10 Besides indoor allergens, outdoor allergens are also important triggering factors that lead to allergic diseases. Hay fever and allergic asthma triggered by grass pollen allergens affect approximately 20% of the population in cool temperate climates. Worldwide more than 200 million individuals are allergic to group 1 grass pollen allergens, and over 100 million individuals exhibit IgE-mediated allergic reactions against Phl p 2, a major allergen from timothy grass (*Phleum pratense*) pollen.

15 Therefore, we propose that recombinant Fve as well as the native Fve may also be applied in the treatment of other allergies that induced by tree pollen allergen (Bet v 1 and Bet v 2 from birch), grass pollen allergen (Phl p 1 and Phl p 2 from timothy grass), weed pollen allergen (antigen E from ragweed), major feline antigen (Fel d 1), major canine allergen (Der f 15), etc. Other allergens will be known to the person skilled in the art.

20 Another useful application of Fve protein in allergy is to conjugate or co-deliver with allergenic crude extracts such as mite extracts, pollen extracts, cat and dog extracts, cockroach extracts, fungal and mold extracts for desensitization by immunotherapy.

	Wheal Diameter (mm)	
	Left hand	Right hand
Saline (negative control)	0	0
Histamine	7	5
Der p 2	22	24
Der p 2 + Fve (1:1 w/w)	15	18

Table 3A: Wheal formation on skin after challenged with Der p 2

	Erythematic Flare Diameter (mm)	
	Left hand	Right hand
Saline (negative control)	0	0
Histamine	30x25	35x30
Der p 2	55x40	50x43
Der p 2 + Fve (1:1 w/w)	45x35	45x35

Table 3B: Erythematic flare formation on skin after challenge with Der p 2

FVE ADJUVANTED ALLERGEN VACCINES**Example 13. Fusion Proteins of Fve and Allergen***Materials and methods*

5 Treatment of recombinant allergen or vaccination with naked DNA encoding a specific allergen has been shown previously to elevate allergen-specific Th1 immune response against Th2 immune reaction (Maecker et al., 2001). To enhance the effectiveness of immunotherapy or DNA vaccine therapy, we generated several fusion proteins consisting of the complete Fve molecule and the mature form of Blo t 5 or Der p 2
 10 allergen. Figure 16 shows the construction of seven fusion proteins of Fve and major house dust mite allergen from *Dermatophagoides ptenyssinus* and *Blomia tropicalis*

The fused cDNAs are successfully expressed in E coli (Figure 17) and the biological properties of the recombinant proteins are examined.

Results

15 The morphology of lymphocyte culture upon stimulation with three recombinant fusion proteins is photographed with inverted microscope (Figure 18A-C). Each of Bt5-Fve, Bt5-FveR27, GST-Dp2-FveR27 are able to increase the number of human PBMC (Figure 19A and 19B), to stimulate the proliferation of human lymphocytes (Figure 20), to polarize human CD8⁺ T cells (Figure 21), and to increase the production of IFN- γ (Th1
 20 response) and IL-10 (Tr response) (Figure 22).

A well-balanced vaccine that induces both Th1 and Tr immune response may be the most valuable and desirable. The Th1 response may very efficiently inhibit the development of Th2 cells via IFN- γ , leading to a life-long protective Th1 memory immune response. Allergen specific Tr cells may in turn dampen the anti-allergic Th1 immune 5 response, ensuring a well-balanced protective but nonpathological Th1 response. Allergen-Fve fusion proteins meet these criteria since they induce cytokine IL-10.

Thus, combining Fve protein with allergen in the form of a fusion protein may be used effectively to induce antigen-specific adjuvant effect that augment the Th1 and Tr responses, which in turn down-regulate the Th2 allergic responses.

10 To test the antigenicity of a Blo t 5-Fve fusion protein, competitive inhibition ELISA is performed using varying concentrations of proteins (GST, GST-Blo t5, GST-Fve, GST-Blo t5-Fve, GST-Fve-Blo t5, Blo t5-Fve). The results show that fusion protein Blo t 5-Fve, un-cleaved GST-Blo t5-Fve and GST-Fve-Blo t5 have lower IgE binding affinity compared to Blo t5 alone and un-cleaved GST-Blo t5 (Figure 23). The fusion 15 protein Blo t5-Fve inhibited IgE binding to a maximum of 70% whereas Blo t5 is able to inhibit the binding of antibody to GST-Bt5 to 100% at inhibitor concentration of 10 μ g/ml. Control GST and GST-Fve are not able to inhibit the binding of IgE to GST-Blo t5 (background levels). In summary, there is a reduction in the IgE binding affinity of Blo t5 when it is in the fusion forms of Blo t5-Fve, GST-Blo t5-Fve and GST-Fve-Blo t5 20 indicating that the antigenicity of Blo t5 with Fve in fusion forms is lowered.

Example 14. Allergen Conjugated to Fve

Beside the use of gene fusions to produce fusion proteins, protein-protein conjugation also provides a convenient and alternative choice to develop allergen vaccine.

25 To date, allergen conjugated adjuvants which have been reported include crystalline bacteria cell surface layer (S-layers) (Jahn-Schmid et al., 1996), CpG

oligodeoxynucleotides (CpG motifs) (Shirota et al., 2000), cholera toxin B subunit (CTB) (Rask et al., 2000), and *Brucella abortus* (Scharf et al., 2001).

Here we disclose Fve protein which is isolated from edible mushroom can also be an ideal adjuvant coupling to allergen vaccine. Poly-lactic acid (PLA) and polyethylene

- 5 glycol (PEG) are two materials which may be used to couple Fve and house dust mite allergen (Der p 2 or Blo t 5), although other materials will be evident to the skilled reader.

Particular cross-linking reagents which may be used to conjugate an allergen and immunomodulator, such as Fve, include N,N'-dicyclohexylcarbodiimide (DCC), N-

- 10 succinimidyl-S-acetyl-thioacetate (SATA), N-succinimidyl-3-(2-pyridyldithio)propionate (SPDP), ortho-phenylenedimaleimide (o-PDM), and sulfosuccinimidyl 4-(N-maleimidomethyl) cyclohexane-1-carboxylate (sulfo-SMCC). A chemical conjugation protocol which may be used is that provided in the Protein-Protein Crosslinking Kit (P6305) from Molecular Probes, Eugene, USA. Protocols for conjugation using SPDP are 15 disclosed in Clinical Experimental Allergy 30: 1024-1032, 2000 and European Journal of Immunology 28: 424-432, 1998.

For example, native Fve or recombinant Fve from *E coli* is conjugated with N-succinimidyl 3-(2-pyridyldithio) propionate (SPDP, Molecular Probes) as a bifunctional coupling reagent. The resulting Allergen-Fve conjugates are purified by gel filtration and 20 characterized for their allergenicity and adjuvanticity by *in vitro* and *in vivo* assays.

Example 15. Human Cytokine Assay in Purified CD4⁺ and CD8⁺ T Cell Subsets

Materials and Methods

To elucidate and identify subsets of human T lymphocytes responding to Fve stimulation, purified CD4⁺ T cells and CD8⁺ T cells from four human tonsillectomy 25 patients (subject 1, 6 yrs-old Chinese; subject 2, 16 yrs-old Indian; subject 3, 17 yrs-old Malay; subject 4, 27 yrs-old Malay) are stimulated with 20μg of Fve after AutoMACS

seperation. AutoMACS is an automated magnetic cell sorter from Miltenyi-Biotec, Germany. The differential cytokine production profiles of these subsets are determined by intracellular cytokines staining after 48 hours in vitro culture.

Results

5 *Fve Triggers Th1/Tc1 Cytokine Production in Human T Cells*

The human cytokines induction studies show that Fve stimulates the production of IL-2, IFN- γ , TNF- α , whereas IL-4 and IL-10 are nearly undetectable. In addition, purified CD4 $^+$ T cells produce higher levels of TNF- α than purified CD8 $^+$ T cells (CD4 $^+$ vs CD8 $^+$: 11.4% vs 2.5%), whereas purified CD8 $^+$ T cells produce higher levels of IFN- γ than purified CD4 $^+$ T cells (CD4 $^+$ vs CD8 $^+$: 3.6% vs 8.5%) upon Fve stimulation (Table 4). Therefore, the enrichment of CD8 $^+$ T cells seems to derive from a protein-cell direct interaction. Taken together, this data supported that Fve could trigger Th1/TC1 cytokines production in human T lymphocytes.

Intracellular Cytokines Scretion	Purified CD8 $^+$ T cells from human tonsil		Purified CD4 $^+$ T cells from human tonsil	
	None	Fve	None	Fve
IL-2	0.1%	0.6%	0.2%	6.8%
IL-4	0.1%	0.3%	0.1%	0.9%
IL-10	0.6%	0.5%	2.3%	0.9%
IFN- γ	0.1%	8.5%	0.6%	3.6%
TNF- α	0.2%	2.5%	0.4%	11.4%

Table 4. Cytokines profile of purified human T cells subsets

Example 16. Lymphocyte Aggregation Activity of Fve

Materials and Methods

Human CD4⁺ and CD8⁺ T cells subset are purified from AutoMACS (an automated magnetic cell sorter from Miltenyi-Biotec, Germany). The morphology of the cells is observed by light microscope at day 3.

Six human cell lines are also used for the cell aggregation study. Promyelocytic HL-60 cells, Jurkat-T cells, monocytic leukemia U937 cells, myeloid leukemia K562 cells, Raji B cells, natural killer NK-92 cells are cultured with native Fve protein with 2.5μg/ml, 5μg/ml, 10μg/ml, 20μg /ml and 40μg/ml, respectively. Cells aggregation is observed by inverted light microscopy after 24 hours.

Results

Fve induced aggregation of human CD4⁺ and CD8⁺ T cells subsets, HL-60, Jurkat-T cells, and NK-92 Cells

Human CD4⁺ and CD8⁺ T cells subset are purified from the tonsil of human subject. The aggregation of CD4⁺ and CD8⁺ T cells upon stimulation with 20μg of Fve protein is observed by confocal microscope at day 3 (photographed data not shown).

From the human cell line study, we found that Fve could induce HL-60 aggregation at low concentration of 2.5μg /ml. Jurkats-T cells and NK-92 also induced aggregation by Fve at concentration of 10μg /ml and 20μg /ml, respectively, where as U937, K562 and Raji didn't induce cell aggregation (Table 5). From the result, it seems that the level of cell aggregation correlates with the level of certain surface protein(s) expression in different cell lines. Promyelocytic cell line HL-60 seems to be an idea cell line to identify Fve receptor.

Human Cell Lines	Fve				
	2.5µg /ml	5µg /ml	10µg /ml	20µg /ml	40µg /ml
HL-60	+	+	+	+	+
Jurkat T	+/-	+/-	+	+	+
U937	-	-	-	-	+/-
K562	-	-	-	-	+/-
Raji	-	-	-	-	-
NK-92	-	-	+/-	+	+

Table 5. Cell aggregation activity of human cell lines

Example 17. *In vitro* Polarization of Human NK cells and CD8⁺ T Cells*Materials and Methods*

Human peripheral blood mononuclear cells (PBMC) from a healthy donor are

5 isolated as standard protocol (Coligan et al., 1998). The cells are then cultured in 24-well plates with native Fve (5µg /ml or 25µg /ml). At days 5 and 10, cell culture are stained with anti-CD4⁺ FITC, anti-CD8⁺ PE, anti-CD16⁺ PE plus anti-CD56⁺ PE monoclonal antibodies (Becton Dickinson), and analyzed by FACScan flow cytometry (Becton Dickinson).

10 Results

Sequential polarization of cells by Fve, NK cells and NKT cells are proportionally increased at day 5 whereas CD8⁺ T cells are increased at day 10

The results show a 10% increase of CD16⁺ and CD56⁺ double positive cells (Natural Killer cells) after stimulation with Fve protein for 5 days (Figure 24). In addition, 15 CD8⁺ T cells but not CD4⁺ cells are increased up to 35% after culturing for 10 days (Figure 25). This result showed that native Fve protein could stimulate both natural killer

cells and CD8⁺ T cells and the stimulation of these cells occurred sequentially with polarization of NK cells and CD8⁺ T cells peaked at day 5 and day 10, respectively.

The data also showed that cell culture consisted of 10% of CD3⁺CD16⁺CD56⁺ NKT cells after stimulation with 25µg /ml of native Fve protein (Figure 24E). This subset of cytotoxic NKT cells has a unique feature in that they mediate non-MHC-restricted cytotoxicity (Lanier et al., 1986).

Example 18. Up- Regulation of a Novel Subset of CD8⁺ T Cells (CD3⁺ CD8⁺ CD18^{+bright})

Materials and Methods

Repeated subcutaneous injection of IL-12 in patients with cancer resulted in the selective expansion of a unique subset of peripheral blood CD8⁺ T cells. This subset expressed high levels of CD18⁺ and up- regulated IL-12 receptor expression after IL-12 treatment in vivo. They appeared morphologically as large granular lymphocytes, increased high IFN-γ production and enhanced non-MHC-restricted cytolytic activity.

Thus, these T cells may play an important role in innate as well as acquired immunity to tumors and infectious pathogens.

To determine whether CD3⁺ CD8⁺ CD18^{+bright} T cells can be enriched by native Fve protein, human peripheral blood mononuclear cells (PBMC) from a healthy donor are isolated and cultured with 20µg /ml of native Fve protein. Cell culture are stained with anti-CD18 FITC, anti-CD8 PE, anti-CD3 PerCP monoclonal antibodies (Becton Dickinson) at day 5, and then analyzed by FACSCalibur flow cytometry (Becton Dickinson).

Results

Result showed that CD3⁺CD18^{+bright} T cells are increased from 8% to 31% of total cell population (Figure 26), and CD3⁺CD8^{+bright}CD18^{+bright} T cells are increased nearly three times, from 3.5% to 9% of the total cell population (Figure 27) after stimulation with

20 μ g /ml of native Fve protein. Furthermore, some CD18 $^+$ CD8 $^-$ cells started to differentiate into CD18 $^+$ CD8 $^{+dim}$ cells after stimulated with native Fve protein (Figure 27B). This data suggested that Fve protein from the golden needle mushroom has a potential ability to stimulate cellular immune responses directed against malignancies in
5 human.

Example 19. *In vivo* Lymphocyte Proliferation Assays

Materials and Methods

Since Fve protein can activate human NK cells and CD8 $^+$ T cells *in vitro*, we sought to determine whether Fve would enhance activation of these cells *in vivo*. Mouse
10 provides a good model system for such a study.

A group of three C57BL/6J mice are subcutaneously injected with 10 μ g, 50 μ g or 250 μ g Fve protein consecutively for three days, respectively. Another three BALB/cJ mice are treated with 125 μ g of Fve protein each for seven days by subcutaneous injection. For continuous BrdU labeling, mice are given 0.5mg/ml BrdU (Sigma) in the drinking water,
15 which is changed every 3 days and then each mouse received one intraperitoneal injection of 1mg of BrdU in PBS at 6 hours before being sacrificed. PBMC, lymph node and spleen are isolated and resuspended in 200ul of washing buffer (1xPBS containing 1% bovine calf serum), then stained with anti CD4 $^+$ -FITC, anti CD8 $^+$ -PE, anti CD19 $^+$ -PE or anti PanNK-PE monoclonal antibody (Pharmingen) for 30 minutes on ice. After two washings
20 with washing buffer, the samples are fixed with FACS Permeabilizing Solution (Becton Dickinson) for 16 hours. After that samples are treated with 50U DNase I (Sigma) for 1hr at room temperature. The cells are washed and stained with anti BrdU-FITC mAb (Becton Dickinson) in PBS for 30 minutes. 1-5 x 10 5 viable (forward and side scatter gated) PBMC, lymphocytes in lymph nodes or splenocytes per sample are analyzed with
25 FACScan (Becton Dickinson) and data are processed using the CellQuest software (Becton Dickinson).

Results

Fve induced NK cells and CD8⁺ T cells proliferation in vivo

FACScan analysis data showed that Fve could induce increased proliferation of NK cells and CD8⁺ T cells in a dose-dependent manner in C57BL/6J mice (Figure 28 and Figure 29). In contrast, CD4⁺ T cells and CD19⁺ B cells showed no significant increase (Figure 30 and Figure 31). Similar CD8⁺ T cell polarization is also seen in lymph nodes of C57BL/6J mice (Figure 32) and so the peripheral blood mononuclear cells (PBMC) of Balb/cJ mice that are subcutaneous injected for seven consecutive days with 125µg of Fve protein. The CD8⁺ versus CD4⁺ T cells ratio is significantly increased in each of the Fve-treated BALB/cJ mouse as compared to the naïve control (Figure 33). Data from the experiment are presented in Table 6 below.

Naïve Balb/cJ mouse	PBMC		CD8 ⁺ /CD4 ⁺ ratio
	CD4 ⁺ T cells	CD8 ⁺ T cells	
#1 None	40.3 %	15.7 %	0.389
#2 125µg nFve	40.2 %	26.2 %	0.651
#3 125µg nFve	40.7 %	21.8 %	0.535
#4 125µg nFve	33.3 %	19.6 %	0.588

Table 6. Data showing results of Figure 33.

In summary, for NK cells in spleen, 10µg Fve caused one fold increase proliferation. The proliferation increased to 5-6 fold when 50µg and 250µg of Fve protein is added. Similar finding is observed in CD8 positive T cells in spleen and lymph nodes. 250µg Fve protein caused 2-3 fold increase proliferation as compared to non-treated mouse. By contrast, Fve failed to stimulate CD4 positive T cells and has very mild stimulation to CD19 B cells (Table 7). Similar phenomenon is also seen in the peripheral blood mononuclear cells. The proportional of CD8 T cells increased up to 6-10% after 125µg of Fve protein are subcutaneous injected to Balb/cJ mice for seven days (Table 8).

These *in vivo* data are in concordance with those derived from *in vitro* studies, which clearly indicate that Fve induces selective polarization of NK cells and CD8⁺ T cells. Furthermore, these immunostimulatory effects of Fve are independent of the genetic background of mouse strains. Thus, Fve appears to be a potent immunostimulator for 5 cellular mediated immune response. Purified NK cells and CD8⁺ T cells will be used for future studies to examine the molecular and cellular basis for the polarization of cell subsets.

Naïve C57BL/6J mouse	Spleen				Lymph nodes
	BrdU incorporated NK cells	BrdU incorporated CD4 ⁺ T cells	BrdU incorporated CD8 ⁺ T cells	BrdU incorporated CD19 ⁺ B cells	BrdU incorporated CD8 ⁺ T cells
#1 None	0.63%	3.49 %	2.22 %	3.48 %	5.83 %
#2 10µg Fve	1.20 %	3.32 %	2.81 %	3.43 %	5.72 %
#3 50µg Fve	3.53 %	3.47 %	3.34 %	4.11 %	9.19 %
#4 250µg Fve	4.00 %	2.55 %	7.31 %	4.55 %	12.05 %

Table 7. *In vivo* stimulation of C57BL/6J mouse lymphocytes

Naïve Balb/cJ mouse	PBMC		
	CD4 ⁺ T cells	CD8 ⁺ T cells	CD8 ⁺ /CD4 ⁺ ratio
#1 None	40.3 %	15.7 %	0.389
#2 125µg Fve	40.2 %	26.2 %	0.651
#3 125µg Fve	40.7 %	21.8 %	0.535
#4 125µg Fve	33.3 %	19.6 %	0.588

Table 8. *In vivo* stimulation of Balb/cJ mouse lymphocytes

Example 20. *In vivo* Evaluation of the Potential Use of Fve for Immunotherapy of Solid Tumors

There are several approaches to treat cancer such as surgery; radiation therapy; given tumor cell arrested drugs; induced apoptosis in cancerous cells; inhibited 5 angiogenesis; elevated tumor recognition and specific killing ability of immune system to eliminate cancerous cells.

Previous data have indicated that Fve protein stimulate enhanced production of various cytokines, particularly IFN- γ , TNF- α and IL-2; induced polarization of natural killer cells and CD8 $^{+}$ T lymphocytes; and triggered a Th1/Tc1-like cellular-mediated 10 immune response. Each of these biological properties may contribute to suppression of tumor growth and to prevent the risk of cancer recurrence by inducing various forms of nonspecific or even specific immunity after surgery.

Malignant melanoma is a very common cancer in the western world. A subset of patient with metastatic melanoma can be successfully treated by the administration of 15 recombinant IL-2, sometimes given together with autologous melanoma-reactive lymphocytes that have been expanded ex vivo. Since melanocyte differentiation antigens, including MART-1/Melan-A, gp100, tyrosinase, TRP-1, and TRP-2, and cancer-testis antigens, including MAGE-3, BAGE, GAGE, NY-ESO-1, are recognized by human T lymphocytes, therefore they become the attractive targets for melanoma vaccines. 20 However, from an immunological point of view, these melanocytes differentiation antigens and cancer-testis antigens are “self” antigens. It may induce central or peripheral tolerance, and thus potentially hampering the induction of powerful anti-melanoma immune responses. Therefore, induction of a strong tumor specific immunity with an immunopotentiator or novel adjuvant could be a useful treatment strategy to overcome 25 immune ignorance and tolerance.

In order to investigate the anti-tumor effect of Fve, C57BL/6J mice are subcutaneously inoculated either with T cell lymphoma EL4 or melanoma B16-F1, the

later is a well established and widely used tumor model for which treatment is notoriously difficult. The tumor growth and survival rate of mice are monitored.

Materials and Methods

Construction of pCIneo-fve and pDisplay-fve recombinant plasmid DNA

5 The pCIneo vector is designed for high level and constitutive expression of cloned DNA inserts in mammalian cells (Figure 34A). Fve DNA is amplified from pGEX-fve and subcloned into the Xho I and EcoR I restriction enzyme cutting sites of pCIneo vector. The pCIneo-fve is used for priming the immune response by intramuscular injection.

10 The pDisplay vector is a mammalian expression vector that is designed to target and to display recombinant proteins to the surface of mammalian cells (Figure 34B). Fusion DNA of Fve and murine Ig kappa chain V-J2-C signal peptide without hemagglutinin A epitope is generated by recombinant PCR and subcloned into the EcoR I and Pst I restriction enzyme cutting sites of pDisplay vector. The Fve protein expressed from the pDisplay-fve acts as triggering signal for immune system and recruiting T 15 lymphocytes to recognize tumor cells.

Transfection of B16-F1 cells with pDisplay-fve

10 The murine melanoma cells B16-F1 is purchased from ATCC, USA. Tumor cells are grown in DMEM supplemented with 10% FBS in 5% CO₂. Cells in the exponential growth phase within four passages are used in this investigation. To obtain stable transfectants, endotoxin free plasmid pDisplay-fve is mixed with polyfect transfection reagent (QIAGEN, Germany) and transfected into B16-F1 cells. Colonies resistant to G418 (Geneticin, GIBCO BRL) at 1000µg /ml for 25-30 days are isolated and designated as B16-Fve. The control B16-F1 cells which are transfected with pDisplay vector alone are designated as B16-vec.

EL4 protection assay

Six to eight weeks old C57BL/6J mice are inoculated with 8×10^6 EL4 cells. Tumor formation is observed at day 3. 100 μ g of pCIneo-fve recombinant plasmid DNA is intramuscularly injected into the tibialis muscle at day 0 and day 7. 20 μ g of Fve protein is given by subcutaneous injection surrounding the tumor site at day 5, 7, 9, 11, 13, 15, and 18, respectively. The diameters of tumors are measured with a caliper and tumor volume is calculated by long diameter times short diameter. Finally the survival rate is recorded.

DNA vaccination and B16-F1 tumor protection experiments

Endotoxin free pCIneo and pCIneo-fve are purified from the QIAGEN plasmid DNA extraction and purification kits. 100 μ g of pCIneo-fve is intramuscularly injected into the tibialis muscle of C57BL/6 mice at day -30 and day -1. Muscles are pulsed with Electro Square Porator ECM830 (BTX, Genetronics, USA) equipped with a two needle array electrode after DNA injection. Mice are inoculated with 5×10^5 B16-F1 cells. Small tumor nodule developed at day 3. 50 μ g of Fve protein is given by subcutaneous injection surrounding the tumor site at day 4, 7, 9, and 12, respectively.

Experimental lung metastasis

B16-F1 cells are trypsinized from monolayer cultures, counted and spun down at 1,200 rpm for 5 min and resuspended with DMEM. Five C57BL/6 syngenic 6-week-old female mice are intravenously injected with 2×10^4 of B16-F1 melanoma cells in a final volume of 120 μ l. About 4 weeks after injection, tumor nodules are established in lung. Mice are kept until they died to assess survival.

Example 21. Prolonged Survival Rate of Tumor-Inoculated Mice Receiving with Fve Gene and Protein

Our results show that tumor established mice that received pCIneo-fve DNA and Fve protein had shown a reduction of T cell lymphoma growth rate (Figure 35) and an

extension of survival time (Figure 36). Similar results are also seen in melanoma B16-F1 inoculated C57BL/6J mice (Figure 37).

These data indicate that Fve induces some protection against the solid EL4 tumor and B16-F1 melanoma, suggesting that Fve could be a potential candidate molecule for the 5 development of the immunotherapeutic reagents for treatment of some cancers. The results also show that DNA vaccine-mediated treatment using the gene of Fve can be further exploited for effective cancer treatment. Nowadays, DNA vaccination is being explored as a potential strategy for combating cancer. However, tumor antigens are often weak and the immune system of patients may be compromised. Like the concept of allergen-Fve fusion protein, fusion of Fve to specific tumor antigen may an effective way to activate protective anti-tumor immune response. Genetic immunization with chimeric gene encoding Fve and tumor antigen may augment and direct immune attack on a range of target tumor antigens. 10 15

Example 22. Life Span in Solid Tumor Model is Extended in Fve Transfected

In previous study, we have proved that using Fve plasmid DNA primed in muscle 15 and Fve protein boosted in tumor region could initiate anti-tumor immune response and thus prolong the survival time of tumor-inoculated mice. Instead of injection Fve surrounding the tumor, we specifically targeted Fve gene into tumor cells and tried to create an inducible-antigenic tumor for cancer therapy. This genetically modified tumor 20 cells may provide signals for antigen presenting cells and both helper and cytotoxic T

To determine whether introduction of the Fve gene into malignant cells would result in enhanced tumor recognition ability via Fve display and lead to extended survival rate in solid tumor experiment. Recombinant plasmid DNA pDisplay-fve is transfected into wild type B16-F1 mouse melanoma and then G418 resistant colonies are selected. 25 Five female of C57BL/6J mice are inoculated with 5×10^4 of B16-Fve transfected. The antigenicity of B16-vec and B16-Fve transfecteds are compared through the life span of two groups of tumor-inoculated mice.

Result demonstrated that artificially expressed Fve on the surface of B16-F1 mouse melanoma extended survival rate as compared to B16-vec inoculated mice (Figure 38). We propose that the characteristics of highly antigenicity and lymphocytes mitogenicity of Fve may elevate immune function to fight against tumor when it displayed on the surface of melanoma. Therefore, Fve may use as immune response activator and enhancer especially for those poorly recognized and low immunogenic tumor, which escaped from cancer surveillance and immune clearance by altering immune recognition and modulating cytotoxic response.

Example 23. Fve DNA Vaccination Retards Tumor Progression

Cancer vaccines are designed to prevent and treat cancer. In general, research has shown that the most effective anti-tumor immune responses are achieved by stimulating T cells, which can recognize and kill tumor cells directly. Most current cancer vaccines try to activate T cells directly, try to enlist APCs to activate T cells, or both. Some novel ways in which researchers are attempting to better activate T cells are: (1) Altering tumor cells so molecules that are normally only express on APCs are now express on the tumor cell. These molecules are capable of giving T cells a stronger activating signal than the original tumor cells. (2) Testing more cytokines and adjuvants to determine which are best candidates for recruiting APCs to areas where they are needed. (3) Using dendritic cells and other APCs fused with tumor cells as the cancer vaccines. These cells go into the body carrying tumor antigen and ready to activate T cells. Early cancer vaccine clinical trials involved mainly patients with melanoma. Currently, cancer vaccines are also being tested in the treatment of many other types of cancer, including prostate cancer, breast cancer, colon cancer, and lymphoma.

Here, we accessed tumor immunity that stimulated by recombinant Fve DNA vaccination alone and the combination of Fve DNA vaccination and Fve-transduced tumor cells. C57BL/6J mice are separated into three groups and each group consisted of ten mice. Mice are inoculated either with 5×10^4 of B16-Fve or B16-vec tumor transfectants in the dorsal back. Tumor formation is observed at day 5-7. 100 μ g of pCIneo-fve plasmid DNA

is intramuscularly injected at the right and left tribilis muscle of C57BL/6J at day -77, day -35 and day -21. Mice are subcutaneously injected with 5×10^4 of B16-Fve transfectant and B16-vec transfectant at day 0, respectively. 100 μ g of pCIneo plasmid DNA is administered following similar experimental procedure and mice are subcutaneously injected with 5×10^4 of B16-vec transfectant as negative control. The fatal rate of mice are recorded and data are presented as survival curves.

Result showed that Fve DNA vaccination contained certain degree of tumor protection (Green line in Figure 39) as compared with vector DNA vaccination (Blue line in Figure 39). In addition, the combination of Fve DNA vaccination and B16-Fve transfectant exerted a stronger tumor protection effect (Red line in Figure 39). Based on the result, we propose Fve is a novel protein to activate T cells directly. This protein is capable of giving T cells a strong activating signal when it is displayed on the surface of poorly immunogenic tumor cells. Therefore, the phenomenon of extended survival rate is observed in the experimental tumor-inoculated mice.

In future, the adjuvant effect of fusion proteins between Fve and tumor antigens to enhance tumor immunity will be determined. In particular, DNA fusion vaccine strategy, whereby target tumor antigen is genetically linked to immunostimulatory molecules such as Fve, is currently being explored. The introduction of fusion gene encoding tumor-associated antigen with Fve into antigen-presenting cells hold considerable promise for the treatment of patients with cancer. The ease of DNA manipulation has allowed incorporation of a wide variety of molecules able to promote antigen uptake, processing and presentation by professional antigen-presenting cells, to provide critical CD4 $^{+}$ T-cell help and to activate more effective immune effector pathways (Zhu and Stevenson 2002). The concept of DNA fusion vaccine strategy is particularly important for cancer vaccines to increase their immunogenicity and to overcome tolerance.

Example 24. Fve Extends the Survival Rate of Lung Metastatic Mice

2x10⁴ of B16-F1 melanoma cells is delivered to C57BL/6J via tail vein injection. The effect of combination of distill water and DNA vector pCIneo versus Fve protein and plasmid DNA pCIneo-fve administration on survival after the establishment of lung
5 metastasis is determined. Survival extended in both metastatic experimental groups undergoing Fve protein orally primed and DNA intramuscularly boosted strategy.

Two groups of five C57BL/6J mice are given with 10mg/ml of Fve protein in the drinking water at days -35, -28 and -21, and each water providing is maintained consecutively for one week. Mice are intravenously injected with 2x10⁴ of B16-F1 (wild type) melanoma cells at day 0. One week after, mice are intramuscularly injected with 100µg of pCIneo-fve plasmid DNA into the right and left tribilis muscle, respectively. The mixture of 5x10⁴ of B16-Fve cells lysate plus 10µg of Fve protein (Red line in Figure 40) or 10µg of Fve protein alone (Green line in Figure 40) are subcutaneously injected into mice at the following three weeks. Negative control group of mice received same amount
10 of 1xPBS in the drinking water, intravenously injected with 2x10⁴ of B16-F1 melanoma cells, followed by intramuscularly injected with plasmid DNA vector pCIneo, and finally
15 subcutaneously injected with B16-vec cells lysate plus 1xPBS (Blue line in Figure 40).

Results showed that the strategy of orally primed with Fve protein before tumor introduced into the lung and intramuscularly boosted the immune response with the
20 plasmid DNA pCIneo-fve after tumor established in lung could extend the survival rate of mice as compared with the control group (Figure 40). This data provided another evidence suggesting Fve could augment anti-tumor immune response against developing or metastatic tumor cells. The inhibition of B16-F1 melanoma experimental lung metastasis by Fve may go through induction of IFN-γ, TNF-α and activation of anti-tumor host
25 mechanisms. IFN-γ^{-/-} and TNF-α^{-/-} gene knockout mice and in vivo depletions of CD4⁺, CD8⁺, or NK1.1⁺ cells may provide supportive evidence to this phenomenon.

Example 25. Global Gene Expression Profiling of Human T Cells and NK Cells After Activation with Fve

The invention of microarray technology allows the simultaneous monitoring of the transcriptional behavior of thousands of genes. This technology has been repeatedly shown to be useful in the analysis of the response of a variety of cellular systems to stimuli, in the classification of human cancer, and in the analysis of animal models of human disease (Churchill 2002; Slonim 2002; van Berkum and Holstege, 2001). To characterize the transcriptional profile of Fve, we analyzed gene expression patterns in T and NK cells from either healthy donor or human cell lines stimulation with Fve by using oligonucleotide microarrays and compared them with gene expression patterns in non-stimulation cells. In future, protein microarray assays would also be used to study protein-protein interactions on a genome-wide scale (Templin et al., 2002; Zhu et al., 2001).

*Materials and Methods**Cells collection and total RNA purification*

Peripheral blood mononuclear cells (PBMC) are collected from healthy donors. CD8-positive T lymphocytes and natural killer cells isolation are performed by immunomagnetic bead selection with monoclonal mouse anti-human CD8 antibodies and monoclonal mouse anti-human CD56 antibodies using the AutoMACS automated separation system (Miltenyi-Biotec, Germany). CD8-positive T cells and CD56-positive natural killer cells purity of more than 94% and 88% homogeneity are confirmed by two-color flow cytometry using CD3⁺/CD8⁺ and CD56⁺ criteria (Becton Dickinson, USA). Human T cell lines (Jurkat T cell, MOLT-4) and NK cell line (NK-92) are grown as recommended (ATCC, USA). Cells are stimulated with Fve and total RNA is isolated with RNeasy Mini Kit (Qiagen, Germany) after 2 and 48 hours, respectively.

Preparation of labeled complementary RNA and hybridization to high-density microarray

Double-stranded complementary DNA (cDNA) and biotinylated complementary RNA (cRNA) are synthesized from total RNA and hybridized to human GeneChip

microarray (Affymetrix, USA), which are washed and scanned according to procedures developed by the manufacturer. The arrays are scanned using laser scanner and visualized using Affymetrix 3.3 software (Affymetrix).

GeneChip data analysis

- 5 Differentially expressed genes are analysed by functional assays

Example 26. X-Ray Crystallographic Study of Fve: Materials and Methods

The three dimensional structural of Fve provides a good basis for the understanding of protein functions, immunomodulations and therapeutic applications in allergy and other diseases. We have crystallized the well-diffracting crystals of Fve and
10 show that it diffracts to 1.4 Å resolution when exposed to synchrotron radiation.

This and the following Examples describe a 1.6 Å° x-ray structure of Fve, determined by Single Anomalous Diffraction (SAD) using the anomalous signal of bromide ions present in the crystal for phasing. Fve represents a novel structure, wherein each monomer consists of an N-terminal α-helix followed by an immunoglobulin fold
15 (beta-sandwich). The structure strongly suggests that dimerization, critical for the activity of FIP proteins, occurs by 3-D domain swapping of these helices and is stabilized predominantly by strong hydrophobic interactions between them.

Crystallization

Fve protein is obtained as described above. It is concentrated to 4 mg/ml in 10 mM
20 Tris-HCl pH 7.5. Initial crystallization screening is done by the sparse matrix crystallization screening kit 1 & 2 from Hampton Research (Jancarik and Kim, 1991; Cudney, *et al.*, 1994). All the screening and crystals growth are accomplished by hanging drops vapor diffusion method at 21°C in VDX multi-well plates with 650 µl reservoir solutions. Drops consisting of 4 µl precipitant buffer from reservoirs and 4 µl protein
25 sample (4 mg/ml) are equilibrated over the well solution for one week.

After extensive screening, plates-like crystals are obtained at two different low salt conditions: (1) 30% PEG 4000, 0.1 M Tris-HCl pH 8.5, 0.2 M MgCl₂; (2) 30% PEG 4000, 0.1 M Tris-HCl pH 8.5, 0.2 M NaOAc. 3D cubic-shaped and octahedral crystals also appeared after 3 days at two different high salt conditions: (1) 2.0 M (NH₄)₂SO₄, 0.1 M Tris-HCl pH 8.5; (2) 2% PEG 400; 0.1 M Na Hepes pH 7.5, 2.0 M (NH₄)₂SO₄. To optimize the crystallization condition, combinations of varied protein and salt concentrations, different molecular weights of PEG, and different pH are screened.

The best crystals formed at the high salt condition is optimized to 2.5% PEG 400, 2.0 M (NH₄)₂SO₄, 0.1 M Tris-HCl pH 8.5 at 21°C. They grew to the approximate dimensions of 1.0 × 0.9 × 0.5 mm within five days. The micrographs of Fve crystals are captured by inverted light microscope (Figure 41).

High resolution protein crystals are therefore grown by vapor diffusion from hanging drop at 2.0% PEG 400, 2.0 M (NH₄)₂SO₄, 0.1 M Tris-Base, pH 8.5 for 1-2 weeks. Heavy atom derivatives are prepared by soaking the crystals in mother liquor containing 25% glycerol and 1M NaBr. The crystals are flash-frozen at 100 K after a 1-min soak in the heavy atom (Br) solution. SAD data from a derivatized crystal are collected at the National Synchrotron Light Source (NSLS) beam line X12C) at one wavelength (****) around the Br absorption edge. The crystal diffracted to 1.7 Å.

X-ray analysis

The X-ray diffraction intensities from Fve crystals are measured at 100 K on beamline BL9-2 from the Stanford Synchrotron Radiation Laboratory facility with ADSC Quantum-315 CCD detector. Data are collected at a wavelength of 1.07 Å. All the data are processed by MOSFLM (Leslie, 1992) and X-ray intensities are scaled with SCALA (CCP4, 1994). Well-ordered diffraction data at 1.4 Å resolution are collected from larger crystals (Figure 42).

Analysis of the collected data (Table 9) indicated that Fve crystals belong to the tetragonal space group $P4_32_12$ with unit cell dimensions of $a = b = 96.92$ Å, $c = 61.42$ Å.

The Matthews parameter (V_M) of these crystals is 2.84 \AA^3 per Da and thus the solvent content is 56.37% assuming two molecules of Fve per asymmetric unit (Matthews, 1968). A total of 344079 observations are obtained at 1.4 \AA resolution giving approximate 56993 unique reflections (99.7% complete, $R_{\text{merge}} = 0.047$).

X-ray source, beamline	SSRL, BL9-2
Wavelength	1.07 \AA
Detector distance	99.97mm, Q-315 CCD Detector
Cell angles (°)	90.00, 90.00, 90.00
Unit cell dimensions (\AA)	96.92, 96.92, 61.42
Space group	$P4_32_12$
Number of molecules per ASU	2
Number of observed reflections	344079
Number of unique reflections	56993
Solvent (%)	56.37
$V_M (\text{\AA}^3 \text{Da}^{-1})$	2.84
Resolution range (\AA)	33.5-1.4
Average $I/\sigma(I)$	10.1
R_{merge} ^a	0.047
Completeness (%)	99.7

5 ^a $R_{\text{merge}} = \sum |I_i - <I>| / \sum I_i$, where I_i is the mean intensity of symmetry-related measurements of this reflection.

Table 9. Data Collection and Statistics of Fve Crystal

Data Processing

The SAD data are processed and scaled using DENZO and SCALEPACK from the
10 HKL2000 suite of programs (Otwinowski and Minor, 1997).

The crystal of Fve belongs to the tetragonal space group P43212 and has unit cell dimensions $a = b = 97.12$, $c = 61.41$ and $\alpha = \beta = \gamma = 90.0$. All of the bromine heavy atom positions are located and refined by the program SOLVE at 1.7 \AA (Terwilliger and Berendzen, 1999) and solvent flattened map is calculated using RESOLVE (Terwilliger, 2001). The resulting electron density map reveals secondary structure elements and side chains. In principle, it is possible to build an initial model by standard protein map-tracing methods. However, the phases obtained from RESOLVE are directly used in ARP/wARP (Morris et al., 2001) for automated main chains tracing, result in 4 continuous fragments

that contained 97% of model. The rest of the model and side chains are fitted manually using XtalView (McRee, 1999). The refinement is carried out in REFMAC 5 (Murshudov et al., 1999) using resolution range 30.02 - 1.7 and water molecules are picked up by ARP/WARP later in the refinement.

5 In chain A, C-terminal residue 114 is modeled as Ala residue, whereas in chain B, C-terminal residue 113 and 114 are omitted from the final model, due to the poor interpretable density. The quality of the final model is verified with PROCHECK (Laskowski et al., 1993). However, the Ramachandran plot shows that Lys 14 in both A and B chains is in the disallowed region, although this residue fits very well in the 2fo-fc map.
 10

Example 27. X-Ray Crystallographic Study of Fve: Results

The crystal structure is solved by single anomalous scattering using Br as the heavy-atom, and is refined to a resolution of 1.7 Å. The atomic coordinates are presented in Appendix C.

15 In total, two chains with a total of 226 residues, 16 bromine atoms and 136 solvent molecules are built into a high quality electron density map. Fve comprises almost exclusively of β-sheet structure with an Ig-like fold, which is formed by seven major antiparallel β-strands arranged into two sheets of four (D, E, H and I) and three (B, C and F) strands packed again each other. The N-terminal domain is composed of a α-helix
 20 which spans a length of 12 residues from Ala2 to Val13 and a β-sheet (A). The N-terminal serine residue is blocked by an acetyl group the density of which is also observed. Six loops connect the two main β-sheets and one loop connects the N-terminal domain with β-sheet structure. The loop between the β-sheets F and H contains a short β-strand and a 3₁₀ helix.

25 The structure of Fve (Figure 43) reveals that exists as a dimer. This is corroborated experimentally by running Fve on a gel filtration column against standard molecular

weight markers (data not shown). From the structure, there are two extended regions of subunit-subunit interactions: between the two N-terminal α -helical regions (residues 2 to 13) and the β -stranded region (A and A').

The buried side chains of the α -helical regions form a hydrophobic core (Figure 5 44A), containing residues Ala 2, Leu 5, Leu 9 and Val 13 whereas the side chains of β -strand (A and A') make inter-subunit hydrogen bonds (Figure 44B). These hydrophobic interactions and hydrogen bonds are responsible for dimer formation. The two monomers, A and B chains, of Fve can be closely superimposed: the RMSD between corresponding C_α positions of 112 residues is 0.29 Å (Figure 44C).

10 *Domain Swapping*

Domain swapping is a very efficient method of forming oligomers since the interactions within the monomer are reused in the dimer. There is thus no need to evolve a new site on the surface which in one monomer mutually recognizes the corresponding site on the other monomer, since in the domain swapped dimer the recognition requirement has 15 already largely been accounted for (Bennett et al., 1995).

Domain-swapped proteins have a C-interface generally with many specifics interaction, formed between domains linked by a hinge loop (Bennett et al., 1995). In p13suc1, two proline residues, located in the hinge region, have been shown to be essential and control the domain-swapping process (Rousseau et al., 2001).

20 As shown in Figure 45A, half of the dimer of Fve contains one N-terminal helix, forming a C-interface with hydrophobic core, which is linked to rest of its subunit by a hinge loop, stretching from residue Val 13 to Pro 22. Furthermore, Fve contains a proline residue at the end of the hinge region, which could adopt alternative conformation in the dimer by releasing the tension in the monomer. These observations suggest that domain-25 swapping could be the mechanism for forming dimer protein from its monomer. The monomer is modeled (Figure 45B).

Structural Similarity with Other Proteins

Fve has no sequence homology to any other non-FIP proteins. However, a search for similar structure in the DALI database (Holm and Sander, 1993) reveals that the protein has a significantly similar fold to 140 proteins but none with the significant 5 sequence similarity to Fve. Among 140 similar fold protein, fibronectin type III family emerged with less topology diversity to Fve β -sandwich fold: the heparin and integrin binding segment of human fibronectin (FN12-15; PDB entry 1FNH), the fragment of human fibronectin type III repeat (FN7-10; 1FNF), The p40 domain of human interleukin-12 (p40; 1F42) and the antibody a6 fragment interferon-gamma receptor alpha chain 10 (IFN γ R1-108; 1JRH). An alignment of FN12-15, FN7-10, p40, IFN γ R1-108 and Fve on the basis of structural similarity shows topology diversity in the range 11-17, calculated by Topp program (Lu, 1996) (Table 10).

	Name	PDB ID	Z-Score	RM SD	Length of aligned segment	Topological Diversity	Superfamily (Family)	Species
1	interleukin-4 receptor alpha chain fragment: b:1-96	1ilar-B	5.8	3	78	8.5	Fn III (FNIII)	<i>Homo sapiens</i>
2	mhc class ii i-ak: a:82-181	1lak-A	5.8	4.7	83	18.6	Ig (C1)	<i>Mus musculus</i>
3	mhc class ii i-ak: b:93-190	1iak-B	5.6	3.5	74	17.8	Ig (C1)	<i>Mus musculus</i>
4	igg2a intact antibody - mab23, kappa L chain: a:1-108	1igt-B	5.5	3.8	86	18.4	Ig (V)	<i>Mus musculus</i>
5	class ii histocompatibility antigen, HLA-DM: a:94-196	1hdm-B	5.3	4.7	82	18.4	Ig (C1)	<i>Homo sapiens</i>
6	fibronectin fragment, heparin & integrin binding segment: a:93-182	1fnh-A	5.3	3	73	11.1	Fn III (FNIII)	<i>Homo sapiens</i>
7	ch3 domain of mak33 antibody fragment:chain a	1cqk-A	5.3	3.3	76	18.5	Ig (C1)	<i>Mus musculus</i>
8	CD1, beta2-microglobulin and alpha-3 domain: d	1cid	5.3	2.8	76	17.8	Ig (V)	<i>Rattus rattus</i>
9	fibronectin fragment, ED-B domain:chain a	2fnb-A	5.2	3.9	72	17	Fn III (FNIII)	<i>Homo sapiens</i>
10	hiv-1 gag peptide: a:182-276	1agd-A	5.2	3.8	84	20.1	Ig (C1)	<i>Homo sapiens</i>
11	igg1 antibody 32c2 fragment: a:1-110	32c2-A	5.1	5.6	80	19.4	Ig (V)	<i>Mus musculus</i>
12	fibronectin repeat 7: 1142-1235	1fnf	5.1	2.7	71	10.8	Fn III (FNIII)	<i>Homo sapiens</i>
13	interleukin-12 beta chain fragment: a:88-211	1f42-A	5.1	2.8	70	12.8	Fn III (FNIII)	<i>Homo sapiens</i>
14	Mutant growth hormone receptor fragment: b:131-236	1axi-B	5.1	3.2	72	14.7	Fn III (FNIII)	<i>Homo sapiens</i>

Table 10

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Each of the applications and patents mentioned in this document, and each document cited or referenced in each of the above applications and patents, including

5 during the prosecution of each of the applications and patents ("application cited documents") and any manufacturer's instructions or catalogues for any products cited or mentioned in each of the applications and patents and in any of the application cited documents, are hereby incorporated herein by reference. Furthermore, all documents cited
in this text, and all documents cited or referenced in documents cited in this text, and any manufacturer's instructions or catalogues for any products cited or mentioned in this text,
are hereby incorporated herein by reference.

10 Various modifications and variations of the described methods and system of the invention will be apparent to those skilled in the art without departing from the scope and spirit of the invention. Although the invention has been described in connection with specific preferred embodiments, it should be understood that the invention as claimed should not be unduly limited to such specific embodiments. Indeed, various modifications of the described modes for carrying out the invention which are obvious to those skilled in molecular biology or related fields are intended to be within the scope of the claims.

APPENDIX A: SEQUENCES

Fve is isolated from Golden Needle Mushroom (*Flammulina velutipes*).

ORGANISM: *Flammulina velutipes*. Eukaryota; Fungi; Basidiomycota; Hymenomycetes; Agaricales; Tricholomataceae; *Flammulina*.

5 ***Fve (Wild type)***

```

ATGTCCGCCACGTCGCTCACCTCCAGCTTGCCTACTTGGTGAAGAAGATCGACTTCGAC
TACACCCCCAACCTGGGGCCGTGGTACCCCAAGCAGCTACATCGACAACCTTACCTTCCC
AAGGTTCTCACCGACAAAAAAACTCGTACCGCGTGTGGTCAATGGCTCTGACCTTGGC
GTCGAGTCCAACCTCGCAGTGACACCGTCCGGTGGCAGACCATCAACTCCTCCAGTAC
10      AACAAAGGGGTATGGTGTGCGGGACACCAAAACGATTCAAGTTTCGTTGTCAATTCCAGAT
ACCGGCAACTCGGAGGAGTACATCATCGCTGAGTGGAAAGAAGACTTGA
msatsltfqlaylvkkidfdytpnwgrgtpssyidnltfpkv1tdkkysyrvvvngsdlg
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20      ACG/ATT/CAA/GTT/TTC/GTT/GTC/ATT/CCA/GAT/ACC/GGC/AAC/TCG/GAG/
GAG/TAC/ATC/ATC/GCT/GAG/TGG/AAG/AAG/ACT/TGA

```

A Fve (Wild type) sequence may also comprise a sequence as set out above, but lacking the initial methionine (M) in the amino acid sequence, or lacking the initial ATG in the nucleic acid sequence.

25 ***GST-Fve (Wild type) Nucleotide Sequence***

```

ATGTCCCCTATACTAGGTTATTGGAAAATTAAGGGCCTTGTGCAACCCACTCGACTTCTT
TTGGAATATCTTGAAGAAAAATATGAAGAGCATTGTATGAGCGCGATGAAGGTGATAAAA
TGGCGAAACAAAAAGTTGAATTGGGTTGGAGTTCCCAATCTCCTTATTATATTGAT
GGTGATGTTAAATTAAACACAGTCTATGGCCATCATACGTTATATAGCTGACAAGCACAAC
30      ATGTTGGGTGGTTGTCCAAAAGAGCGTGCAGAGATTCAATGTTGAAGGAGCGGTTTG
GATATTAGATA CGGTGTTCGAGAATTGCAATATAGTAAAGACTTGAAAACTCTCAAAGTT
GATTTCTTAGCAAGCTACCTGAAATGCTGAAAATGTTGAAGATCGTTATGTCAATAAA
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GTTGTTTATACATGGACCAATGTGCCTGGATGCGTTCCAAAATTAGTTGTTTAAA
35      AAACGTATTGAAGCTATCCCACAAATTGATAAGTACTTGAATCCAGCAAGTATATAGCA
TGGCCTTGCAGGGCTGGCAAGGCCACGTTGGTGGTGGCGACCATCCTCCAAAATCGGAT
CTGGAAGTTCTGTTCCAGGGGCCCTGGGATCCTCCGCCACGTCGCTCACCTCCAGCTT
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AGCAGCTACATCGACAA CCTTACCTTCCCAAGGTTCTCACCGACAAAAAAACTCGTAC
40      CGCGTCGTGGTCAATGGCTCTGACCTTGGCGTCAGTCCAAC TCGCAGTGACACCGTCC

```

GGTGGGCAGACCATCAACTCCTCCAGTACAACAAGGGGTATGGTGTGCGGGACACCAAA
 ACGATTCAAGTTCTGTCATTCCAGATACCGGCAACTCGGAGGAGTACATCATCGCT
 GAGTGGAAAGAAGACTTGA

5 *GST-Fve (Wild type) Amino Acid Sequence*

MSPILGYWKIKGLVQPTRLLLEYLEEKYEEHLYERDEGDKWRNKKFELGLEFPNLPLYID
 GDVKLTQSMAIIRYIADKHNMLGGCPKERAEISMLEGAVLDIYGVSRAYSKDFETLK
 DFLSKLPEMLKMFEDRLCHKTYLNGDHVTPDFMLYDALDVLYMDPMCLDAFPKLVCFK
 KRIEAIHQIDKYLKSSKYIAWPLQGWQATEFGGDHPPKSDLEVLFQGPLGSSATSLTFQL
 10 AYLVKKIDFDYTBNWGRGTPSSYIDNLTFPKVLTDDKKYSYRVVVNGSDLGVESNFAVTPS
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FVE DELETION MUTANTS

Fve D6-18

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 C/AAC/TTC/GCA/GTG/ACA/CCG/TCC/GGT/GGG/CAG/ACC/ATC/AAC/TTC/CTC/CAG/
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 T/GTC/ATT/CCA/GAT/ACC/GGC/AAC/TCG/GAG/GAG/TAC/ATC/ATC/GCT/GAG/TGG/
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Fve D19-33

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C/ATT/CCA/GAT/ACC/GGC/AAC/TCG/GAG/GAG/TAC/ATC/ATC/GCT/GAG/TGG/AAG/
AAG/ACT/TGA

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Fve p55-100

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FVE MUTANTS WITH SINGLE AMINO ACID SUBSTITUTIONS*FveR27A*

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FveG28A

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 40 A
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FveT29A

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avtpsggqtinflqynkgvgadtktiqvfvviptgnseeyiaewkkt

FUSION PROTEINS OF MAJOR HOUSE DUST MITE ALLERGEN (BLT 5 OR DER P 2) AND 10 Fungal Immunomodulatory Protein FvE

Blo t 5-Fve (two-in-one chimeric wild type)

15 caagagcacaaggccaaagaaggatgattccgaaacgaattcgatcaacttgttgcgaaacaggca
aaccatgtatcgaaaagg'gagaacatcaattgcttacttgcaacacccaactcgacgaattgaat
gaaaacaagagcaaggaattgcaagagaaaaatcattcgagaacttgcgttgcgcattgatgc
gaaggagccccaaaggagctttggAACGTGAATTGAAAGCAGACTGATCTTAACATTGGAAACGATTG
aactacgaagaggctcaaactctcgcaagatcttgcgttgcgttgcgttgcgcattgatgc
gtgaaggatattccaaacccaaTCCGCCACGTCGCTCACCTCAGCTGCCTACTTGTTGAAGAAG
ATCGACTCGACTACACCCCCAActGGGGCCGTGGTACCCCAAGCAGCTACATCGACAACTTACC
TTCACCAAGGTTCTCACCGACAAAAAAACTCGTACCGCGTGTGGTCAATGGCTCTGACCTTGGC
GTCGAGTCCAActTCGCACTGACACCGTCCGGTGGCAGACCATCAACTTCCTCCAGTACAACAAAG
GGGTATGGTGTGCGGGACACCAAAACGATTCAAGTTTCGTTGTCAATTCCAGATACCGGCAACTCG
GAGGAGTACATCATCGCTGAGTGGAAAGAAGACTTGA
QEHKPKKKDDFRNEFDHLLIEQANHAIKEKGHEQQLYLQHQLDELNENKSKELEQEKKIRELDVVCAMI
EGAQGALERELKRTDLNILERFNYEEAQTLSKILLKDLKETEQKVVDIQTQsatsltfqlaylvkk
idfdytpnwgrgtppsyidnltfpkvltdkkyssyrvvvngsdlgvesnfavtpsggqtinflqynk
gygvadtktiqvfvvipdtgnseeyjiaewkkt

Blo t 5-FveR27A (two-in-one chimeric mutant)

30 caagagcacaaggccaaagaaggatgttccgaaacgaattcgatcaacttgttgcataacaggca
aaccatgttatcgaaaaggagaacatcaattgtttacttgcaacaccaactcgacgaattgaat
gaaaacaagagcaaggaatttgcagagaaaaatcattcgagaacttgcattgtttgcgcattgatc
gaaggagcccaaggagcttggAACGTGAATTGAGCAGACTGCATCTTAACATTGGAAACGATT
aactacgaagaggctcaaactctcagcaagatcttgcatttaaggattgaaggaaaccgaacaaaaa
gtgaaggatattcaaacccaaTCCGCCACGTGCGTCACCTTCAGCTTGCGCTACTTGGTGAAGAAG
· ATCGACTTCGACTACACCCCCAATGGGGC**GCA**GGTACCCAAAGCAGCTACATCGACAACCTTAC
35 CTTCCCCAAGGTTCTCACCGACAAAAAAATCTCGTACCGCGTCGTGGTCAATGGCTCTGACCTTGG
CGTCGAGTCCAACCTCGCAGTGACACCGTCCGGTGGCAGACCATACTCCTCCAGTACAACAA
GGGGTATGGTGTGGCGACACCAAAACGATTCAAGTTTCGTTGTCAATTCCAGATAACGGCAACTC
GGAGGGAGTACATCATCGCTGAGTGGAAAGAAGACTTGA
40 QEHPKKDDFRNEFDHLLIEQANHAIKEKGEHQLLYLQHQLDELNENKSKELEQEKKIRELDVVCAMI
EGAQGALERELKRTDLNILERFNYEEAQTLSKILLKDLKETEQKVKDIDIQTQsatsltfqlaylvkk
idfdytpnwga~~g~~tpssyidnltfpkvltdkkysyrvvvngsdlgvesnfavtpsgqqtinf1qyn
kgygvadtktiqvfvvipdtqnseeyiiawkk

Blot 5-FveT29A (two-in-one chimeric mutant)

45 caagagcacaaggccaaagaaggatgatttccgaaacgaattcgatcacttgttgcacaggca
aaccatgttatcgaaaaggagaacatcaattgtttacttgcaacacccaactcgacgaattgaat
gaaaacaagagcaaggaattgcaagagaaaaatcattcgagaacttgcattgttgcgcattgatc
gaaggagcccaaggagcttggAACGTGAATTGAAAGCGAACTGCATCTAACATTGGAAACGATTG
aactacgaagaggctcaaactctcagcaagatcttgcattqaaggattqaaggaaaccqaacaaaaaa

gtgaaggatattcaaacccaaTCCGCCACGTCGCTCACCTCCAGCTTGCCTACTTGGTGAAGAAG
 5 ATCGACTTCGACTACACCCCCAACTGGGGCCGTGGT**GC**ACCAAGCAGCTACATCGACAACCTTAC
 CTCCCCAAGGTTCTCACCGACAAAAAAACTCGTACCGCGTGTGGTCAATGGCTCTGACCTTGG
 CGTCGAGTCCAACCTCGCAGTGACACCGTCCGGTGGCAGACCATACTCCTCCAGTACAACAA
 GGGGTATGGTGTGCGGGACACCAAAACGATTCAAGTTTCGTTGTATTCCAGATACCGGCAACTC
 GGAGGAGTACATCATCGCTGAGTGGAAAGAAGACTTGA
 QEHKPKKKDDFRNEFDHLLIEQANHAIKEHQLYLQHQLDELNENKSKELOEKIIIRELDVVCAMI
 EGAQGALERELKRTDLNILERFNYEEAQTLISKILLKDLKETEQKVVDIQTQsatsltfqlaylvkk
 10 idfdytpnwrgapssyidnltfpkvltdkkysyrvvvngsdlgvesnfavtpsggqtinflqyn
 kgyvadtktiqvfvvipdtgnseeyiaewkkt

Der p 2-FveR27A (two-in-one chimeric mutant)

gatcaagtgcgttcaaaagattgtgccaatcatgaaatcaaaaaagtttggtaccaggatgccat
 gttcagaaccatgttatcattcatcggttaaaccattccaaattggaaagccgtttcgaaagccaaac
 15 caaaacacaaaaacggctaaaattgaaatcaaaggcctcaatcgatggtttagaagttgatgttccc
 ggtatcgatccaaatgcgttccattacatgaaatgcccattggtaaaggacaacaatatgatatt
 aaatatacatggaatgttccgaaaattgcacccaaatctgaaaatgttgcgtcactgttaaagtt
 atgggtgatgatgggttttgcctgtctattgtactcatgctaaaatccgcattCCGCCACG
 TCGCTCACCTCCAGCTTGCCTACTTGGTGAAGAAGATCGACTACACCCCCAACTGGGGC
GCAGGTACCCCAAGCAGCTACATCGACAACCTTACCTTCCCAAGGTTCTCACCGACAAAAAAATA
 20 CTCGTACCGCGTGTGGTCAATGGCTCTGACCTTGGCGTCAGTCCAACCTCGCAGTGACACCGTC
 CGGTGGCAGACCATCAACTCCTCCAGTACAACAAGGGGTATGGTGTGCGGGACACCAAAACGAT
 TCAAGTTTCGTTGTCAATTCCAGATACCGGCAACTCGGAGGAGTACATCATCGCTGAGTGGAAAGAA
 GACTTGA
 25 DQVDVKDCANHEIKKVLVPGCHGSEPCI IHRGKPFQLEAVFEANQNTKTAKIEIKASIDGLEVDVP
 GIDPNACHYMKCPLVKGQQYDIKYTNVPKIAPKSENVVTVKVMGDDGVLAACIAATHAKIRDsat
 sltfqlaylvkkidfdytpnwrgapssyidnltfpkvltdkkysyrvvvngsdlgvesnfavtp
 sggqtinflqynkgyvadtktiqvfvvipdtgnseeyiaewkkt

Der p 2-FveT29A (two-in-one chimeric mutant)

gatcaagtgcgttcaaaagattgtgccaatcatgaaatcaaaaaagtttggtaccaggatgccat
 gttcagaaccatgttatcattcatcggttaaaccattccaaattggaaagccgtttcgaaagccaaac
 30 caaaacacaaaaacggctaaaattgaaatcaaaggcctcaatcgatggtttagaagttgatgttccc
 ggtatcgatccaaatgcgttccattacatgaaatgcccattggtaaaggacaacaatatgatatt
 aaatatacatggaatgttccgaaaattgcacccaaatctgaaaatgttgcgtcactgttaaagtt
 atgggtgatgatgggttttgcctgtctattgtactcatgctaaaatccgcattCCGCCACG
 TCGCTCACCTCCAGCTTGCCTACTTGGTGAAGAAGATCGACTACACCCCCAACTGGGGC
CGTGGTGCA**CCAAGCAGCTACATCGACAACCTTACCTTCCCAAGGTTCTCACCGACAAAAAAATA
 35 CTCGTACCGCGTGTGGTCAATGGCTCTGACCTTGGCGTCAGTCCAACCTCGCAGTGACACCGTC
 CGGTGGCAGACCATCAACTCCTCCAGTACAACAAGGGGTATGGTGTGCGGGACACCAAAACGAT
 TCAAGTTTCGTTGTCAATTCCAGATACCGGCAACTCGGAGGAGTACATCATCGCTGAGTGGAAAGAA
 GACTTGA
 40 DQVDVKDCANHEIKKVLVPGCHGSEPCI IHRGKPFQLEAVFEANQNTKTAKIEIKASIDGLEVDVP
 GIDPNACHYMKCPLVKGQQYDIKYTNVPKIAPKSENVVTVKVMGDDGVLAACIAATHAKIRDsat
 sltfqlaylvkkidfdytpnwrgapssyidnltfpkvltdkkysyrvvvngsdlgvesnfavtp
 sggqtinflqynkgyvadtktiqvfvvipdtgnseeyiaewkkt**

Blo t 5-Der p 2-FveR27A (three-in-one chimeric mutant)

45 caagagcacaaggccaaagaaggatattccgaaacgaattcgatcacttgttgcacaggca
 aaccatgctatcgaaaaggagaacatcaattgtttacttgcacccaaactcgacgaattgaat
 gaaaacaagagcaaggaattgcaagagaaaatcattcgagaacttgatgttgcgcattgc
 gaaggagccaaaggagcttggAACGTGAATTGAAGCGAACTGATCTTAACATTGGAAACGATT
 50 aactacgaagaggctcaaactctcagcaagatcttgcattaaaggattgaaggaaaccgaacaaaaa

5 gtgaaggatattcaaacccaagatcaagtcatgtcaaaagatttgccaatcatgaaatcaaaaaaa
 gtttggtaccaggatgccatggttcagaaccatgtatcattcatcgtaaaaccattccaaattg
 gaagccgtttcgaaagccaacaaaacacaaaaacggctaaaattgaaatcaaagcctcaatcgat
 ggtagaaagtatgttcccggtatcgatccaaatgcattacatgaaatgcccattggtt
 aaaggacaacaatatacatgaaatgttccgaaaattgacccaaaatctgaaat
 gttgtcgtactgttaaggatgggtatgggtatgttgcattgtctattgtactcatgct
 aaaatcccgatTCCGCCACGTCGCTCACCTCAGCTGCCTACTGGTGAAGAAGATCGACTTC
 GACTACACCCCCAACTGGGGC**GCA**GGTACCCAAAGCAGCTACATCGACAACCTTACCTTCCCCAA
 GGTTCTCACCGACAAAAAAACTCGTACCGCGTCGTGGTCAATGGCTCTGACCTTGGCGTCGAGTC
 10 CAACTTCGAGTGACACCGTCCGGTGGCAGACCATCAACTCCAGTACAACAAGGGGTATGG
 TGTCGCGGACACCAAAACGATTCAAGTTTCGTTGTCAATTCCAGATAACGGCAACTCGGAGGAGTA
 CATCATCGCTGAGTGGAAAGAAGACTTGA
 15 QEHKPKKDDFRNEFDHLLIEQANHAIEKGEHQQLYLOQHQLDELNENKSKEIQEKIIRELDVV
 CAMI
 EGAQGALERELKRTDLNILERFNYEEAQTLSKILLKDLKETEQVKVDIQTQDQV
 DVKDCANHEIKK
 VLVPGCHGSEPCIIHRGKPFQLEAVFEANQNTKTAKIEIKASIDGLEVDVPGIDPNACHY
 MKCPLV
 KGQQYDIKYTNVPKIAPKSENVVVTVKVMDDGVLA
 CAIA
 THAKIRDsatstfqlaylvkkidf
 dytpnwga~~g~~tpssyidnltfpkvltdkkysyrvvvngsdlgvesnfavtpsgqqtinflqynkgy
 gvadtktiqvfvvipdtgnseeyiiawekkt

FUSION PROTEINS OF VIRAL ANTIGEN AND FVE

20 *HPV E7-FveT29A*

MHGDTPTLHEYMLDLQPETTDLYCYEQLNDSSEEDEIDGPAGQAEPDRAHYNIVTFCCCDSTLR
 LCVQSTHVDIRTLEDLLMGTGIVCPICSQKPsatsltfqlaylvkkidfdytpnwrgapssyid
 nltfpkvltdkkysyrvvvngsdlgvesnfavtpsgqqtinflqynkgyvadtktiqvfvipdt
 gnseeyiiawekkt
 25 atgcatggagatacacctacattgcataatatatgttagatttgcaaccagagacaactgatctc
 tactgttatgacaattaaatgacagctcagaggaggatggaaatagatggtcagctggacaa
 gcagaaccggacagagccattacaatattgtaaccttttttttttttttttttttttttttttttt
 ttgtgcgtacaaaggcacacacgttagacatttgtacttttttttttttttttttttttttttt
 atttgtgtccccatctgttctcagaaaaccaTCCGCCACGTCGTCACCTCCAGCTTGCCTACTTG
 30 GTGAAGAAGATCGACTTCGACTACACCCCCAACTGGGGCCGTGGTGCACCAAGCAGCTACATCGAC
 AACCTTACCTCCCCAAGGTTCTCACCGACAAAAAAACTCGTACCGCGTCGTGGTCAATGGCTCT
 GACCTTGGCGTCGAGTCCAACCTCGCAGTGCACACCCTGGTGGCAGACCATCAACTCCCTCCAG
 TACAACAAGGGGTATGGTGTGCGGGACACCAAAACGATTCAAGTTTCGTTGTCAATTCCAGATACC
 GGCAACTCGGAGGAGTACATCATCGCTGAGTGGAAAGAAGACTTGA
 35

35 *HCV Core23-FveT29A*

Deletion of the 23 amino acids of core antigen from 141-163 amino acid residues
 leads to increased protein production efficiency

40 MSTNPKPQRKTKRNTNRRPQDVKFPGGGQIVGGVYLLPRRGPRLGVRATRKTSERSQPRGRROPIP
 KARQPEGRAWAQPGYPWPLYGN EGLGWAGWLLSPRGSRPSWGPTDPRRRSRNLGKVIDLT
 CGFAD
 LMGYLPLVYATGNLPGCSFSIFLLALLSCLTIPASA
 satsltfqlaylvkkidfdytpnwrgaps
 syidnltfpkvltdkkysyrvvvngsdlgvesnfavtpsgqqtinflqynkgyvadtktiqvfv
 ipdtgnseeyiiawekkt
 45 atgagcacgaatcctaaacctaagaaaaaccaacgtaaacacccaaccgcgcacaggacgtc
 aagttcccggccgggtggtcagatcgtcggtggagttacctgttgcgcgcaggggccccaggtt
 ggtgtgcgcgcacttaggaagactccgagcggtcgcaacctcgtggaaaggcgacaacctatccc
 aaggctcgccagcccagggtagggcctggctcagccggtacccctggccctatggcaat

gaggggcttgggtgggcaggatggctcctgtcaccccgtaagttggcccttagttggggccccacg
 gaccccccggcgttaggtcgcaatttggtaaggcatcgataacctcacgtcgccgttcgcgat
 ctcatgggtaccttcgcgtcggcaacaggaaactgtcccggttgctccttttatcttc
 cttttggcttgcgtcctgtttgaccatcccagttccgcttatgaagTCCGCCACGTCGCTCAC
 5 CTTCCAGCTGCCTACTTGGTGAAGAAGATCGACTTCGACTACACCCCCAACTGGGGCGTGGTGC
 ACCAACAGCTACATCGACAACCTTACCTCCCCAAGGTCTCACCGACAAAAAAATACTCGTACCG
 CGTCGTGGTCAATGGCTCTGACCTTGGCGTCGAGTCCAACCTCGCAGTGCACACCGTCCGGTGGCA
 GACCATCAACTTCCCTCCAGTACAACAAGGGTATGGTGTGCGGGACACCAAAACGATTCAAGTTT
 CGTTGTATTCCAGATAACCGGCAACTCGGAGGAGTACATCATCGCTGAGTGBAAGAAGACTTGA

10 FUSION PROTEINS OF TUMOR-ASSOCIATED ANTIGEN AND FVE

MAGE3-FveT29A

mpleqrshckpeegleargealglvgaqapateeqeaasssstlvevtlgevpaaespdpqqspq
 gasslpmtmnyplwsqsyedssnqeeegpstfpdlesefqaalsrkvaelvhfllykryarepvtk
 15 aemlgsvvgnwqyffpvifskassslqlvfgielmevdighlyifatclglsydgllgdqnqimpk
 aglliivlaiiaregdcapeekiweelsvlevfegredsilgdpkklltqhfvqenyleyrqvpgs
 dpacyeflwgralvettsyvkvlhhmvkisggphisypplhewvlregeesatsltfqlaylvkki
 dfdytpnwrgapssyidnlfpkvldkkysyrvvvngsdlgvesnfavtpsggqtinflqynkg
 ygvadtktiqvfvvipdtgnseeyiiawekkt
 atgcctcttgcagcagaggactcagcactgcaagcctgaagaaggccttgaggcccggaggagggcc
 20 ctgggcctgggtgggtgcgcaggctcctgtactgaggagcaggaggctgcctcctccttctact
 ctagttgaagtccacccctggggaggtgcctgtcggcaggactcaccatcctctggagccaatcctatgaggactccagc
 ggagcctccagcctccactaccatgaactaccctctggagccaatcctatgaggactccagc
 aaccaagaagaggagggccaagcacctccctgacctggagtcgagttccaagcagcactcagt
 25 aggaagggtggccagttggttcatttgcctcaagtatcgagccaggagccggtcacaaag
 gcagaaaatgtgggagtgtcgccaaattgcagttttcctgtatcttcagcaagact
 tccagttccctgcagctggctttggcatcgactgtatggagtcggaccatcgccacttgtac
 atctttgccacctgcctggccctctcctacgtggcctgtgggtgacaatcagatcatgcccaag
 30 gcaggcctcctgataatcgccctggccataatcgcaagagaggcgactgtgcccctgaggagaaa
 atctgggaggagctgagttgttagaggtttgagggagggaaagacagatcttggggatccc
 aagaagctgcacccaacattcgtcaggaaaactacctggagttaccggcaggtccccggcagt
 35 gatcctgcatttatgaattcctgtgggtccaaaggccctcgtgaaaccagctatgtgaaagtc
 ctgcaccatatggtaaagatcagtggaggacccatattcctaccaccctgcattgtgggtt
 ttgagagagggggaaagagTCGCCACGTCGCTCACCTCCAGCTGCCTACTTGGTAAGAAGATC
 GACTTCGACTACACCCCCAACTGGGGCGTGGTCACCAAGCAGCTACATCGACAAACCTTACCTTC
 CCCAAGGTTCTCACCGACAAAAAAATACTCGTACCGCGTCGTGGTCATGGCTCTGACCTTGGCGTC
 GAGTCCAACCTCGCAGTGACACCGTCCGGTGGGCAGACCATCAACTTCCCTCCAGTACAACAAAGGGG
 TATGGTGTGCGGGACACCAAAACGATTCAAGTTTCGTTGTATTCCAGATAACCGGAACTCGGAG
 GAGTACATCATCGCTGAGTGBAAGAAGACTTGA

MART1-FveT29A

40 mpredahfiygyppkghghsyttaeaaagigiltvilgvllligcwycrrrnyralmdkslhvgt
 qcalrrcpqegfdhrdskvslqekncepvpnappayeklsaeqspypspatsltfqlaylvk
 kidfdytpnwrgapssyidnlfpkvldkkysyrvvvngsdlgvesnfavtpsggqtinflqyn
 kgygvadtktiqvfvvipdtgnseeyiiawekkt
 atgccaagagaagatgctactcatctatggttacccaagaaggggcacggccactcttacacc
 45 acggctgaagaggccgctggatcgccatcctgacagtgtatcctggagtcctactgtctcatcgcc
 tgggttattgtagaagacgaaatggatacagagccttgcattggataaaagtcttcatgttggact
 caatgtgcctaacaagaagatgccacaaagaagggttgcattcatcgccacagcaagtgtctt
 caagagaaaaactgtgaacctgtggtccaaatgtccacctgcattatgagaaaactctgtcagaa
 cagtcaccaccacatttcacccTCCGCCACGTCGCTCACCTCCAGCTGCCTACTTGGTGAAG
 50 AAGATCGACTTCGACTACACCCCCAACTGGGGCGTGGTCACCAAGCAGCTACATCGACAAACCTT
 ACCTCCCCAAGGTTCTCACCGACAAAAAAATACTCGTACCGCGTCGTGGTCATGGCTCTGACCTT

GGCGTCGAGTCCAACCTCGCAGTGACACCGTCCGGTGGCAGACCATCAACTTCCTCCAGTACAAC
AAGGGGTATGGTGTGCGGGACACCAAAACGATTCAAGTTTCGTTGTCATTCCAGATAACCGGCAAC
TCGGAGGAGTACATCATCGCTGAGTGGAAAGAAGACTTGA

5 *CEA-FveT29A*

kltestpfnvaegkevlllvhnlpqhlfgyswykgervdgnrqiigyvigtqqatpgpaysrei
iypnasliiqniqndtgytlhvksdlvneeatgqfrvypelpksissnnspvedkdavaft
cepetqdatylwwvnnqslpvsprlqlsngnrtlfnvtrndtasykcetqnpvsarrsdsviln
vlygpadaptisplntsyrsgenlnlschaasnpaqsywfvngtfqqstqelfipnitvnnsgsyt
cqahnsdtglrnrttvttitvyaeppkpfitsnnsnpvededavaltcepeiqnttylwvnnqslp
vsprlqlsndnrnltl1sutrndvgpyecgiqnelsvdhspvilmvlygpddptispsyttyrpg
vnlslschaasnpaqsywlidgniqqhtqelfisniteknsglytcqannsasghsrttvktitv
saelpkpsissnnspvedkdavaftcepeaqnttylwvngqslpvsprlqlsngnrtlfnvt
rndarayvcgiqnsvsanrsdpvtldvlygpdtspiisppdssylsganlnlschisasnpspqswr
15 ingipqqhtqvlfiakitpnnngtyacfvsnlatgrnnnsivksitvsasgtspglsagatvgimig
vlvgvalisatsltfqlaylvkkidfdytpnwrgapssyidnltfpkvltdkkysyrvvvngsdl
gvesnfavtpsggqtinflqynkgygvadtktiqvfvvipdtgnseeyiiawekkt
aagctcaatttgaatccacgcgttcaatgtcgccagaggaaaggaggtgcttacttgccac
aatctgccccagcatctttggctacagctggtaaaaggtgaaagagtggatggcaaccgtcaa
20 attataggatatgtaatagaaactcaacaagctaccccaggccgcatacagtggtcgagagata
atatacccaatgcattccctgctgatccagaacatcatccagaatgacacaggattcaccccta
cacgtcataaagtcatcttgcataatgaaagaagcaactggccagttccgggtataccggagctg
cccaagccctccatctccagcaacaactccaaacccgtggaggacaaggatgctgtggccttacc
tgtgaacctgagactcaggacgcaacctacctgtggtaaaacaatcagagcctccggcgtcagt
25 cccaggctcagctgtccatggcaacaggaccctcactctattcaatgtcacaagaaatgacaca
gcaagctacaaatgtgaaacccagaacccagtgagtgccaggcgcagtgattcagtcattcgt
gtcctctatggcccgatgccccaccattccctctaaacacatcttacagatcagggaaaat
ctgaacctctcctgcatgcaggcttaaccacactgacatctttgtcaatggact
ttccagcaatccacccaagagctcttatccccacatcactgtgaaataatgtggatcctatacg
30 tgccaagcccataactcagacactgcctcaataggaccacagtcacgacgatcacagtctatgca
gagccacccaaacccttcatcaccacgcaacaactccaaacccgtggaggatgaggatgcttagcc
ttaacctgtgaaacctgagattcagaacacaaacctacctgtggtaaaataatcagagcctcccg
gtcagtcctcaggctgcagctgtccatgacaacacaggaccctcactctactcgtcacaaggaaat
gatgttaggaccctatgagtgaaatccagaacgaattaagtgtgaccacagcgcaccaggatc
35 ctgaatgtcctctatggccagacgacccaccattccctcatacacacttaccgtccagg
gtgaacctcagcctctcctgcatgcaggcttaaccacactgacatcttcttggctgattgat
ggAACATCCAGCAACACACACAAGAGCTCTTATCTCAACATCACTGAGAAGAACAGCGGACTC
tatacctgccaggccaataactcagccaggccacagcaggactacagtcaagacaatcacagtc
tctgcggagctgcccagccctccatctccagcaacaactccaaacccgtggaggacaaggatgct
40 gtggccttcacctgtgaaacctgaggctcagaacacacaacctacctgtggtaaaatggtcagac
ctcccagtcagttccaggctgcagctgtccatggcaacaggaccctcactctattcaatgtcaca
agaaatgacgcaagagcctatgtatgtgaaatccagaactcagtgagtgaaaccgcagtgacc
gtcacccctggatgtcctctatggccgacaccccatcattccccccagactcgttacctt
tcgggagcgaacctcaacctctcctgcccactcggcctctaaccatcccccagacttattctggcgt
45 atcaatggataccgcagcaacacacacaaggctctttatcggccaaatcacgcacaaataataac
gggacctatgcctgtttgtctctaacttggctactggccgcaataattccatagtcagagcatt
acagtctctgcattggacttctcttgcattctcgtggctactggccgcaataattccatagtc
gtgctgttgggttgctgtataTCCGCCACGTGCGTCACCTTCCAGCTTGCCTACTTGGTGAAG
AAGATCGACTTCGACTACACCCCCAACTGGGGCCGTGGTGCACCAAGCAGCTACATCGACAACCTT
50 ACCTTCCCCAAGGTTCTCACCGACAAAAAAATACTCGTACCGCGTCGTGGTCAATGGCTCTGACCTT
GGCGTCGAGTCCAACCTCGCAGTGACACCGTCCGGTGGCAGACCATCAACTTCCTCCAGTACAAC
AAGGGGTATGGTGTGCGGGACACCAAAACGATTCAAGTTTCGTTGTCATTCCAGATAACCGGCAAC
TCGGAGGAGTACATCATCGCTGAGTGGAAAGAAGACTTGA

PRIMERS FOR CONSTRUCTION OF FVE DELETION MUTANTS

Fd6-18F (36 mer)

5' -ggA/TCC/TCC/gCC/ACg/TCg/TTC/gAC/TAC/ACC/CCC/AAC- 3'

Fd6-18R (36 mer)

5 5' -gTT/ggg/ggT/gTA/gTC/gAA/CgA/CgT/ggC/ggA/ggA/TCC- 3'

Fd19-33F (36 mer)

5' -TTg/gTg/AAg/AAg/ATC/gAC/ATC/gAC/AAC/CTT/ACC/TTC- 3'

Fd19-33R (36 mer)

5' -gAA/ggT/AAg/gTT/gTC/gAT/gTC/gAT/CTT/CTT/CAC/CAA- 3'

10 *Fd34-46F (36 mer)*

5' -ggT/ACC/CCA/AgC/AgC/TAC/AAA/TAC/TCg/TAC/CgC/gTC- 3'

Fd34-46R (36 mer)

5' -gAC/gCg/gTA/CgA/gTA/TTT/gTA/gCT/gCT/Tgg/ggT/ACC- 3'

Fd47-60F (36 mer)

15 5' -AAg/gTT/CTC/ACC/gAC/AAA/gTC/gAg/TCC/AAC/TTC/gCA- 3'

Fd47-60R (36 mer)

5' -TgC/gAA/gTT/ggA/CTC/gAC/TTT/gTC/ggT/gAg/AAC/CTT- 3'

Fd61-72F (36 mer)

5' -AAT/ggC/TCT/gAC/CTT/ggC/CAg/ACC/ATC/AAC/TTC/CTC- 3'

20 *Fd61-72R (36 mer)*

5' -gAg/gAA/gTT/gAT/ggT/CTg/gCC/AAg/gTC/AgA/gCC/ATT- 3'

Fd73-84F (36 mer)

5' -gTg/ACA/CCg/TCC/ggT/ggg/ggT/gTC/gCg/gAC/ACC/AAA- 3'

Fd73-84R (36 mer)

25 5' -TTT/ggT/gTC/CgC/gAC/ACC/CCC/ACC/ggA/Cgg/Tgg/CAC- 3'

Fd85-97F (36 mer)

5' -CAg/TAC/AAC/AAg/ggg/TAT/ATT/CCA/gAT/ACC/ggC/AAC- 3'

Fd85-97R (36 mer)

5' -gTT/gCC/ggt/ATC/Tgg/AAT/ATA/CCC/CTT/gTT/gTA/CTg- 3'

30 *Fd98-106F (36 mer)*

5' -ATT/CAA/gTT/TTC/gTT/gTC/TAC/ATC/ATC/gCT/gAg/Tgg- 3'

Fd98-106R (36 mer)

5' -CCA/CTC/AgC/gAT/gAT/gTA/gAC/AAC/gAA/AAC/TTg/AAT- 3'

Fd107-115R (39 mer)

5' -gAT/gCA/ACT/gAA/TTC/TTA/CTC/CTC/CgA/gTT/gCC/ggT- 3'

PRIMERS FOR CONSTRUCTION OF LARGE FRAGMENT DELETION OF FVE*d(61-97)-F (36mer)*

5 5' -/AAT/ggC/TCT/gAC/CTT/ggC/ATT/CCA/gAT/ACC/ggC/AAC/-3'

d(61-97)-R (36mer)

5' -/gTT/gCC/ggT/ATC/Tgg/AAT/gCC/AAG/gTC/AgA/gCC/ATT/-3'

PRIMERS FOR CONSTRUCTION OF SMALL FRAGMENT OF FVE (FROM 55AA TO 100AA)*[Fv55-100]-F (48mer)*

10 5' -
/gTT/CCg/CgT/ggA/TCC/ATC/gAA/ggT/CgT/AAT/ggC/TCT/gAC/CTT/ggC/gTC/-
3'

[Fv55-100]-R (42mer)

5' -/gAT/gCA/ACT/gAA/TTC/TTA/TCA/ATC/Tgg/AAT/gAC/AAC/gAA/AAC/-3'

15 PRIMERS FOR CONSTRUCTION OF POINT MUTANTS OF FVE*F(R27A)-F (27 mer)*

5' - CCC/AAC/Tgg/ggC/gCA/ggT/ACC/CCA/AgC - 3'

F(R27A)-R (27 mer)

5' - gCT/Tgg/ggT/ACC/TgC/GCC/CCA/gTT/ggg - 3'

20 F(G28A)-F (27 mer)

5' - AAC/Tgg/ggC/CgT/gCA/ACC/CCA/AgC/AgC - 3'

F(G28A)-R (27 mer)

5' - gCT/gCT/Tgg/ggT/TgC/ACg/gCC/CCA/gTT - 3'

F(T29A)-F (27 mer)

25 5' - Tgg/ggC/CgT/ggT/gCA/CCA/AgC/AgC/TAC - 3'

F(T29A)-R (27 mer)

5' - gTA/gCT/gCT/Tgg/TgC/ACC/ACg/gCC/CCA - 3'

PRIMERS FOR BLO T 5-FVE FUSION PROTEIN*Bt5Fv-F (36mer)*

30 5' -/AAG/gAT/ATT/CAA/ACC/CAA/TCC/gCC/ACg/TCg/CTC/ACC/-3'

Bt5Fv-R (36mer)

5' -/ggT/gAg/CgA/CgT/ggC/ggA/TTg/ggT/TTg/AAT/ATC/CTT/-3'

PRIMERS FOR DER P 2-FVE FUSION PROTEIN*Dp2Fv-F (36mer)*

5 5' -/CAT/gCT/AAA/ATC/CgC/gAT/TCC/gCC/ACg/TCg/CTC/ACC-3'

Dp2Fv-R (36mer)

5' -/ggT/gAg/CgA/CgT/ggC/ggA/ATC/gCg/gAT/TTT/AgC/ATg-3'

PRIMERS FOR BLO T 5-DER P 2-FVE FUSION PROTEIN*Bt5Dp2-F (36mer)*

10 5' -/aag/gat/att/caa/acc/caa/gat/caa/gtc/gat/gtc/aaa/-3'

Bt5Dp2-R (36mer)

5' -/ttt/gac/atc/gac/ttg/atc/ttg/ggt/ttg/aat/atc/ctt/-3'

APPENDIX B: FVE FRAGMENTS (RGT TRIPLET HIGHLIGHTED)

Fragment Number	Residues	Sequence
1	24-28	WGRGT
2	25-29	GRGTP
3	26-30	RGTPS
4	27-31	GTPSS
5	28-32	TPSSY
6	23-28	NWGRGT
7	24-29	WGRGTP
8	25-30	GRGTPS
9	26-31	RGTPSS
10	27-32	GTPSSY
11	28-33	TPSSYI
12	22-28	PNWGRGT
13	23-29	NWGRGTP
14	24-30	WGRGTPS
15	25-31	GRGTPSS
16	26-32	RGTPSSY
17	27-33	GTPSSYI
18	28-34	TPSSYID
19	21-28	TPNWGRGT
20	22-29	PNWGRGTP
21	23-30	NWGRGTPS
22	24-31	WGRGTPSS
23	25-32	GRGTPSSY
24	26-33	RGTPSSYI
25	27-34	GTPSSYID
26	28-35	TPSSYIDN
27	20-28	YTPNWGRGT
28	21-29	TPNWGRGTP
29	22-30	PNWGRGTPS
30	23-31	NWGRGTPSS
31	24-32	WGRGTPSSY
32	25-33	GRGTPSSYI
33	26-34	RGTPSSYID
34	27-35	GTPSSYIDN
35	28-36	TPSSYIDNL
36	19-28	DYTPNWGRGT
37	20-29	YTPNWGRGTP
38	21-30	TPNWGRGTPS
39	22-31	PNWGRGTPSS
40	23-32	NWGRGTPSSY

Fragment Number	Residues	Sequence
41	24-33	WGRGTPSSYI
42	25-34	GRGTPSSYID
43	26-35	RGTTPSSYIDN
44	27-36	GTPSSYIDNL
45	28-37	TPSSYIDNL
46	18-28	FDYTPNWGRGT
47	19-29	DYTPNWGRGTP
48	20-30	YTPNWGRGTPS
49	21-31	TPNWGRGTPSS
50	22-32	PNWGRGTPSSY
51	23-33	NWGRGTPSSYI
52	24-34	WGRGTPSSYID
53	25-35	GRGTPSSYIDN
54	26-36	RGTTPSSYIDNL
55	27-37	GTPSSYIDNL
56	28-38	TPSSYIDNL
57	17-28	DFDYTPNWGRGT
58	18-29	FDYTPNWGRGTP
59	19-30	DYTPNWGRGTPS
60	20-31	YTPNWGRGTPSS
61	21-32	TPNWGRGTPSSY
62	22-33	PNWGRGTPSSYI
63	23-34	NWGRGTPSSYID
64	24-35	WGRGTPSSYIDN
65	25-36	GRGTPSSYIDNL
66	26-37	RGTTPSSYIDNL
67	27-38	GTPSSYIDNL
68	28-39	TPSSYIDNL
69	16-28	IDFDYTPNWGRGT
70	17-29	DFDYTPNWGRGTP
71	18-30	FDYTPNWGRGTPS
72	19-31	DYTPNWGRGTPSS
73	20-32	YTPNWGRGTPSSY
74	21-33	TPNWGRGTPSSYI
75	22-34	PNWGRGTPSSYID
76	23-35	NWGRGTPSSYIDN
77	24-36	WGRGTPSSYIDNL
78	25-37	GRGTPSSYIDNL
79	26-38	RGTTPSSYIDNL
80	27-39	GTPSSYIDNL
81	28-40	TPSSYIDNL
82	15-28	KIDFDYTPNWGRGT
83	16-29	IDFDYTPNWGRGTP

Fragment Number	Residues	Sequence
84	17-30	DFDYTPNWGRGTPS
85	18-31	FDYTPNWGRGTPSS
86	19-32	DYTPNWGRGTPSSY
87	20-33	YTPNWGRGTPSSYI
88	21-34	TPNWGRGTPSSYID
89	22-35	PNWGRGTPSSYIDN
90	23-36	NWGRGTPSSYIDNL
91	24-37	WGRGTPSSYIDNL
92	25-38	GRGTPSSYIDNLTF
93	26-39	RGTPSSYIDNLTFP
94	27-40	GTPSSYIDNLTFPK
95	28-41	TPSSYIDNLTFPKV
96	14-28	KKIDFDYTPNWGRGT
97	15-29	KIDFDYTPNWGRGTP
98	16-30	IDFDYTPNWGRGTPS
99	17-31	DFDYTPNWGRGTPSS
100	18-32	FDYTPNWGRGTPSSY
101	19-33	DYTPNWGRGTPSSYI
102	20-34	YTPNWGRGTPSSYID
103	21-35	TPNWGRGTPSSYIDN
104	22-36	PNWGRGTPSSYIDNL
105	23-37	NWGRGTPSSYIDNL
106	24-38	WGRGTPSSYIDNLTF
107	25-39	GRGTPSSYIDNLTFP
108	26-40	RGTPSSYIDNLTFPK
109	27-41	GTPSSYIDNLTFPKV
110	28-42	TPSSYIDNLTFPKVL
111	13-28	VKKIDFDYTPNWGRGT
112	14-29	KKIDFDYTPNWGRGTP
113	15-30	KIDFDYTPNWGRGTPS
114	16-31	IDFDYTPNWGRGTPSS
115	17-32	DFDYTPNWGRGTPSSY
116	18-33	FDYTPNWGRGTPSSYI
117	19-34	DYTPNWGRGTPSSYID
118	20-35	YTPNWGRGTPSSYIDN
119	21-36	TPNWGRGTPSSYIDNL
120	22-37	PNWGRGTPSSYIDNL
121	23-38	NWGRGTPSSYIDNL
122	24-39	WGRGTPSSYIDNLTFP
123	25-40	GRGTPSSYIDNLTFPK
124	26-41	RGTPSSYIDNLTFPKV
125	27-42	GTPSSYIDNLTFPKVL
126	28-43	TPSSYIDNLTFPKVLT

Fragment Number	Residues	Sequence
127	12-28	LVKKIDFDYTPNWGRGT
128	13-29	VKKIDFDYTPNWGRGTP
129	14-30	KKIDFDYTPNWGRGTPS
130	15-31	KIDFDYTPNWGRGTPSS
131	16-32	IDFDYTPNWGRGTPSSY
132	17-33	DFDYTPNWGRGTPSSYI
133	18-34	FDYTPNWGRGTPSSYID
134	19-35	DYTPNWGRGTPSSYIDN
135	20-36	YTPNWGRGTPSSYIDNL
136	21-37	TPNWGRGTPSSYIDNLT
137	22-38	PNWGRGTPSSYIDNLTF
138	23-39	NWGRGTPSSYIDNLTFP
139	24-40	WGRGTPSSYIDNLTFPK
140	25-41	GRGTPSSYIDNLTFPKV
141	26-42	RGTPSSYIDNLTFPKVL
142	27-43	GTPSSYIDNLTFPKVLT
143	28-44	TPSSYIDNLTFPKVLD
144	11-28	YLVKKIDFDYTPNWGRGT
145	12-29	LVKKIDFDYTPNWGRGTP
146	13-30	VKKIDFDYTPNWGRGTPS
147	14-31	KKIDFDYTPNWGRGTPSS
148	15-32	KIDFDYTPNWGRGTPSSY
149	16-33	IDFDYTPNWGRGTPSSYI
150	17-34	DFDYTPNWGRGTPSSYID
151	18-35	FDYTPNWGRGTPSSYIDN
152	19-36	DYTPNWGRGTPSSYIDNL
153	20-37	YTPNWGRGTPSSYIDNL
154	21-38	TPNWGRGTPSSYIDNLTF
155	22-39	PNWGRGTPSSYIDNLTFP
156	23-40	NWGRGTPSSYIDNLTFPK
157	24-41	WGRGTPSSYIDNLTFPKV
158	25-42	GRGTPSSYIDNLTFPKVL
159	26-43	RGTPSSYIDNLTFPKVLT
160	27-44	GTPSSYIDNLTFPKVLD
161	28-45	TPSSYIDNLTFPKVLDK
162	10-28	AYLVKKIDFDYTPNWGRGT
163	11-29	YLVKKIDFDYTPNWGRGTP
164	12-30	LVKKIDFDYTPNWGRGTPS
165	13-31	VKKIDFDYTPNWGRGTPSS
166	14-32	KKIDFDYTPNWGRGTPSSY
167	15-33	KIDFDYTPNWGRGTPSSYI
168	16-34	IDFDYTPNWGRGTPSSYID
169	17-35	DFDYTPNWGRGTPSSYIDN

Fragment Number	Residues	Sequence
170	18-36	FDYTPNWGRGTPSSYIDNL
171	19-37	DYTPNWGRGTPSSYIDNL
172	20-38	YTPNWGRGTPSSYIDNLTF
173	21-39	TPNWGRGTPSSYIDNLTFP
174	22-40	PNWGRGTPSSYIDNLTFPK
175	23-41	NWGRGTPSSYIDNLTFPKV
176	24-42	WGRGTPSSYIDNLTFPKVL
177	25-43	GRGTPSSYIDNLTFPKVLT
178	26-44	RGTTPSSYIDNLTFPKVLT
179	27-45	GTPSSYIDNLTFPKVLTDK
180	28-46	TPSSYIDNLTFPKVLTDKK
181	9-28	LAYLVKKIDFDYTPNWGRG
182	10-29	AYLVKKIDFDYTPNWGRGTP
183	11-30	YLVKKIDFDYTPNWGRGTPS
184	12-31	LVKKIDFDYTPNWGRGTPSS
185	13-32	VKKIDFDYTPNWGRGTPSSY
186	14-33	KKIDFDYTPNWGRGTPSSYI
187	15-34	KIDFDYTPNWGRGTPSSYID
188	16-35	IDFDYTPNWGRGTPSSYIDN
189	17-36	DFDYTPNWGRGTPSSYIDNL
190	18-37	FDYTPNWGRGTPSSYIDNL
191	19-38	DYTPNWGRGTPSSYIDNLTF
192	20-39	YTPNWGRGTPSSYIDNLTFP
193	21-40	TPNWGRGTPSSYIDNLTFPK
194	22-41	PNWGRGTPSSYIDNLTFPKV
195	23-42	NWGRGTPSSYIDNLTFPKV
196	24-43	WGRGTPSSYIDNLTFPKVLT
197	25-44	GRGTPSSYIDNLTFPKVLT
198	26-45	RGTTPSSYIDNLTFPKVLT
199	27-46	GTPSSYIDNLTFPKVLTDKK
200	28-47	TPSSYIDNLTFPKVLTDKK
201	8-28	QAYLVKKIDFDYTPNWGRG
202	9-29	AYLVKKIDFDYTPNWGRGTP
203	10-30	AYLVKKIDFDYTPNWGRGTPS
204	11-31	YLVKKIDFDYTPNWGRGTPSS
205	12-32	LVKKIDFDYTPNWGRGTPSSY
206	13-33	VKKIDFDYTPNWGRGTPSSYI
207	14-34	KKIDFDYTPNWGRGTPSSYID
208	15-35	KIDFDYTPNWGRGTPSSYIDN
209	16-36	IDFDYTPNWGRGTPSSYIDNL
210	17-37	DFDYTPNWGRGTPSSYIDNL
211	18-38	FDYTPNWGRGTPSSYIDNL
212	19-39	DYTPNWGRGTPSSYIDNL

Fragment Number	Residues	Sequence
213	20-40	YTPNWGRGTPSSYIDNLTFPK
214	21-41	TPNWGRGTPSSYIDNLTFPKV
215	22-42	PNWGRGTPSSYIDNLTFPKVL
216	23-43	NWGRGTPSSYIDNLTFPKVLT
217	24-44	WGRGTPSSYIDNLTFPKVLT
218	25-45	GRGTPSSYIDNLTFPKVLT
219	26-46	RGT
220	27-47	GTPSSYIDNLTFPKVLT
221	28-48	T
222	7-28	FQLAYLVKKIDFDYTPNWGRG
223	8-29	T
224	9-30	LAYLVKKIDFDYTPNWGRG
225	10-31	T
226	11-32	A
227	12-33	YLVKKIDFDYTPNWGRG
228	13-34	T
229	14-35	VKKIDFDYTPNWGRG
230	15-36	T
231	16-37	KIDFDYTPNWGRG
232	17-38	T
233	18-39	DYTPNWGRG
234	19-40	T
235	20-41	YTPNWGRG
236	21-42	T
237	22-43	PNWGRG
238	23-44	T
239	24-45	WGRG
240	25-46	T
241	26-47	GRGTPSSYIDNLTFPKVLT
242	27-48	RGT
243	28-49	GTPSSYIDNLTFPKVLT
244	6-28	T
245	7-29	FQLAYLVKKIDFDYTPNWGRG
246	8-30	T
247	9-31	QLAYLVKKIDFDYTPNWGRG
248	10-32	T
249	11-33	LAYLVKKIDFDYTPNWGRG
250	12-34	T
251	13-35	A
252	14-36	YLVKKIDFDYTPNWGRG
253	15-37	T
254	16-38	VKKIDFDYTPNWGRG
255	17-39	T

Fragment Number	Residues	Sequence
256	18-40	FDYTPNWGRGTPSSYIDNLTFPK
257	19-41	DYTPNWGRGTPSSYIDNLTFPKV
258	20-42	YTPNWGRGTPSSYIDNLTFPKVL
259	21-43	TPNWGRGTPSSYIDNLTFPKVLT
260	22-44	PNWGRGTPSSYIDNLTFPKVLTD
261	23-45	NWGRGTPSSYIDNLTFPKVLTDK
262	24-46	WGRGTPSSYIDNLTFPKVLTDKK
263	25-47	GRGTPSSYIDNLTFPKVLTDKKY
264	26-48	RGTTPSSYIDNLTFPKVLTDKKYS
265	27-49	GTPSSYIDNLTFPKVLTDKKYSY
266	28-50	TPSSYIDNLTFPKVLTDKKYSR
267	5-28	LTFQLAYLVKKIDFDYTPNWGRGRT
268	6-29	TFQLAYLVKKIDFDYTPNWGRGTP
269	7-30	FQLAYLVKKIDFDYTPNWGRGTPS
270	8-31	QLAYLVKKIDFDYTPNWGRGTPSS
271	9-32	LAYLVKKIDFDYTPNWGRGTPSSY
272	10-33	AYLVKKIDFDYTPNWGRGTPSSYI
273	11-34	YLVKKIDFDYTPNWGRGTPSSYID
274	12-35	LVKKIDFDYTPNWGRGTPSSYIDN
275	13-36	VKKIDFDYTPNWGRGTPSSYIDNL
276	14-37	KKIDFDYTPNWGRGTPSSYIDNLT
277	15-38	KIDFDYTPNWGRGTPSSYIDNLTF
278	16-39	IDFDYTPNWGRGTPSSYIDNLTFP
279	17-40	DFDYTPNWGRGTPSSYIDNLTFPK
280	18-41	FDYTPNWGRGTPSSYIDNLTFPKV
281	19-42	DYTPNWGRGTPSSYIDNLTFPKVL
282	20-43	YTPNWGRGTPSSYIDNLTFPKVLT
283	21-44	TPNWGRGTPSSYIDNLTFPKVLTD
284	22-45	PNWGRGTPSSYIDNLTFPKVLTDK
285	23-46	NWGRGTPSSYIDNLTFPKVLTDKK
286	24-47	WGRGTPSSYIDNLTFPKVLTDKKY
287	25-48	GRGTPSSYIDNLTFPKVLTDKKYS
288	26-49	RGTTPSSYIDNLTFPKVLTDKKYSY
289	27-50	GTPSSYIDNLTFPKVLTDKKYSR
290	28-51	TPSSYIDNLTFPKVLTDKKYSYRV
291	4-28	SLTFQLAYLVKKIDFDYTPNWGRGRT
292	5-29	LTFQLAYLVKKIDFDYTPNWGRGTP
293	6-30	TFQLAYLVKKIDFDYTPNWGRGTPS
294	7-31	FQLAYLVKKIDFDYTPNWGRGTPSS
295	8-32	QLAYLVKKIDFDYTPNWGRGTPSSY
296	9-33	LAYLVKKIDFDYTPNWGRGTPSSYI
297	10-34	AYLVKKIDFDYTPNWGRGTPSSYID
298	11-35	YLVKKIDFDYTPNWGRGTPSSYIDN

Fragment Number	Residues	Sequence
299	12-36	LVKKIDFDYTPNWGRGTPSSYIDNL
300	13-37	VKKIDFDYTPNWGRGTPSSYIDNL
301	14-38	KKIDFDYTPNWGRGTPSSYIDNL
302	15-39	KIDFDYTPNWGRGTPSSYIDNL
303	16-40	IDFDYTPNWGRGTPSSYIDNL
304	17-41	DFDYTPNWGRGTPSSYIDNL
305	18-42	F DYTPNWGRGTPSSYIDNL
306	19-43	DYTPNWGRGTPSSYIDNL
307	20-44	YTPNWGRGTPSSYIDNL
308	21-45	TPNWGRGTPSSYIDNL
309	22-46	PNWGRGTPSSYIDNL
310	23-47	NWGRGTPSSYIDNL
311	24-48	WGRGTPSSYIDNL
312	25-49	GRGTPSSYIDNL
313	26-50	RGT PSSYIDNL
314	27-51	GTPSSYIDNL
315	28-52	TPSSYIDNL
316	3-28	TSLTFQLAYLVKKIDFDYTPNWGRG
317	4-29	SLTFQLAYLVKKIDFDYTPNWGRG
318	5-30	LTFQLAYLVKKIDFDYTPNWGRG
319	6-31	TFQLAYLVKKIDFDYTPNWGRG
320	7-32	FQLAYLVKKIDFDYTPNWGRG
321	8-33	QLAYLVKKIDFDYTPNWGRG
322	9-34	LAYLVKKIDFDYTPNWGRG
323	10-35	AYLVKKIDFDYTPNWGRG
324	11-36	YLVKKIDFDYTPNWGRG
325	12-37	LVKKIDFDYTPNWGRG
326	13-38	VKKIDFDYTPNWGRG
327	14-39	KKIDFDYTPNWGRG
328	15-40	KIDFDYTPNWGRG
329	16-41	IDFDYTPNWGRG
330	17-42	DFDYTPNWGRG
331	18-43	F DYTPNWGRG
332	19-44	DYTPNWGRG
333	20-45	YTPNWGRG
334	21-46	TPNWGRG
335	22-47	PNWGRG
336	23-48	NWGRG
337	24-49	WGRG
338	25-50	GRGTPSSYIDNL
339	26-51	RGT PSSYIDNL
340	27-52	GTPSSYIDNL
341	28-53	TPSSYIDNL

Fragment Number	Residues	Sequence
342	2-28	ATSLTFQLAYLVKKIDFDYTPNWGRGT
343	3-29	TSLTFQLAYLVKKIDFDYTPNWGRGTP
344	4-30	SLTFQLAYLVKKIDFDYTPNWGRGTPS
345	5-31	LTFQLAYLVKKIDFDYTPNWGRGTPSS
346	6-32	TFQLAYLVKKIDFDYTPNWGRGTPSSY
347	7-33	FQLAYLVKKIDFDYTPNWGRGTPSSYI
348	8-34	QLAYLVKKIDFDYTPNWGRGTPSSYID
349	9-35	LAYLVKKIDFDYTPNWGRGTPSSYIDN
350	10-36	AYLVKKIDFDYTPNWGRGTPSSYIDNL
351	11-37	YLVKKIDFDYTPNWGRGTPSSYIDNLT
352	12-38	LVKKIDFDYTPNWGRGTPSSYIDNLTF
353	13-39	VKKIDFDYTPNWGRGTPSSYIDNLTFP
354	14-40	KKIDFDYTPNWGRGTPSSYIDNLTFPK
355	15-41	KIDFDYTPNWGRGTPSSYIDNLTFPKV
356	16-42	IDFDYTPNWGRGTPSSYIDNLTFPKVL
357	17-43	DFDYTPNWGRGTPSSYIDNLTFPKVLT
358	18-44	FDYTPNWGRGTPSSYIDNLTFPKVLT
359	19-45	DYTPNWGRGTPSSYIDNLTFPKVLTDK
360	20-46	YTPNWGRGTPSSYIDNLTFPKVLTDKK
361	21-47	TPNWGRGTPSSYIDNLTFPKVLTDKKY
362	22-48	PNWGRGTPSSYIDNLTFPKVLTDKKYS
363	23-49	NWGRGTPSSYIDNLTFPKVLTDKKYSY
364	24-50	WGRGTPSSYIDNLTFPKVLTDKKYSYR
365	25-51	GRGTPSSYIDNLTFPKVLTDKKYSYRV
366	26-52	RGTPSSYIDNLTFPKVLTDKKYSYRVVV
367	27-53	GTPSSYIDNLTFPKVLTDKKYSYRVVV
368	28-54	TPSSYIDNLTFPKVLTDKKYSYRVVVN
369	1-28	SATSLTFQLAYLVKKIDFDYTPNWGRGT
370	2-29	ATSLTFQLAYLVKKIDFDYTPNWGRGTP
371	3-30	TSLTFQLAYLVKKIDFDYTPNWGRGTPS
372	4-31	SLTFQLAYLVKKIDFDYTPNWGRGTPSS
373	5-32	LTFQLAYLVKKIDFDYTPNWGRGTPSSY
374	6-33	TFQLAYLVKKIDFDYTPNWGRGTPSSYI
375	7-34	FQLAYLVKKIDFDYTPNWGRGTPSSYID
376	8-35	QLAYLVKKIDFDYTPNWGRGTPSSYIDN
377	9-36	LAYLVKKIDFDYTPNWGRGTPSSYIDNL
378	10-37	AYLVKKIDFDYTPNWGRGTPSSYIDNL
379	11-38	YLVKKIDFDYTPNWGRGTPSSYIDNLTF
380	12-39	LVKKIDFDYTPNWGRGTPSSYIDNLTFP
381	13-40	VKKIDFDYTPNWGRGTPSSYIDNLTFPK
382	14-41	KKIDFDYTPNWGRGTPSSYIDNLTFPKV
383	15-42	KIDFDYTPNWGRGTPSSYIDNLTFPKVL
384	16-43	IDFDYTPNWGRGTPSSYIDNLTFPKVLT

Fragment Number	Residues	Sequence
385	17-44	DFDYTPNWGRGTPSSYIDNLTFPKVLTD
386	18-45	FDYTPNWGRGTPSSYIDNLTFPKVLTDK
387	19-46	DYTPNWGRGTPSSYIDNLTFPKVLTDKK
388	20-47	YTPNWGRGTPSSYIDNLTFPKVLTDKKY
389	21-48	TPNWGRGTPSSYIDNLTFPKVLTDKKYS
390	22-49	PNWGRGTPSSYIDNLTFPKVLTDKKSY
391	23-50	NWGRGTPSSYIDNLTFPKVLTDKKYSYR
392	24-51	WGRGTPSSYIDNLTFPKVLTDKKYSYRV
393	25-52	GRGTPSSYIDNLTFPKVLTDKKYSYRVV
394	26-53	RGTPSSYIDNLTFPKVLTDKKYSYRVVV
395	27-54	GTPSSYIDNLTFPKVLTDKKYSYRVVVN
396	28-55	TPSSYIDNLTFPKVLTDKKYSYRVVVNG

APPENDIX C: CRYSTAL COORDINATES OF FVE PROTEIN

		XX-XXX-XX	xxxx
HEADER	----		
COMPND	---		
REMARK	3		
5 REMARK	3	REFINEMENT.	
REMARK	3	PROGRAM : REFMAC 5.0	
REMARK	3	AUTHORS : MURSHUDOV, VAGIN, DODSON	
REMARK	3		
REMARK	3	REFINEMENT TARGET : MAXIMUM LIKELIHOOD	
10 REMARK	3		
REMARK	3	DATA USED IN REFINEMENT.	
REMARK	3	RESOLUTION RANGE HIGH (ANGSTROMS) : 1.70	
REMARK	3	RESOLUTION RANGE LOW (ANGSTROMS) : 30.02	
REMARK	3	DATA CUTOFF (SIGMA(F)) : NONE	
15 REMARK	3	COMPLETENESS FOR RANGE (%) : 98.80	
REMARK	3	NUMBER OF REFLECTIONS : 30783	
REMARK	3		
REMARK	3	FIT TO DATA USED IN REFINEMENT.	
REMARK	3	CROSS-VALIDATION METHOD : THROUGHOUT	
20 REMARK	3	FREE R VALUE TEST SET SELECTION : RANDOM	
REMARK	3	R VALUE (WORKING + TEST SET) : 0.18358	
REMARK	3	R VALUE (WORKING SET) : 0.18218	
REMARK	3	FREE R VALUE : 0.21016	
REMARK	3	FREE R VALUE TEST SET SIZE (%) : 5.1	
25 REMARK	3	FREE R VALUE TEST SET COUNT : 1650	
REMARK	3		
REMARK	3	FIT IN THE HIGHEST RESOLUTION BIN.	
REMARK	3	TOTAL NUMBER OF BINS USED : 20	
REMARK	3	BIN RESOLUTION RANGE HIGH : 1.701	
30 REMARK	3	BIN RESOLUTION RANGE LOW : 1.745	
REMARK	3	REFLECTION IN BIN (WORKING SET) : 2183	
REMARK	3	BIN R VALUE (WORKING SET) : 0.160	
REMARK	3	BIN FREE R VALUE SET COUNT : 114	
REMARK	3	BIN FREE R VALUE : 0.197	
35 REMARK	3		
REMARK	3	NUMBER OF NON-HYDROGEN ATOMS USED IN REFINEMENT.	
REMARK	3	ALL ATOMS : 1940	
REMARK	3		
REMARK	3	B VALUES.	
40 REMARK	3	FROM WILSON PLOT (A**2) : NULL	
REMARK	3	MEAN B VALUE (OVERALL, A**2) : 13.666	
REMARK	3	OVERALL ANISOTROPIC B VALUE.	
REMARK	3	B11 (A**2) : -0.02	
REMARK	3	B22 (A**2) : -0.02	
45 REMARK	3	B33 (A**2) : 0.03	
REMARK	3	B12 (A**2) : 0.00	
REMARK	3	B13 (A**2) : 0.00	
REMARK	3	B23 (A**2) : 0.00	
REMARK	3		
50 REMARK	3	ESTIMATED OVERALL COORDINATE ERROR. (A) : 0.092	
REMARK	3	ESU BASED ON R VALUE (A) : 0.092	
REMARK	3	ESU BASED ON FREE R VALUE (A) : 0.075	
REMARK	3	ESU BASED ON MAXIMUM LIKELIHOOD (A**2) : 2.208	
REMARK	3	ESU FOR B VALUES BASED ON MAXIMUM LIKELIHOOD (A**2) :	
55 REMARK	3		
REMARK	3	CORRELATION COEFFICIENTS.	
REMARK	3	CORRELATION COEFFICIENT FO-FC : 0.947	
REMARK	3	CORRELATION COEFFICIENT FO-FC FREE : 0.933	
REMARK	3		
60 REMARK	3	RMS DEVIATIONS FROM IDEAL VALUES COUNT RMS WEIGHT	

REMARK 3 BOND LENGTHS REFINED ATOMS (A) : 1830 ; 0.010 ; 0.022
 REMARK 3 BOND LENGTHS OTHERS (A) : 1593 ; 0.001 ; 0.020
 REMARK 3 BOND ANGLES REFINED ATOMS (DEGREES) : 2490 ; 1.466 ; 1.941
 REMARK 3 BOND ANGLES OTHERS (DEGREES) : 3724 ; 0.921 ; 3.000
 5 REMARK 3 TORSION ANGLES, PERIOD 1 (DEGREES) : 224 ; 4.899 ; 3.000
 REMARK 3 TORSION ANGLES, PERIOD 3 (DEGREES) : 311 ; 16.844 ; 15.000
 REMARK 3 CHIRAL-CENTER RESTRAINTS (A**3) : 280 ; 0.231 ; 0.200
 REMARK 3 GENERAL PLANES REFINED ATOMS (A) : 2026 ; 0.006 ; 0.020
 REMARK 3 GENERAL PLANES OTHERS (A) : 374 ; 0.003 ; 0.020
 10 REMARK 3 NON-BONDED CONTACTS REFINED ATOMS (A) : 327 ; 0.271 ; 0.300
 REMARK 3 NON-BONDED CONTACTS OTHERS (A) : 1447 ; 0.212 ; 0.300
 REMARK 3 H-BOND (X...Y) REFINED ATOMS (A) : 131 ; 0.131 ; 0.500
 REMARK 3 SYMMETRY VDW REFINED ATOMS (A) : 8 ; 0.310 ; 0.300
 REMARK 3 SYMMETRY VDW OTHERS (A) : 17 ; 0.291 ; 0.300
 15 REMARK 3 SYMMETRY H-BOND REFINED ATOMS (A) : 14 ; 0.144 ; 0.500
 REMARK 3
 REMARK 3 ISOTROPIC THERMAL FACTOR RESTRAINTS. COUNT RMS WEIGHT
 REMARK 3 MAIN-CHAIN BOND REFINED ATOMS (A**2) : 1124 ; 0.898 ; 1.500
 REMARK 3 MAIN-CHAIN ANGLE REFINED ATOMS (A**2) : 1827 ; 1.603 ; 2.000
 20 REMARK 3 SIDE-CHAIN BOND REFINED ATOMS (A**2) : 706 ; 2.292 ; 3.000
 REMARK 3 SIDE-CHAIN ANGLE REFINED ATOMS (A**2) : 663 ; 3.839 ; 4.500
 REMARK 3
 REMARK 3 NCS RESTRAINTS STATISTICS
 REMARK 3 NUMBER OF NCS GROUPS : NULL
 25 REMARK 3
 REMARK 3
 REMARK 3 TLS DETAILS
 REMARK 3 NUMBER OF TLS GROUPS : 2
 REMARK 3
 30 REMARK 3 TLS GROUP : 1
 REMARK 3 NUMBER OF COMPONENTS GROUP : 1
 REMARK 3 COMPONENTS C SSSEQI TO C SSSEQI
 REMARK 3 RESIDUE RANGE : A 1 A 113
 REMARK 3 ORIGIN FOR THE GROUP (A) : 31.8380 34.4130 15.9540
 35 REMARK 3 T TENSOR
 REMARK 3 T11: 0.0826 T22: 0.0528
 REMARK 3 T33: 0.0022 T12: 0.0085
 REMARK 3 T13: 0.0118 T23: 0.0066
 REMARK 3 L TENSOR
 40 REMARK 3 L11: 0.3236 L22: 1.6346
 REMARK 3 L33: 0.0319 L12: -0.4538
 REMARK 3 L13: -0.1060 L23: -0.1134
 REMARK 3 S TENSOR
 REMARK 3 S11: 0.0668 S12: 0.0317 S13: 0.0266
 45 REMARK 3 S21: -0.0158 S22: -0.0508 S23: -0.0656
 REMARK 3 S31: -0.0111 S32: 0.0027 S33: -0.0160
 REMARK 3
 REMARK 3 TLS GROUP : 2
 REMARK 3 NUMBER OF COMPONENTS GROUP : 1
 50 REMARK 3 COMPONENTS C SSSEQI TO C SSSEQI
 REMARK 3 RESIDUE RANGE : B 1 B 112
 REMARK 3 ORIGIN FOR THE GROUP (A) : 33.7580 2.5150 18.4210
 REMARK 3 T TENSOR
 REMARK 3 T11: 0.0638 T22: 0.0608
 55 REMARK 3 T33: 0.0227 T12: 0.0019
 REMARK 3 T13: -0.0064 T23: -0.0055
 REMARK 3 L TENSOR
 REMARK 3 L11: 0.0923 L22: 0.6926
 REMARK 3 L33: 0.1427 L12: -0.1092
 60 REMARK 3 L13: -0.1135 L23: -0.0160
 REMARK 3 S TENSOR
 REMARK 3 S11: 0.0096 S12: 0.0276 S13: -0.0212
 REMARK 3 S21: -0.0046 S22: -0.0327 S23: 0.0279

REMARK 3 S31: -0.0061 S32: -0.0095 S33: 0.0231
 REMARK 3
 REMARK 3
 REMARK 3 BULK SOLVENT MODELLING.
 5 REMARK 3 METHOD USED : BABBINET MODEL WITH MASK
 REMARK 3 PARAMETERS FOR MASK CALCULATION
 REMARK 3 VDW PROBE RADIUS : 1.40
 REMARK 3 ION PROBE RADIUS : 0.80
 REMARK 3 SHRINKAGE RADIUS : 0.80
 10 REMARK 3
 REMARK 3 OTHER REFINEMENT REMARKS:
 REMARK 3 HYDROGENS HAVE BEEN ADDED IN THE RIDING POSITIONS
 REMARK 3
 CISPEP 1 THR A 28 PRO A 29 0.00
 15 CISPEP 2 THR B 28 PRO B 29 0.00
 CRYST1 97.118 97.118 61.413 90.00 90.00 90.00 P 43 21 2
 SCALE1 0.010297 0.000000 0.000000 0.000000
 SCALE2 0.000000 0.010297 0.000000 0.000000
 SCALE3 0.000000 0.000000 0.016283 0.000000
 20 ATOM 1 O ACE A 0 39.758 17.815 6.621 1.00 32.04 O
 ATOM 2 C ACE A 0 38.470 17.959 6.297 1.00 30.44 C
 ATOM 3 CA ACE A 0 37.841 19.332 5.940 1.00 30.13 C
 ATOM 4 N SER A 1 37.877 16.775 5.643 1.00 19.18 N
 ATOM 6 CA SER A 1 36.408 16.741 5.468 1.00 17.19 C
 25 ATOM 8 CB SER A 1 35.991 15.421 4.841 1.00 17.15 C
 ATOM 11 OG SER A 1 36.194 14.363 5.768 1.00 16.56 O
 ATOM 13 C SER A 1 35.748 16.842 6.834 1.00 16.94 C
 ATOM 14 O SER A 1 36.412 16.630 7.854 1.00 16.93 O
 ATOM 17 N ALA A 2 34.500 17.297 6.850 1.00 17.11 N
 30 ATOM 19 CA ALA A 2 33.637 17.247 8.031 1.00 16.12 C
 ATOM 21 CB ALA A 2 32.200 17.465 7.619 1.00 16.40 C
 ATOM 25 C ALA A 2 33.762 15.907 8.757 1.00 15.10 C
 ATOM 26 O ALA A 2 33.901 15.848 9.975 1.00 13.93 C
 ATOM 27 N THR A 3 33.680 14.823 8.009 1.00 14.66 N
 35 ATOM 29 CA THR A 3 33.773 13.515 8.630 1.00 13.12 C
 ATOM 31 CB THR A 3 33.497 12.440 7.599 1.00 13.38 C
 ATOM 33 OG1 THR A 3 32.154 12.599 7.122 1.00 13.50 C
 ATOM 35 CG2 THR A 3 33.517 11.067 8.238 1.00 14.13 C
 ATOM 39 C THR A 3 35.111 13.272 9.307 1.00 12.51 C
 40 ATOM 40 O THR A 3 35.141 12.780 10.440 1.00 10.83 O
 ATOM 41 N SER A 4 36.216 13.578 8.632 1.00 11.39 N
 ATOM 43 CA SER A 4 37.538 13.356 9.244 1.00 12.60 C
 ATOM 45 CB SER A 4 38.694 13.609 8.266 1.00 13.31 C
 ATOM 48 OG SER A 4 38.566 14.874 7.668 1.00 19.57 O
 45 ATOM 50 C SER A 4 37.726 14.223 10.471 1.00 11.69 C
 ATOM 51 O SER A 4 38.223 13.765 11.484 1.00 10.87 O
 ATOM 52 N LEU A 5 37.331 15.484 10.379 1.00 11.95 N
 ATOM 54 CA LEU A 5 37.478 16.382 11.515 1.00 11.00 C
 ATOM 56 CB LEU A 5 37.047 17.801 11.149 1.00 11.44 C
 50 ATOM 59 CG LEU A 5 37.928 18.509 10.117 1.00 13.46 C
 ATOM 61 CD1 LEU A 5 37.267 19.790 9.651 1.00 15.05 C
 ATOM 65 CD2 LEU A 5 39.270 18.807 10.731 1.00 15.52 C
 ATOM 69 C LEU A 5 36.658 15.900 12.698 1.00 10.25 C
 ATOM 70 O LEU A 5 37.114 15.947 13.852 1.00 9.79 O
 55 ATOM 71 N THR A 6 35.440 15.446 12.417 1.00 9.51 N
 ATOM 73 CA THR A 6 34.547 14.953 13.459 1.00 9.80 C
 ATOM 75 CB THR A 6 33.250 14.425 12.840 1.00 9.84 C
 ATOM 77 OG1 THR A 6 32.454 15.510 12.319 1.00 10.30 O
 ATOM 79 CG2 THR A 6 32.388 13.749 13.859 1.00 9.40 C
 60 ATOM 83 C THR A 6 35.186 13.816 14.236 1.00 9.72 C
 ATOM 84 O THR A 6 35.215 13.845 15.451 1.00 9.30 O
 ATOM 85 N PHE A 7 35.679 12.796 13.545 1.00 9.95 C
 ATOM 87 CA PHE A 7 36.185 11.642 14.278 1.00 8.92 C

										C	
										C	
										C	
										C	
										C	
5	ATOM	89	CB	PHE	A	7	35.993	10.367	13.490	1.00	8.90
	ATOM	92	CG	PHE	A	7	34.552	9.988	13.365	1.00	8.19
	ATOM	93	CD1	PHE	A	7	33.848	9.583	14.485	1.00	10.40
	ATOM	95	CE1	PHE	A	7	32.512	9.267	14.407	1.00	10.95
	ATOM	97	CZ	PHE	A	7	31.848	9.370	13.217	1.00	11.35
	ATOM	99	CE2	PHE	A	7	32.532	9.791	12.080	1.00	10.55
	ATOM	101	CD2	PHE	A	7	33.872	10.127	12.165	1.00	10.65
	ATOM	103	C	PHE	A	7	37.603	11.819	14.812	1.00	9.58
	ATOM	104	O	PHE	A	7	37.970	11.203	15.811	1.00	9.17
10	ATOM	105	N	GLN	A	8	38.405	12.669	14.177	1.00	9.36
	ATOM	107	CA	GLN	A	8	39.683	12.999	14.778	1.00	10.36
	ATOM	109	CB	GLN	A	8	40.476	13.937	13.891	1.00	10.90
	ATOM	112	CG	GLN	A	8	41.097	13.322	12.692	1.00	14.14
	ATOM	115	CD	GLN	A	8	41.805	14.419	11.894	1.00	16.75
15	ATOM	116	OE1	GLN	A	8	41.409	14.742	10.787	1.00	21.77
	ATOM	117	NE2	GLN	A	8	42.799	15.056	12.517	1.00	20.28
	ATOM	120	C	GLN	A	8	39.409	13.716	16.116	1.00	10.53
	ATOM	121	O	GLN	A	8	40.049	13.416	17.118	1.00	10.95
	ATOM	122	N	LEU	A	9	38.457	14.654	16.122	1.00	9.95
20	ATOM	124	CA	LEU	A	9	38.145	15.413	17.332	1.00	9.62
	ATOM	126	CB	LEU	A	9	37.162	16.537	17.057	1.00	9.66
	ATOM	129	CG	LEU	A	9	36.767	17.375	18.278	1.00	9.80
	ATOM	131	CD1	LEU	A	9	37.974	18.098	18.862	1.00	10.08
	ATOM	135	CD2	LEU	A	9	35.701	18.397	17.886	1.00	12.75
25	ATOM	139	C	LEU	A	9	37.541	14.467	18.346	1.00	9.58
	ATOM	140	O	LEU	A	9	37.935	14.484	19.514	1.00	9.46
	ATOM	141	N	ALA	A	10	36.588	13.637	17.917	1.00	9.20
	ATOM	143	CA	ALA	A	10	35.952	12.701	18.856	1.00	9.03
	ATOM	145	CB	ALA	A	10	34.875	11.850	18.154	1.00	8.72
30	ATOM	149	C	ALA	A	10	36.949	11.802	19.605	1.00	8.50
	ATOM	150	O	ALA	A	10	36.855	11.615	20.825	1.00	8.50
	ATOM	151	N	TYR	A	11	37.918	11.242	18.899	1.00	9.18
	ATOM	153	CA	TYR	A	11	38.865	10.359	19.541	1.00	8.12
	ATOM	155	CB	TYR	A	11	39.716	9.664	18.491	1.00	8.30
35	ATOM	158	CG	TYR	A	11	40.642	8.638	19.075	1.00	7.61
	ATOM	159	CD1	TYR	A	11	40.156	7.495	19.699	1.00	9.07
	ATOM	161	CE1	TYR	A	11	41.008	6.560	20.229	1.00	10.41
	ATOM	163	CZ	TYR	A	11	42.359	6.768	20.170	1.00	13.73
	ATOM	164	OH	TYR	A	11	43.210	5.831	20.740	1.00	15.09
40	ATOM	166	CE2	TYR	A	11	42.868	7.898	19.571	1.00	10.94
	ATOM	168	CD2	TYR	A	11	42.014	8.827	19.027	1.00	10.00
	ATOM	170	C	TYR	A	11	39.752	11.139	20.530	1.00	8.66
	ATOM	171	O	TYR	A	11	40.158	10.596	21.550	1.00	8.96
	ATOM	172	N	LEU	A	12	40.012	12.412	20.245	1.00	8.35
45	ATOM	174	CA	LEU	A	12	40.899	13.238	21.081	1.00	9.68
	ATOM	176	CB	LEU	A	12	41.501	14.374	20.257	1.00	10.19
	ATOM	179	CG	LEU	A	12	42.469	13.943	19.152	1.00	15.33
	ATOM	181	CD1	LEU	A	12	43.187	15.145	18.549	1.00	18.28
	ATOM	185	CD2	LEU	A	12	43.464	12.905	19.653	1.00	18.55
50	ATOM	189	C	LEU	A	12	40.242	13.812	22.351	1.00	9.19
	ATOM	190	O	LEU	A	12	40.851	13.776	23.445	1.00	10.13
	ATOM	191	N	VAL	A	13	39.010	14.301	22.232	1.00	8.92
	ATOM	193	CA	VAL	A	13	38.357	14.969	23.368	1.00	8.52
	ATOM	195	CB	VAL	A	13	38.013	16.426	23.050	1.00	8.78
55	ATOM	197	CG1	VAL	A	13	39.251	17.141	22.537	1.00	10.74
	ATOM	201	CG2	VAL	A	13	36.864	16.560	22.057	1.00	9.49
	ATOM	205	C	VAL	A	13	37.131	14.252	23.904	1.00	8.44
	ATOM	206	O	VAL	A	13	36.592	14.631	24.947	1.00	8.60
	ATOM	207	N	LYS	A	14	36.709	13.218	23.178	1.00	8.48
60	ATOM	209	CA	LYS	A	14	35.583	12.339	23.536	1.00	8.98
	ATOM	211	CB	LYS	A	14	35.771	11.687	24.909	1.00	8.33
	ATOM	214	CG	LYS	A	14	37.127	11.029	25.118	1.00	7.66
	ATOM	217	CD	LYS	A	14	37.513	10.044	23.992	1.00	8.44

	ATOM	220	CE	LYS	A	14	38.818	9.318	24.229	1.00	7.68	N
	ATOM	223	NZ	LYS	A	14	39.160	8.416	23.087	1.00	7.55	C
	ATOM	227	C	LYS	A	14	34.187	12.932	23.465	1.00	10.23	O
	ATOM	228	O	LYS	A	14	33.306	12.332	22.864	1.00	9.28	C
5	ATOM	229	N	LYS	A	15	33.976	14.083	24.089	1.00	10.78	N
	ATOM	231	CA	LYS	A	15	32.636	14.648	24.202	1.00	12.04	C
	ATOM	233	CB	LYS	A	15	32.058	14.428	25.615	1.00	13.87	C
	ATOM	236	CG	LYS	A	15	30.626	14.970	25.767	1.00	18.29	C
	ATOM	239	CD	LYS	A	15	30.411	15.838	26.991	1.00	25.35	C
10	ATOM	242	CE	LYS	A	15	29.648	17.144	26.648	1.00	26.80	C
	ATOM	245	NZ	LYS	A	15	30.479	18.398	26.848	1.00	28.04	N
	ATOM	249	C	LYS	A	15	32.701	16.124	23.876	1.00	11.99	C
	ATOM	250	O	LYS	A	15	33.603	16.825	24.333	1.00	12.92	O
	ATOM	251	N	ILE	A	16	31.770	16.587	23.054	1.00	11.71	N
15	ATOM	253	CA	ILE	A	16	31.631	18.011	22.795	1.00	11.45	C
	ATOM	255	CB	ILE	A	16	32.644	18.502	21.769	1.00	12.21	C
	ATOM	257	CG1	ILE	A	16	32.966	19.980	22.019	1.00	12.61	C
	ATOM	260	CD1	ILE	A	16	34.167	20.459	21.239	1.00	16.67	C
20	ATOM	264	CG2	ILE	A	16	32.154	18.226	20.357	1.00	12.62	C
	ATOM	268	C	ILE	A	16	30.193	18.273	22.375	1.00	11.19	C
	ATOM	269	O	ILE	A	16	29.515	17.396	21.835	1.00	10.05	O
	ATOM	270	N	ASP	A	17	29.729	19.495	22.614	1.00	11.77	N
	ATOM	272	CA	ASP	A	17	28.357	19.861	22.315	1.00	11.36	C
25	ATOM	274	CB	ASP	A	17	27.503	19.570	23.548	1.00	12.18	C
	ATOM	277	CG	ASP	A	17	26.019	19.854	23.363	1.00	13.83	C
	ATOM	278	OD1	ASP	A	17	25.558	20.190	22.262	1.00	14.93	O
	ATOM	279	OD2	ASP	A	17	25.207	19.726	24.327	1.00	17.34	O
	ATOM	280	C	ASP	A	17	28.354	21.342	22.018	1.00	10.94	C
30	ATOM	281	O	ASP	A	17	28.505	22.158	22.930	1.00	12.08	O
	ATOM	282	N	PHE	A	18	28.220	21.709	20.754	1.00	9.97	N
	ATOM	284	CA	PHE	A	18	28.208	23.121	20.420	1.00	9.42	C
	ATOM	286	CB	PHE	A	18	29.621	23.630	20.070	1.00	9.10	C
	ATOM	289	CG	PHE	A	18	30.262	22.990	18.849	1.00	9.30	C
35	ATOM	290	CD1	PHE	A	18	31.457	22.269	18.966	1.00	11.84	C
	ATOM	292	CE1	PHE	A	18	32.069	21.704	17.850	1.00	11.09	C
	ATOM	294	CZ	PHE	A	18	31.520	21.860	16.619	1.00	10.73	C
	ATOM	296	CE2	PHE	A	18	30.335	22.573	16.470	1.00	11.19	C
	ATOM	298	CD2	PHE	A	18	29.725	23.157	17.586	1.00	8.90	C
40	ATOM	300	C	PHE	A	18	27.226	23.431	19.299	1.00	9.78	C
	ATOM	301	O	PHE	A	18	26.794	22.537	18.568	1.00	9.84	O
	ATOM	302	N	ASP	A	19	26.899	24.711	19.156	1.00	10.97	N
	ATOM	304	CA	ASP	A	19	26.059	25.169	18.060	1.00	10.37	C
	ATOM	306	CB	ASP	A	19	24.575	25.130	18.429	1.00	10.87	C
45	ATOM	309	CG	ASP	A	19	23.674	25.452	17.267	1.00	11.55	C
	ATOM	310	OD1	ASP	A	19	24.180	25.843	16.178	1.00	11.30	O
	ATOM	311	OD2	ASP	A	19	22.418	25.322	17.350	1.00	12.10	O
	ATOM	312	C	ASP	A	19	26.497	26.590	17.705	1.00	10.71	C
	ATOM	313	O	ASP	A	19	26.136	27.575	18.388	1.00	10.19	O
50	ATOM	314	N	TYR	A	20	27.297	26.678	16.646	1.00	10.10	N
	ATOM	316	CA	TYR	A	20	27.788	27.942	16.103	1.00	9.68	C
	ATOM	318	CB	TYR	A	20	29.308	27.879	15.911	1.00	9.82	C
	ATOM	321	CG	TYR	A	20	30.089	28.043	17.181	1.00	8.36	C
	ATOM	322	CD1	TYR	A	20	30.459	26.943	17.934	1.00	9.01	C
55	ATOM	324	CE1	TYR	A	20	31.175	27.087	19.115	1.00	9.44	C
	ATOM	326	CZ	TYR	A	20	31.514	28.335	19.546	1.00	10.02	C
	ATOM	327	OH	TYR	A	20	32.228	28.469	20.703	1.00	9.07	O
	ATOM	329	CE2	TYR	A	20	31.167	29.441	18.804	1.00	10.02	C
	ATOM	331	CD2	TYR	A	20	30.451	29.303	17.648	1.00	8.62	C
60	ATOM	333	C	TYR	A	20	27.054	28.282	14.786	1.00	10.92	C
	ATOM	334	O	TYR	A	20	27.600	28.930	13.878	1.00	11.60	O
	ATOM	335	N	THR	A	21	25.800	27.857	14.694	1.00	12.18	N
	ATOM	337	CA	THR	A	21	24.980	28.261	13.567	1.00	12.37	C
	ATOM	339	CB	THR	A	21	23.584	27.692	13.676	1.00	12.82	C

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5	ATOM	341	OG1	THR A	21	23.623	26.259	13.737	1.00	12.95	O
	ATOM	343	CG2	THR A	21	22.832	27.997	12.401	1.00	13.70	C
	ATOM	347	C	THR A	21	24.871	29.776	13.598	1.00	12.58	C
	ATOM	348	O	THR A	21	24.445	30.332	14.595	1.00	12.57	O
5	ATOM	349	N	PRO A	22	25.259	30.460	12.528	1.00	12.83	C
	ATOM	350	CA	PRO A	22	25.263	31.917	12.549	1.00	12.54	C
	ATOM	352	CB	PRO A	22	26.214	32.276	11.409	1.00	12.71	C
	ATOM	355	CG	PRO A	22	26.064	31.150	10.423	1.00	12.51	C
	ATOM	358	CD	PRO A	22	25.773	29.925	11.259	1.00	12.22	C
0	ATOM	361	C	PRO A	22	23.890	32.509	12.337	1.00	12.87	C
	ATOM	362	O	PRO A	22	23.281	32.302	11.282	1.00	14.33	O
	ATOM	363	N	ASN A	23	23.405	33.202	13.363	1.00	12.69	N
	ATOM	365	CA	ASN A	23	22.145	33.920	13.285	1.00	12.96	C
	ATOM	367	CB	ASN A	23	21.290	33.568	14.497	1.00	13.22	C
15	ATOM	370	CG	ASN A	23	20.761	32.141	14.427	1.00	16.73	O
	ATOM	371	OD1	ASN A	23	19.705	31.904	13.821	1.00	22.06	N
	ATOM	372	ND2	ASN A	23	21.511	31.174	14.977	1.00	18.52	C
	ATOM	375	C	ASN A	23	22.449	35.415	13.208	1.00	12.31	C
	ATOM	376	O	ASN A	23	22.904	36.007	14.185	1.00	12.34	O
20	ATOM	377	N	TRP A	24	22.216	36.016	12.048	1.00	12.92	C
	ATOM	379	CA	TRP A	24	22.554	37.408	11.814	1.00	12.37	C
	ATOM	381	CB	TRP A	24	22.990	37.612	10.367	1.00	13.22	C
	ATOM	384	CG	TRP A	24	24.130	36.740	9.944	1.00	12.12	C
	ATOM	385	CD1	TRP A	24	24.039	35.556	9.279	1.00	11.86	C
25	ATOM	387	NE1	TRP A	24	25.292	35.046	9.042	1.00	13.92	N
	ATOM	389	CE2	TRP A	24	26.230	35.904	9.547	1.00	11.19	C
	ATOM	390	CD2	TRP A	24	25.536	36.989	10.123	1.00	10.96	C
	ATOM	391	CE3	TRP A	24	26.276	38.003	10.726	1.00	11.62	C
	ATOM	393	CZ3	TRP A	24	27.660	37.925	10.707	1.00	13.20	C
30	ATOM	395	CH2	TRP A	24	28.317	36.833	10.136	1.00	11.66	C
	ATOM	397	CZ2	TRP A	24	27.619	35.814	9.545	1.00	10.81	C
	ATOM	399	C	TRP A	24	21.343	38.268	12.120	1.00	12.73	C
	ATOM	400	O	TRP A	24	20.282	38.076	11.532	1.00	13.03	O
	ATOM	401	N	GLY A	25	21.488	39.222	13.029	1.00	12.00	C
35	ATOM	403	CA	GLY A	25	20.370	40.074	13.398	1.00	11.21	C
	ATOM	406	C	GLY A	25	20.495	41.423	12.706	1.00	11.71	C
	ATOM	407	O	GLY A	25	21.592	41.969	12.603	1.00	11.26	O
	ATOM	408	N	ARG A	26	19.375	41.957	12.233	1.00	11.79	N
	ATOM	410	CA	ARG A	26	19.388	43.192	11.486	1.00	12.49	C
40	ATOM	412	CB	ARG A	26	18.460	43.083	10.267	1.00	12.94	C
	ATOM	415	CG	ARG A	26	18.999	42.137	9.202	1.00	16.01	C
	ATOM	418	CD	ARG A	26	18.019	41.888	8.062	1.00	20.32	C
	ATOM	421	NE	ARG A	26	18.565	41.043	6.998	1.00	24.78	N
	ATOM	423	CZ	ARG A	26	19.426	41.460	6.071	1.00	25.04	N
45	ATOM	424	NH1	ARG A	26	19.860	40.607	5.149	1.00	29.16	N
	ATOM	427	NH2	ARG A	26	19.863	42.715	6.057	1.00	19.47	N
	ATOM	430	C	ARG A	26	19.010	44.365	12.357	1.00	12.60	C
	ATOM	431	O	ARG A	26	18.369	44.206	13.398	1.00	12.88	O
	ATOM	432	N	GLY A	27	19.411	45.549	11.917	1.00	12.77	C
50	ATOM	434	CA	GLY A	27	19.173	46.761	12.675	1.00	12.32	C
	ATOM	437	C	GLY A	27	18.090	47.628	12.071	1.00	13.21	C
	ATOM	438	O	GLY A	27	17.167	47.128	11.435	1.00	11.98	O
	ATOM	439	N	THR A	28	18.203	48.928	12.316	1.00	14.26	C
	ATOM	441	CA	THR A	28	17.261	49.925	11.819	1.00	14.60	C
55	ATOM	443	CB	THR A	28	16.523	50.576	13.006	1.00	13.94	C
	ATOM	445	OG1	THR A	28	15.801	49.590	13.761	1.00	12.38	O
	ATOM	447	CG2	THR A	28	15.460	51.569	12.517	1.00	14.06	C
	ATOM	451	C	THR A	28	18.039	51.002	11.041	1.00	15.60	C
	ATOM	452	O	THR A	28	18.823	51.756	11.636	1.00	15.37	O
60	ATOM	453	N	PRO A	29	17.874	51.082	9.718	1.00	17.62	C
	ATOM	454	CA	PRO A	29	17.025	50.182	8.928	1.00	17.21	C
	ATOM	456	CB	PRO A	29	16.956	50.887	7.570	1.00	17.70	C
	ATOM	459	CG	PRO A	29	18.211	51.657	7.483	1.00	17.48	C

	ATOM	582	CG2	THR	A	37	22.397	34.382	19.308	1.00	11.78	C
	ATOM	586	C	THR	A	37	24.484	33.647	17.365	1.00	10.36	C
	ATOM	587	O	THR	A	37	24.103	33.128	16.314	1.00	11.69	O
5	ATOM	588	N	PHE	A	38	25.183	33.002	18.288	1.00	10.33	N
	ATOM	590	CA	PHE	A	38	25.435	31.568	18.220	1.00	10.67	C
	ATOM	592	CB	PHE	A	38	26.892	31.285	18.520	1.00	10.97	C
	ATOM	595	CG	PHE	A	38	27.844	31.792	17.480	1.00	9.37	C
	ATOM	596	CD1	PHE	A	38	28.952	32.543	17.835	1.00	10.78	C
	ATOM	598	CE1	PHE	A	38	29.844	32.982	16.879	1.00	10.42	C
10	ATOM	600	CZ	PHE	A	38	29.659	32.667	15.590	1.00	12.03	C
	ATOM	602	CE2	PHE	A	38	28.559	31.898	15.215	1.00	10.68	C
	ATOM	604	CD2	PHE	A	38	27.660	31.476	16.146	1.00	10.68	C
	ATOM	606	C	PHE	A	38	24.595	30.912	19.303	1.00	11.13	C
15	ATOM	607	O	PHE	A	38	24.678	31.328	20.444	1.00	11.20	O
	ATOM	608	N	PRO	A	39	23.777	29.911	18.995	1.00	11.41	N
	ATOM	609	CA	PRO	A	39	22.920	29.317	20.033	1.00	11.03	C
	ATOM	611	CB	PRO	A	39	22.047	28.347	19.251	1.00	11.55	C
	ATOM	614	CG	PRO	A	39	22.138	28.792	17.827	1.00	11.41	C
20	ATOM	617	CD	PRO	A	39	23.501	29.337	17.671	1.00	10.69	C
	ATOM	620	C	PRO	A	39	23.593	28.585	21.186	1.00	10.78	C
	ATOM	621	O	PRO	A	39	23.007	28.537	22.272	1.00	11.13	O
	ATOM	622	N	LYS	A	40	24.756	27.986	20.961	1.00	10.03	N
	ATOM	624	CA	LYS	A	40	25.420	27.246	22.033	1.00	11.35	C
25	ATOM	626	CB	LYS	A	40	24.930	25.808	22.100	1.00	11.81	C
	ATOM	629	CG	LYS	A	40	25.329	25.153	23.413	1.00	15.47	C
	ATOM	632	CD	LYS	A	40	25.020	23.673	23.445	1.00	21.03	C
	ATOM	635	CE	LYS	A	40	25.654	23.024	24.665	1.00	26.85	C
	ATOM	638	NZ	LYS	A	40	24.928	23.362	25.917	1.00	35.22	N
30	ATOM	642	C	LYS	A	40	26.939	27.297	21.877	1.00	11.13	C
	ATOM	643	O	LYS	A	40	27.540	26.454	21.211	1.00	11.86	O
	ATOM	644	N	VAL	A	41	27.549	28.310	22.479	1.00	10.84	N
	ATOM	646	CA	VAL	A	41	28.995	28.462	22.410	1.00	10.68	C
	ATOM	648	CB	VAL	A	41	29.449	29.903	22.641	1.00	10.07	C
35	ATOM	650	CG1	VAL	A	41	28.907	30.826	21.533	1.00	10.33	C
	ATOM	654	CG2	VAL	A	41	29.040	30.419	24.007	1.00	10.45	C
	ATOM	658	C	VAL	A	41	29.690	27.564	23.425	1.00	11.85	C
	ATOM	659	O	VAL	A	41	29.093	27.111	24.425	1.00	12.38	O
	ATOM	660	N	LEU	A	42	30.957	27.305	23.165	1.00	13.03	N
40	ATOM	662	CA	LEU	A	42	31.803	26.664	24.159	1.00	15.12	C
	ATOM	664	CB	LEU	A	42	33.126	26.219	23.556	1.00	14.97	C
	ATOM	667	CG	LEU	A	42	32.873	25.139	22.491	1.00	15.42	C
	ATOM	669	CD1	LEU	A	42	34.128	24.763	21.705	1.00	16.85	C
	ATOM	673	CD2	LEU	A	42	32.303	23.917	23.125	1.00	17.26	C
45	ATOM	677	C	LEU	A	42	32.012	27.709	25.245	1.00	17.87	C
	ATOM	678	O	LEU	A	42	32.083	28.897	24.974	1.00	17.12	O
	ATOM	679	N	THR	A	43	32.171	27.279	26.476	1.00	21.75	N
	ATOM	681	CA	THR	A	43	32.188	28.272	27.549	1.00	24.61	C
	ATOM	683	CB	THR	A	43	30.761	28.365	28.043	1.00	24.82	C
50	ATOM	685	OG1	THR	A	43	29.883	29.292	27.424	1.00	27.15	O
	ATOM	687	CG2	THR	A	43	30.199	27.229	28.835	1.00	24.68	C
	ATOM	691	C	THR	A	43	33.197	27.863	28.620	1.00	26.54	C
	ATOM	692	O	THR	A	43	33.185	28.377	29.738	1.00	27.45	O
	ATOM	693	N	ASP	A	44	34.103	26.963	28.249	1.00	28.93	N
55	ATOM	695	CA	ASP	A	44	35.103	26.469	29.179	1.00	29.14	C
	ATOM	697	CB	ASP	A	44	35.855	25.271	28.602	1.00	28.74	C
	ATOM	700	CG	ASP	A	44	36.401	25.521	27.217	1.00	28.34	C
	ATOM	701	OD1	ASP	A	44	37.572	25.172	26.990	1.00	26.28	O
	ATOM	702	OD2	ASP	A	44	35.734	26.028	26.286	1.00	24.46	O
60	ATOM	703	C	ASP	A	44	36.063	27.575	29.547	1.00	30.53	C
	ATOM	704	O	ASP	A	44	36.513	27.663	30.699	1.00	30.19	O
	ATOM	705	N	LYS	A	45	36.372	28.422	28.568	1.00	31.95	N
	ATOM	707	CA	LYS	A	45	37.275	29.547	28.790	1.00	31.72	C
	ATOM	709	CB	LYS	A	45	38.701	29.244	28.320	1.00	31.90	C

	ATOM	842	N	VAL	A	52	36.074	41.685	11.206	1.00	14.90	N
	ATOM	844	CA	VAL	A	52	36.768	42.287	10.070	1.00	14.99	C
	ATOM	846	CB	VAL	A	52	37.834	41.307	9.534	1.00	15.36	C
	ATOM	848	CG1	VAL	A	52	38.577	41.908	8.360	1.00	15.94	C
5	ATOM	852	CG2	VAL	A	52	38.819	40.945	10.636	1.00	15.62	C
	ATOM	856	C	VAL	A	52	35.733	42.590	8.981	1.00	15.27	C
	ATOM	857	O	VAL	A	52	35.001	41.691	8.577	1.00	14.98	O
	ATOM	858	N	VAL	A	53	35.680	43.840	8.506	1.00	15.37	N
	ATOM	860	CA	VAL	A	53	34.663	44.255	7.542	1.00	16.36	C
	ATOM	862	CB	VAL	A	53	33.805	45.395	8.090	1.00	16.50	C
10	ATOM	864	CG1	VAL	A	53	32.827	45.905	7.043	1.00	16.92	C
	ATOM	868	CG2	VAL	A	53	33.037	44.923	9.314	1.00	16.68	C
	ATOM	872	C	VAL	A	53	35.366	44.712	6.284	1.00	17.90	C
	ATOM	873	O	VAL	A	53	36.121	45.670	6.321	1.00	17.86	O
	ATOM	874	N	ASN	A	54	35.099	44.024	5.182	1.00	19.87	N
	ATOM	876	CA	ASN	A	54	35.764	44.316	3.916	1.00	20.93	C
15	ATOM	878	CB	ASN	A	54	35.225	45.606	3.324	1.00	20.73	C
	ATOM	881	CG	ASN	A	54	33.946	45.408	2.504	1.00	20.64	C
	ATOM	882	OD1	ASN	A	54	33.395	46.382	1.976	1.00	22.37	O
	ATOM	883	ND2	ASN	A	54	33.474	44.168	2.388	1.00	18.46	N
	ATOM	886	C	ASN	A	54	37.281	44.421	4.100	1.00	22.08	C
	ATOM	887	O	ASN	A	54	37.924	45.291	3.513	1.00	22.88	O
20	ATOM	888	N	GLY	A	55	37.851	43.545	4.924	1.00	23.68	N
	ATOM	890	CA	GLY	A	55	39.288	43.532	5.134	1.00	22.59	C
	ATOM	893	C	GLY	A	55	39.767	44.478	6.212	1.00	22.03	C
	ATOM	894	O	GLY	A	55	40.936	44.441	6.586	1.00	22.03	O
	ATOM	895	N	SER	A	56	38.883	45.332	6.712	1.00	21.22	N
	ATOM	897	CA	SER	A	56	39.268	46.257	7.764	1.00	20.83	C
25	ATOM	899	CB	SER	A	56	38.434	47.521	7.666	1.00	21.16	C
	ATOM	902	OG	SER	A	56	38.925	48.496	8.556	1.00	24.04	O
	ATOM	904	C	SER	A	56	39.068	45.628	9.138	1.00	19.96	O
	ATOM	905	O	SER	A	56	37.961	45.229	9.477	1.00	18.84	O
	ATOM	906	N	ASP	A	57	40.129	45.590	9.937	1.00	19.21	N
	ATOM	908	CA	ASP	A	57	40.100	44.953	11.252	1.00	19.05	C
30	ATOM	910	CB	ASP	A	57	41.547	44.599	11.610	1.00	19.38	C
	ATOM	913	CG	ASP	A	57	41.704	43.926	12.947	1.00	20.67	C
	ATOM	914	OD1	ASP	A	57	40.717	43.476	13.545	1.00	19.91	O
	ATOM	915	OD2	ASP	A	57	42.833	43.786	13.472	1.00	25.20	O
	ATOM	916	C	ASP	A	57	39.483	45.908	12.263	1.00	18.66	C
	ATOM	917	O	ASP	A	57	40.031	46.992	12.524	1.00	17.62	O
35	ATOM	918	N	LEU	A	58	38.337	45.517	12.823	1.00	18.14	N
	ATOM	920	CA	LEU	A	58	37.660	46.339	13.821	1.00	17.44	C
	ATOM	922	CB	LEU	A	58	36.140	46.283	13.638	1.00	17.54	C
	ATOM	925	CG	LEU	A	58	35.587	46.711	12.271	1.00	18.21	C
	ATOM	927	CD1	LEU	A	58	34.067	46.915	12.314	1.00	18.79	C
	ATOM	931	CD2	LEU	A	58	36.271	47.970	11.777	1.00	20.33	C
40	ATOM	935	C	LEU	A	58	38.058	45.955	15.248	1.00	17.13	C
	ATOM	936	O	LEU	A	58	37.539	46.510	16.221	1.00	17.54	O
	ATOM	937	N	GLY	A	59	38.978	45.010	15.381	1.00	16.87	N
	ATOM	939	CA	GLY	A	59	39.503	44.667	16.686	1.00	16.43	C
	ATOM	942	C	GLY	A	59	38.781	43.524	17.361	1.00	16.14	C
	ATOM	943	O	GLY	A	59	37.953	42.845	16.768	1.00	13.91	O
45	ATOM	944	N	VAL	A	60	39.070	43.377	18.641	1.00	16.63	N
	ATOM	946	CA	VAL	A	60	38.664	42.216	19.409	1.00	16.80	C
	ATOM	948	CB	VAL	A	60	39.909	41.452	19.859	1.00	17.07	C
	ATOM	950	CG1	VAL	A	60	39.536	40.267	20.694	1.00	17.82	C
	ATOM	954	CG2	VAL	A	60	40.719	40.997	18.636	1.00	17.98	C
	ATOM	958	C	VAL	A	60	37.883	42.635	20.638	1.00	17.13	C
50	ATOM	959	O	VAL	A	60	38.254	43.594	21.331	1.00	17.22	O
	ATOM	960	N	GLU	A	61	36.806	41.913	20.913	1.00	16.81	N
	ATOM	962	CA	GLU	A	61	35.954	42.215	22.058	1.00	17.72	C
	ATOM	964	CB	GLU	A	61	34.759	43.060	21.623	1.00	18.26	C
	ATOM	967	CG	GLU	A	61	35.079	44.412	20.956	1.00	20.64	C

	ATOM	1087	CA	GLY	A	70	24.455	28.736	26.089	1.00	16.37	C
	ATOM	1090	C	GLY	A	70	24.635	29.701	24.941	1.00	14.59	C
	ATOM	1091	O	GLY	A	70	25.655	29.678	24.275	1.00	14.71	CON
5	ATOM	1092	N	GLY	A	71	23.655	30.564	24.707	1.00	12.55	CC
	ATOM	1094	CA	GLY	A	71	23.758	31.485	23.587	1.00	11.79	CC
	ATOM	1097	C	GLY	A	71	24.646	32.689	23.872	1.00	11.04	CC
	ATOM	1098	O	GLY	A	71	24.827	33.109	25.024	1.00	11.27	CC
	ATOM	1099	N	GLN	A	72	25.209	33.247	22.807	1.00	10.97	CC
10	ATOM	1101	CA	GLN	A	72	26.016	34.462	22.914	1.00	10.54	CC
	ATOM	1103	CB	GLN	A	72	27.497	34.125	23.115	1.00	10.98	CC
	ATOM	1106	CG	GLN	A	72	28.414	35.293	23.430	1.00	12.20	CC
	ATOM	1109	CD	GLN	A	72	29.834	34.862	23.853	1.00	15.65	CC
	ATOM	1110	OE1	GLN	A	72	30.449	35.487	24.742	1.00	17.66	CC
15	ATOM	1111	NE2	GLN	A	72	30.354	33.820	23.222	1.00	10.29	CC
	ATOM	1114	C	GLN	A	72	25.807	35.312	21.675	1.00	11.06	CC
	ATOM	1115	O	GLN	A	72	25.877	34.821	20.533	1.00	11.31	CC
	ATOM	1116	N	THR	A	73	25.535	36.589	21.904	1.00	10.95	CC
	ATOM	1118	CA	THR	A	73	25.337	37.526	20.830	1.00	9.80	CC
20	ATOM	1120	CB	THR	A	73	24.021	38.290	21.035	1.00	10.73	CCO
	ATOM	1122	OG1	THR	A	73	22.912	37.385	21.013	1.00	11.04	CC
	ATOM	1124	CG2	THR	A	73	23.786	39.270	19.891	1.00	10.78	CC
	ATOM	1128	C	THR	A	73	26.475	38.540	20.782	1.00	9.92	CC
	ATOM	1129	O	THR	A	73	26.722	39.283	21.745	1.00	10.19	CC
25	ATOM	1130	N	ILE	A	74	27.161	38.554	19.643	1.00	9.37	CC
	ATOM	1132	CA	ILE	A	74	28.232	39.493	19.364	1.00	10.19	CC
	ATOM	1134	CB	ILE	A	74	29.235	38.855	18.371	1.00	10.48	CC
	ATOM	1136	CG1	ILE	A	74	29.843	37.581	18.972	1.00	12.71	CC
	ATOM	1139	CD1	ILE	A	74	30.471	36.666	17.946	1.00	16.05	CC
30	ATOM	1143	CG2	ILE	A	74	30.296	39.860	17.986	1.00	10.70	CC
	ATOM	1147	C	ILE	A	74	27.609	40.733	18.756	1.00	10.18	CC
	ATOM	1148	O	ILE	A	74	27.052	40.677	17.660	1.00	11.08	CC
	ATOM	1149	N	ASN	A	75	27.674	41.851	19.489	1.00	9.17	CC
	ATOM	1151	CA	ASN	A	75	27.079	43.102	19.040	1.00	9.50	CC
35	ATOM	1153	CB	ASN	A	75	26.600	43.849	20.274	1.00	9.51	CC
	ATOM	1156	CG	ASN	A	75	25.994	45.177	19.950	1.00	10.45	CC
	ATOM	1157	OD1	ASN	A	75	25.558	45.424	18.827	1.00	9.62	CC
	ATOM	1158	ND2	ASN	A	75	25.931	46.046	20.959	1.00	12.30	CC
	ATOM	1161	C	ASN	A	75	28.050	43.975	18.248	1.00	9.58	CC
40	ATOM	1162	O	ASN	A	75	28.992	44.543	18.807	1.00	10.09	CC
	ATOM	1163	N	PHE	A	76	27.817	44.088	16.945	1.00	10.23	CC
	ATOM	1165	CA	PHE	A	76	28.751	44.809	16.087	1.00	10.31	CC
	ATOM	1167	CB	PHE	A	76	28.464	44.552	14.610	1.00	10.82	CC
	ATOM	1170	CG	PHE	A	76	28.596	43.096	14.199	1.00	11.07	CC
45	ATOM	1171	CD1	PHE	A	76	29.568	42.277	14.737	1.00	13.37	CC
	ATOM	1173	CE1	PHE	A	76	29.681	40.936	14.328	1.00	10.49	CC
	ATOM	1175	CZ	PHE	A	76	28.820	40.441	13.411	1.00	10.42	CC
	ATOM	1177	CE2	PHE	A	76	27.856	41.259	12.865	1.00	11.96	CC
	ATOM	1179	CD2	PHE	A	76	27.746	42.568	13.258	1.00	12.00	CC
50	ATOM	1181	C	PHE	A	76	28.780	46.301	16.409	1.00	10.53	CC
	ATOM	1182	O	PHE	A	76	29.743	46.978	16.059	1.00	10.34	CC
	ATOM	1183	N	LEU	A	77	27.746	46.826	17.073	1.00	10.19	CC
	ATOM	1185	CA	LEU	A	77	27.754	48.242	17.446	1.00	11.27	CC
	ATOM	1187	CB	LEU	A	77	26.443	48.652	18.120	1.00	11.26	CC
55	ATOM	1190	CG	LEU	A	77	25.267	48.913	17.154	1.00	12.41	CC
	ATOM	1192	CD1	LEU	A	77	24.989	47.774	16.232	1.00	12.55	CC
	ATOM	1196	CD2	LEU	A	77	23.977	49.223	17.911	1.00	13.90	CC
	ATOM	1200	C	LEU	A	77	28.933	48.577	18.368	1.00	12.07	CC
	ATOM	1201	O	LEU	A	77	29.399	49.717	18.371	1.00	13.10	CC
60	ATOM	1202	N	GLN	A	78	29.416	47.580	19.112	1.00	12.87	CC
	ATOM	1204	CA	GLN	A	78	30.562	47.741	20.011	1.00	13.30	CC
	ATOM	1206	CB	GLN	A	78	30.588	46.602	21.048	1.00	13.42	CC
	ATOM	1209	CG	GLN	A	78	29.408	46.690	22.022	1.00	14.19	CC
	ATOM	1212	CD	GLN	A	78	29.251	45.560	23.009	1.00	17.92	CC

	ATOM	1335	C	ALA	A	86	25.861	46.128	5.186	1.00	15.57
	ATOM	1336	O	ALA	A	86	26.941	45.575	5.059	1.00	15.01
	ATOM	1337	N	ASP	A	87	24.708	45.558	4.842	1.00	16.64
	ATOM	1339	CA	ASP	A	87	24.658	44.160	4.402	1.00	16.37
5	ATOM	1341	CB	ASP	A	87	23.253	43.550	4.495	1.00	16.92
	ATOM	1344	CG	ASP	A	87	22.293	44.088	3.472	1.00	17.15
	ATOM	1345	OD1	ASP	A	87	21.117	43.677	3.520	1.00	17.33
	ATOM	1346	OD2	ASP	A	87	22.615	44.920	2.605	1.00	17.89
10	ATOM	1347	C	ASP	A	87	25.316	43.899	3.046	1.00	16.95
	ATOM	1348	O	ASP	A	87	25.392	42.753	2.623	1.00	16.19
	ATOM	1349	N	THR	A	88	25.812	44.949	2.398	1.00	18.29
	ATOM	1351	CA	THR	A	88	26.566	44.803	1.146	1.00	17.50
	ATOM	1353	CB	THR	A	88	26.427	46.084	0.327	1.00	17.73
15	ATOM	1355	OG1	THR	A	88	26.702	47.225	1.150	1.00	16.82
	ATOM	1357	CG2	THR	A	88	25.020	46.269	-0.109	1.00	18.14
	ATOM	1361	C	THR	A	88	28.052	44.563	1.361	1.00	17.57
	ATOM	1362	O	THR	A	88	28.820	44.404	0.409	1.00	16.74
	ATOM	1363	N	LYS	A	89	28.477	44.594	2.609	1.00	17.46
20	ATOM	1365	CA	LYS	A	89	29.871	44.389	2.919	1.00	17.25
	ATOM	1367	CB	LYS	A	89	30.312	45.388	3.978	1.00	17.22
	ATOM	1370	CG	LYS	A	89	30.058	46.844	3.579	1.00	18.92
	ATOM	1373	CD	LYS	A	89	30.818	47.788	4.471	1.00	22.27
	ATOM	1376	CE	LYS	A	89	30.590	49.242	4.055	1.00	25.20
25	ATOM	1379	NZ	LYS	A	89	31.208	50.160	5.042	1.00	29.86
	ATOM	1383	C	LYS	A	89	30.069	42.968	3.411	1.00	17.09
	ATOM	1384	O	LYS	A	89	29.122	42.311	3.818	1.00	16.84
	ATOM	1385	N	THR	A	90	31.300	42.493	3.343	1.00	17.10
	ATOM	1387	CA	THR	A	90	31.662	41.181	3.842	1.00	16.27
30	ATOM	1389	CB	THR	A	90	32.860	40.644	3.086	1.00	17.13
	ATOM	1391	OG1	THR	A	90	32.533	40.515	1.704	1.00	15.79
	ATOM	1393	CG2	THR	A	90	33.199	39.226	3.543	1.00	17.35
	ATOM	1397	C	THR	A	90	32.068	41.322	5.296	1.00	15.60
	ATOM	1398	O	THR	A	90	32.930	42.137	5.613	1.00	14.66
35	ATOM	1399	N	ILE	A	91	31.451	40.543	6.170	1.00	15.11
	ATOM	1401	CA	ILE	A	91	31.823	40.561	7.577	1.00	14.05
	ATOM	1403	CB	ILE	A	91	30.596	40.777	8.475	1.00	14.05
	ATOM	1405	CG1	ILE	A	91	29.771	41.971	7.995	1.00	13.77
	ATOM	1408	CD1	ILE	A	91	28.482	42.119	8.725	1.00	15.25
40	ATOM	1412	CG2	ILE	A	91	31.039	40.924	9.949	1.00	13.82
	ATOM	1416	C	ILE	A	91	32.435	39.221	7.914	1.00	14.03
	ATOM	1417	O	ILE	A	91	31.795	38.191	7.702	1.00	14.85
	ATOM	1418	N	GLN	A	92	33.679	39.230	8.382	1.00	12.86
	ATOM	1420	CA	GLN	A	92	34.298	38.028	8.919	1.00	13.25
45	ATOM	1422	CB	GLN	A	92	35.678	37.818	8.338	1.00	14.29
	ATOM	1425	CG	GLN	A	92	35.645	37.428	6.904	1.00	16.51
	ATOM	1428	CD	GLN	A	92	37.020	37.515	6.275	1.00	21.30
	ATOM	1429	OE1	GLN	A	92	37.536	36.517	5.775	1.00	25.59
	ATOM	1430	NE2	GLN	A	92	37.627	38.701	6.319	1.00	23.68
50	ATOM	1433	C	GLN	A	92	34.443	38.120	10.423	1.00	12.74
	ATOM	1434	O	GLN	A	92	34.914	39.127	10.940	1.00	12.27
	ATOM	1435	N	VAL	A	93	34.072	37.051	11.115	1.00	11.90
	ATOM	1437	CA	VAL	A	93	34.217	36.985	12.564	1.00	11.73
	ATOM	1439	CB	VAL	A	93	32.865	36.841	13.257	1.00	11.43
55	ATOM	1441	CG1	VAL	A	93	33.048	36.856	14.771	1.00	12.22
	ATOM	1445	CG2	VAL	A	93	31.925	37.956	12.809	1.00	12.10
	ATOM	1449	C	VAL	A	93	35.118	35.797	12.912	1.00	11.97
	ATOM	1450	O	VAL	A	93	34.953	34.707	12.379	1.00	11.23
	ATOM	1451	N	PHE	A	94	36.096	36.055	13.773	1.00	11.48
60	ATOM	1453	CA	PHE	A	94	37.069	35.064	14.188	1.00	12.22
	ATOM	1455	CB	PHE	A	94	38.473	35.563	13.871	1.00	12.62
	ATOM	1458	CG	PHE	A	94	38.736	35.743	12.404	1.00	12.77
	ATOM	1459	CD1	PHE	A	94	38.345	36.900	11.763	1.00	14.64
	ATOM	1461	CE1	PHE	A	94	38.598	37.083	10.420	1.00	16.13

	ATOM	1587	CB	SER A	103	49.068	35.070	18.214	1.00	31.29	C
	ATOM	1590	OG	SER A	103	49.175	34.552	19.532	1.00	32.26	O
	ATOM	1592	C	SER A	103	46.981	34.072	17.315	1.00	28.93	C
5	ATOM	1593	O	SER A	103	47.135	33.752	16.140	1.00	29.01	O
	ATOM	1594	N	GLU A	104	46.308	33.320	18.173	1.00	26.74	N
	ATOM	1596	CA	GLU A	104	45.648	32.098	17.739	1.00	23.55	C
	ATOM	1598	CB	GLU A	104	45.821	30.969	18.759	1.00	23.10	C
	ATOM	1601	CG	GLU A	104	45.217	29.652	18.294	1.00	22.29	C
10	ATOM	1604	CD	GLU A	104	45.267	28.539	19.335	1.00	20.79	C
	ATOM	1605	OE1	GLU A	104	44.705	27.459	19.063	1.00	18.27	O
	ATOM	1606	OE2	GLU A	104	45.872	28.735	20.405	1.00	19.73	O
	ATOM	1607	C	GLU A	104	44.166	32.431	17.527	1.00	21.26	C
	ATOM	1608	O	GLU A	104	43.463	32.788	18.468	1.00	20.20	O
15	ATOM	1609	N	GLU A	105	43.706	32.342	16.286	1.00	19.27	N
	ATOM	1611	CA	GLU A	105	42.310	32.652	15.989	1.00	17.92	C
	ATOM	1613	CB	GLU A	105	42.119	34.141	15.658	1.00	18.34	C
	ATOM	1616	CG	GLU A	105	42.614	34.515	14.283	1.00	19.41	C
	ATOM	1619	CD	GLU A	105	42.443	35.986	13.960	1.00	21.46	C
20	ATOM	1620	OE1	GLU A	105	42.657	36.346	12.779	1.00	22.55	O
	ATOM	1621	OE2	GLU A	105	42.097	36.770	14.872	1.00	19.79	O
	ATOM	1622	C	GLU A	105	41.807	31.788	14.851	1.00	16.31	C
	ATOM	1623	O	GLU A	105	42.589	31.268	14.050	1.00	16.35	O
	ATOM	1624	N	TYR A	106	40.489	31.642	14.779	1.00	14.92	N
25	ATOM	1626	CA	TYR A	106	39.856	30.789	13.784	1.00	13.70	C
	ATOM	1628	CB	TYR A	106	39.466	29.416	14.400	1.00	12.91	C
	ATOM	1631	CG	TYR A	106	40.630	28.705	15.037	1.00	13.45	C
	ATOM	1632	CD1	TYR A	106	41.441	27.863	14.296	1.00	15.31	C
	ATOM	1634	CE1	TYR A	106	42.517	27.226	14.869	1.00	15.32	C
30	ATOM	1636	CZ	TYR A	106	42.812	27.428	16.186	1.00	15.25	C
	ATOM	1637	OH	TYR A	106	43.904	26.776	16.728	1.00	16.78	O
	ATOM	1639	CE2	TYR A	106	42.027	28.251	16.961	1.00	15.05	C
	ATOM	1641	CD2	TYR A	106	40.934	28.890	16.379	1.00	13.75	C
	ATOM	1643	C	TYR A	106	38.605	31.460	13.230	1.00	13.16	C
35	ATOM	1644	O	TYR A	106	37.789	31.993	14.001	1.00	12.90	O
	ATOM	1645	N	ILE A	107	38.432	31.416	11.911	1.00	12.87	N
	ATOM	1647	CA	ILE A	107	37.219	31.954	11.296	1.00	13.04	C
	ATOM	1649	CB	ILE A	107	37.271	31.865	9.734	1.00	13.61	C
	ATOM	1651	CG1	ILE A	107	36.049	32.531	9.105	1.00	16.08	C
40	ATOM	1654	CD1	ILE A	107	36.054	33.996	9.131	1.00	19.25	C
	ATOM	1658	CG2	ILE A	107	37.277	30.431	9.234	1.00	14.39	C
	ATOM	1662	C	ILE A	107	36.026	31.203	11.890	1.00	12.69	C
	ATOM	1663	O	ILE A	107	36.050	29.967	11.991	1.00	12.26	O
	ATOM	1664	N	ILE A	108	34.994	31.931	12.314	1.00	10.96	N
45	ATOM	1666	CA	ILE A	108	33.831	31.283	12.892	1.00	11.58	C
	ATOM	1668	CB	ILE A	108	33.823	31.470	14.438	1.00	11.47	C
	ATOM	1670	CG1	ILE A	108	32.825	30.527	15.117	1.00	12.45	C
	ATOM	1673	CD1	ILE A	108	33.138	29.042	14.913	1.00	14.07	C
	ATOM	1677	CG2	ILE A	108	33.541	32.903	14.825	1.00	11.20	C
50	ATOM	1681	C	ILE A	108	32.516	31.695	12.234	1.00	11.67	C
	ATOM	1682	O	ILE A	108	31.510	31.041	12.437	1.00	13.19	O
	ATOM	1683	N	ALA A	109	32.512	32.756	11.438	1.00	11.35	N
	ATOM	1685	CA	ALA A	109	31.319	33.139	10.675	1.00	12.23	C
	ATOM	1687	CB	ALA A	109	30.290	33.798	11.582	1.00	12.13	C
55	ATOM	1691	C	ALA A	109	31.699	34.113	9.557	1.00	12.08	C
	ATOM	1692	O	ALA A	109	32.648	34.879	9.714	1.00	12.05	O
	ATOM	1693	N	GLU A	110	30.956	34.090	8.448	1.00	13.59	N
	ATOM	1695	CA	GLU A	110	31.147	35.085	7.399	1.00	13.30	C
	ATOM	1697	CB	GLU A	110	32.149	34.594	6.336	1.00	14.11	C
60	ATOM	1700	CG	GLU A	110	32.258	35.509	5.123	1.00	15.16	C
	ATOM	1703	CD	GLU A	110	33.187	34.947	4.059	1.00	18.76	C
	ATOM	1704	OE1	GLU A	110	34.393	35.270	4.085	1.00	20.47	O
	ATOM	1705	OE2	GLU A	110	32.706	34.165	3.204	1.00	21.90	O
	ATOM	1706	C	GLU A	110	29.814	35.470	6.762	1.00	13.12	C

	ATOM	1835	C	SER	B	4	41.042	22.939	15.427	1.00	11.99	C
	ATOM	1836	O	SER	B	4	40.232	23.408	14.613	1.00	11.15	O
	ATOM	1837	N	LEU	B	5	41.045	21.652	15.755	1.00	11.62	N
5	ATOM	1839	CA	LEU	B	5	40.036	20.744	15.224	1.00	10.95	C
	ATOM	1841	CB	LEU	B	5	40.253	19.326	15.755	1.00	11.58	C
	ATOM	1844	CG	LEU	B	5	41.493	18.602	15.191	1.00	13.26	C
	ATOM	1846	CD1	LEU	B	5	41.671	17.274	15.878	1.00	14.30	C
	ATOM	1850	CD2	LEU	B	5	41.430	18.388	13.679	1.00	16.20	C
10	ATOM	1854	C	LEU	B	5	38.633	21.207	15.621	1.00	10.05	C
	ATOM	1855	O	LEU	B	5	37.713	21.115	14.830	1.00	9.60	O
	ATOM	1856	N	THR	B	6	38.482	21.669	16.858	1.00	9.83	N
	ATOM	1858	CA	THR	B	6	37.187	22.108	17.359	1.00	10.01	C
	ATOM	1860	CB	THR	B	6	37.300	22.622	18.794	1.00	9.92	C
15	ATOM	1862	OG1	THR	B	6	37.622	21.536	19.681	1.00	10.43	O
	ATOM	1864	CG2	THR	B	6	35.965	23.168	19.289	1.00	9.67	C
	ATOM	1868	C	THR	B	6	36.616	23.197	16.490	1.00	10.19	C
	ATOM	1869	O	THR	B	6	35.478	23.121	16.047	1.00	10.13	O
	ATOM	1870	N	PHE	B	7	37.416	24.217	16.232	1.00	10.95	N
20	ATOM	1872	CA	PHE	B	7	36.898	25.372	15.532	1.00	10.37	C
	ATOM	1874	CB	PHE	B	7	37.576	26.643	16.024	1.00	10.33	C
	ATOM	1877	CG	PHE	B	7	37.149	27.021	17.415	1.00	10.12	C
	ATOM	1878	CD1	PHE	B	7	35.833	27.366	17.673	1.00	10.88	C
	ATOM	1880	CE1	PHE	B	7	35.417	27.659	18.945	1.00	11.19	C
25	ATOM	1882	CZ	PHE	B	7	36.296	27.605	19.969	1.00	11.18	C
	ATOM	1884	CE2	PHE	B	7	37.605	27.245	19.734	1.00	12.59	C
	ATOM	1886	CD2	PHE	B	7	38.021	26.936	18.466	1.00	11.83	C
	ATOM	1888	C	PHE	B	7	36.909	25.194	14.025	1.00	10.94	C
	ATOM	1889	O	PHE	B	7	36.103	25.820	13.353	1.00	11.93	O
30	ATOM	1890	N	GLN	B	8	37.767	24.329	13.489	1.00	11.93	N
	ATOM	1892	CA	GLN	B	8	37.647	24.010	12.067	1.00	11.34	C
	ATOM	1894	CB	GLN	B	8	38.761	23.087	11.621	1.00	12.48	C
	ATOM	1897	CG	GLN	B	8	40.113	23.720	11.528	1.00	14.22	C
	ATOM	1900	CD	GLN	B	8	41.117	22.698	11.051	1.00	15.81	C
35	ATOM	1901	OE1	GLN	B	8	42.036	22.331	11.781	1.00	19.91	O
	ATOM	1902	NE2	GLN	B	8	40.902	22.184	9.843	1.00	17.92	N
	ATOM	1905	C	GLN	B	8	36.316	23.286	11.855	1.00	10.37	C
	ATOM	1906	O	GLN	B	8	35.580	23.546	10.908	1.00	10.98	O
	ATOM	1907	N	LEU	B	9	36.006	22.360	12.758	1.00	10.06	N
40	ATOM	1909	CA	LEU	B	9	34.757	21.608	12.648	1.00	9.71	C
	ATOM	1911	CB	LEU	B	9	34.726	20.455	13.634	1.00	9.51	C
	ATOM	1914	CG	LEU	B	9	33.493	19.574	13.606	1.00	10.41	C
	ATOM	1916	CD1	LEU	B	9	33.447	18.825	12.265	1.00	11.23	C
	ATOM	1920	CD2	LEU	B	9	33.561	18.587	14.753	1.00	9.36	C
45	ATOM	1924	C	LEU	B	9	33.552	22.498	12.880	1.00	9.33	C
	ATOM	1925	O	LEU	B	9	32.566	22.409	12.160	1.00	10.21	O
	ATOM	1926	N	ALA	B	10	33.618	23.376	13.874	1.00	9.78	N
	ATOM	1928	CA	ALA	B	10	32.476	24.246	14.138	1.00	9.75	C
	ATOM	1930	CB	ALA	B	10	32.727	25.091	15.353	1.00	10.03	C
50	ATOM	1934	C	ALA	B	10	32.145	25.126	12.919	1.00	9.57	C
	ATOM	1935	O	ALA	B	10	30.982	25.275	12.554	1.00	8.63	O
	ATOM	1936	N	TYR	B	11	33.155	25.688	12.269	1.00	9.75	N
	ATOM	1938	CA	TYR	B	11	32.885	26.566	11.136	1.00	10.72	C
	ATOM	1940	CB	TYR	B	11	34.159	27.237	10.688	1.00	10.48	C
55	ATOM	1943	CG	TYR	B	11	33.979	28.188	9.535	1.00	11.27	C
	ATOM	1944	CD1	TYR	B	11	34.664	27.988	8.352	1.00	11.30	C
	ATOM	1946	CE1	TYR	B	11	34.534	28.867	7.292	1.00	13.85	C
	ATOM	1948	CZ	TYR	B	11	33.706	29.949	7.409	1.00	14.97	C
	ATOM	1949	OH	TYR	B	11	33.579	30.827	6.350	1.00	16.32	O
60	ATOM	1951	CE2	TYR	B	11	33.022	30.183	8.582	1.00	14.29	C
	ATOM	1953	CD2	TYR	B	11	33.159	29.300	9.640	1.00	12.05	C
	ATOM	1955	C	TYR	B	11	32.250	25.801	9.987	1.00	11.09	C
	ATOM	1956	O	TYR	B	11	31.380	26.310	9.272	1.00	11.19	O
	ATOM	1957	N	LEU	B	12	32.672	24.556	9.823	1.00	12.21	N

	ATOM	1959	CA	LEU	B	12	32.179	23.730	8.742	1.00	12.45
	ATOM	1961	CB	LEU	B	12	33.187	22.614	8.470	1.00	12.95
	ATOM	1964	CG	LEU	B	12	33.011	21.779	7.209	1.00	16.64
	ATOM	1966	CD1	LEU	B	12	32.907	22.662	5.962	1.00	17.27
5	ATOM	1970	CD2	LEU	B	12	34.193	20.804	7.093	1.00	18.06
	ATOM	1974	C	LEU	B	12	30.767	23.156	8.956	1.00	12.37
	ATOM	1975	O	LEU	B	12	29.914	23.302	8.071	1.00	13.09
	ATOM	1976	N	VAL	B	13	30.514	22.525	10.103	1.00	12.47
10	ATOM	1978	CA	VAL	B	13	29.256	21.805	10.335	1.00	12.51
	ATOM	1980	CB	VAL	B	13	29.485	20.381	10.894	1.00	13.37
	ATOM	1982	CG1	VAL	B	13	30.492	19.620	10.042	1.00	15.30
	ATOM	1986	CG2	VAL	B	13	29.922	20.398	12.336	1.00	14.08
	ATOM	1990	C	VAL	B	13	28.273	22.541	11.237	1.00	12.00
15	ATOM	1991	O	VAL	B	13	27.106	22.172	11.319	1.00	11.68
	ATOM	1992	N	LYS	B	14	28.775	23.558	11.917	1.00	11.40
	ATOM	1994	CA	LYS	B	14	27.991	24.459	12.764	1.00	11.19
	ATOM	1996	CB	LYS	B	14	26.759	25.042	12.035	1.00	11.48
	ATOM	1999	CG	LYS	B	14	27.039	25.761	10.731	1.00	11.14
20	ATOM	2002	CD	LYS	B	14	28.190	26.750	10.824	1.00	9.92
	ATOM	2005	CE	LYS	B	14	28.422	27.540	9.526	1.00	10.31
	ATOM	2008	NZ	LYS	B	14	29.602	28.448	9.653	1.00	12.24
	ATOM	2012	C	LYS	B	14	27.549	23.850	14.104	1.00	10.98
	ATOM	2013	O	LYS	B	14	27.795	24.432	15.165	1.00	10.55
25	ATOM	2014	N	LYS	B	15	26.914	22.684	14.069	1.00	10.86
	ATOM	2016	CA	LYS	B	15	26.282	22.127	15.256	1.00	12.66
	ATOM	2018	CB	LYS	B	15	24.753	22.337	15.149	1.00	13.78
	ATOM	2021	CG	LYS	B	15	23.878	21.739	16.237	1.00	18.00
	ATOM	2024	CD	LYS	B	15	22.382	22.146	16.068	1.00	23.19
30	ATOM	2027	CE	LYS	B	15	21.671	21.394	14.921	1.00	27.17
	ATOM	2030	NZ	LYS	B	15	20.279	21.902	14.547	1.00	34.39
	ATOM	2034	C	LYS	B	15	26.637	20.640	15.355	1.00	12.47
	ATOM	2035	O	LYS	B	15	26.526	19.913	14.363	1.00	13.03
	ATOM	2036	N	ILE	B	16	27.094	20.207	16.526	1.00	13.16
35	ATOM	2038	CA	ILE	B	16	27.368	18.784	16.760	1.00	12.11
	ATOM	2040	CB	ILE	B	16	28.707	18.380	16.149	1.00	12.61
	ATOM	2042	CG1	ILE	B	16	28.660	16.894	15.756	1.00	13.09
	ATOM	2045	CD1	ILE	B	16	29.822	16.447	14.941	1.00	14.91
	ATOM	2049	CG2	ILE	B	16	29.831	18.704	17.105	1.00	12.04
40	ATOM	2053	C	ILE	B	16	27.276	18.487	18.258	1.00	12.09
	ATOM	2054	O	ILE	B	16	27.516	19.360	19.098	1.00	11.30
	ATOM	2055	N	ASP	B	17	26.903	17.257	18.587	1.00	11.37
	ATOM	2057	CA	ASP	B	17	26.701	16.850	19.977	1.00	11.97
	ATOM	2059	CB	ASP	B	17	25.238	17.060	20.347	1.00	12.19
45	ATOM	2062	CG	ASP	B	17	24.929	16.770	21.795	1.00	15.47
	ATOM	2063	OD1	ASP	B	17	25.834	16.488	22.602	1.00	15.71
	ATOM	2064	OD2	ASP	B	17	23.746	16.838	22.212	1.00	21.94
	ATOM	2065	C	ASP	B	17	27.026	15.373	20.040	1.00	11.79
	ATOM	2066	O	ASP	B	17	26.246	14.552	19.554	1.00	12.85
50	ATOM	2067	N	PHE	B	18	28.190	15.029	20.566	1.00	10.57
	ATOM	2069	CA	PHE	B	18	28.552	13.620	20.686	1.00	10.47
	ATOM	2071	CB	PHE	B	18	29.385	13.115	19.479	1.00	9.71
	ATOM	2074	CG	PHE	B	18	30.728	13.797	19.316	1.00	9.88
	ATOM	2075	CD1	PHE	B	18	31.732	13.663	20.275	1.00	7.77
	ATOM	2077	CE1	PHE	B	18	32.936	14.318	20.131	1.00	9.85
55	ATOM	2079	CZ	PHE	B	18	33.172	15.094	19.013	1.00	10.47
	ATOM	2081	CE2	PHE	B	18	32.194	15.226	18.062	1.00	9.60
	ATOM	2083	CD2	PHE	B	18	30.979	14.584	18.211	1.00	9.21
	ATOM	2085	C	PHE	B	18	29.281	13.324	21.983	1.00	9.72
	ATOM	2086	O	PHE	B	18	29.760	14.220	22.691	1.00	10.09
60	ATOM	2087	N	ASP	B	19	29.326	12.041	22.306	1.00	9.08
	ATOM	2089	CA	ASP	B	19	30.050	11.562	23.459	1.00	9.72
	ATOM	2091	CB	ASP	B	19	29.193	11.549	24.716	1.00	9.54
	ATOM	2094	CG	ASP	B	19	29.999	11.225	25.937	1.00	11.36

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5	ATOM	2095	OD1	ASP	B	19	29.498	11.408	27.093	1.00	13.80
	ATOM	2096	OD2	ASP	B	19	31.149	10.767	25.834	1.00	10.64
	ATOM	2097	C	ASP	B	19	30.543	10.155	23.160	1.00	9.57
	ATOM	2098	O	ASP	B	19	29.765	9.177	23.218	1.00	10.53
	ATOM	2099	N	TYR	B	20	31.830	10.085	22.813	1.00	9.01
	ATOM	2101	CA	TYR	B	20	32.505	8.831	22.518	1.00	8.11
	ATOM	2103	CB	TYR	B	20	33.232	8.896	21.163	1.00	7.80
	ATOM	2106	CG	TYR	B	20	32.292	8.800	19.966	1.00	7.18
	ATOM	2107	CD1	TYR	B	20	31.765	9.933	19.381	1.00	8.23
10	ATOM	2109	CE1	TYR	B	20	30.897	9.849	18.286	1.00	8.63
	ATOM	2111	CZ	TYR	B	20	30.570	8.623	17.777	1.00	8.76
	ATOM	2112	OH	TYR	B	20	29.690	8.504	16.698	1.00	7.11
	ATOM	2114	CE2	TYR	B	20	31.081	7.492	18.358	1.00	7.59
	ATOM	2116	CD2	TYR	B	20	31.929	7.580	19.435	1.00	8.21
15	ATOM	2118	C	TYR	B	20	33.453	8.446	23.657	1.00	7.82
	ATOM	2119	O	TYR	B	20	34.453	7.777	23.416	1.00	8.72
	ATOM	2120	N	THR	B	21	33.128	8.834	24.892	1.00	8.34
	ATOM	2122	CA	THR	B	21	33.897	8.365	26.046	1.00	8.81
	ATOM	2124	CB	THR	B	21	33.320	8.922	27.334	1.00	9.96
20	ATOM	2126	OG1	THR	B	21	33.372	10.363	27.299	1.00	10.43
	ATOM	2128	CG2	THR	B	21	34.222	8.512	28.491	1.00	9.86
	ATOM	2132	C	THR	B	21	33.804	6.831	26.040	1.00	9.35
	ATOM	2133	O	THR	B	21	32.692	6.291	26.011	1.00	9.22
	ATOM	2134	N	PRO	B	22	34.922	6.111	26.025	1.00	9.19
25	ATOM	2135	CA	PRO	B	22	34.844	4.647	25.905	1.00	9.50
	ATOM	2137	CB	PRO	B	22	36.209	4.280	25.352	1.00	9.82
	ATOM	2140	CG	PRO	B	22	37.138	5.334	25.941	1.00	9.26
	ATOM	2143	CD	PRO	B	22	36.320	6.589	26.015	1.00	9.34
	ATOM	2146	C	PRO	B	22	34.616	3.930	27.227	1.00	10.16
30	ATOM	2147	O	PRO	B	22	35.520	3.898	28.070	1.00	10.91
	ATOM	2148	N	ASN	B	23	33.413	3.394	27.413	1.00	10.40
	ATOM	2150	CA	ASN	B	23	33.082	2.645	28.614	1.00	10.53
	ATOM	2152	CB	ASN	B	23	31.680	3.014	29.089	1.00	11.14
	ATOM	2155	CG	ASN	B	23	31.595	4.472	29.590	1.00	13.44
35	ATOM	2156	OD1	ASN	B	23	31.816	4.721	30.763	1.00	19.39
	ATOM	2157	ND2	ASN	B	23	31.342	5.441	28.685	1.00	14.28
	ATOM	2160	C	ASN	B	23	33.228	1.143	28.312	1.00	9.85
	ATOM	2161	O	ASN	B	23	32.489	0.595	27.483	1.00	10.11
	ATOM	2162	N	TRP	B	24	34.208	0.502	28.942	1.00	8.96
40	ATOM	2164	CA	TRP	B	24	34.524	-0.899	28.684	1.00	8.99
	ATOM	2166	CB	TRP	B	24	36.032	-1.117	28.760	1.00	8.98
	ATOM	2169	CG	TRP	B	24	36.799	-0.256	27.823	1.00	8.22
	ATOM	2170	CD1	TRP	B	24	37.375	0.965	28.116	1.00	9.52
	ATOM	2172	NE1	TRP	B	24	38.003	1.469	27.000	1.00	9.37
45	ATOM	2174	CE2	TRP	B	24	37.860	0.574	25.967	1.00	9.60
	ATOM	2175	CD2	TRP	B	24	37.099	-0.517	26.446	1.00	9.21
	ATOM	2176	CE3	TRP	B	24	36.824	-1.577	25.560	1.00	9.09
	ATOM	2178	CZ3	TRP	B	24	37.292	-1.502	24.264	1.00	9.03
	ATOM	2180	CH2	TRP	B	24	38.045	-0.414	23.829	1.00	7.76
50	ATOM	2182	CZ2	TRP	B	24	38.337	0.636	24.661	1.00	8.73
	ATOM	2184	C	TRP	B	24	33.806	-1.771	29.715	1.00	10.09
	ATOM	2185	O	TRP	B	24	34.024	-1.610	30.922	1.00	11.35
	ATOM	2186	N	GLY	B	25	32.931	-2.655	29.234	1.00	10.43
	ATOM	2188	CA	GLY	B	25	32.143	-3.516	30.098	1.00	9.95
55	ATOM	2191	C	GLY	B	25	32.749	-4.894	30.176	1.00	10.10
	ATOM	2192	O	GLY	B	25	33.220	-5.444	29.184	1.00	10.44
	ATOM	2193	N	ARG	B	26	32.721	-5.470	31.372	1.00	9.91
	ATOM	2195	CA	ARG	B	26	33.393	-6.738	31.594	1.00	10.74
	ATOM	2197	CB	ARG	B	26	34.185	-6.692	32.897	1.00	11.28
60	ATOM	2200	CG	ARG	B	26	35.418	-5.792	32.764	1.00	14.61
	ATOM	2203	CD	ARG	B	26	36.168	-5.505	34.025	1.00	18.47
	ATOM	2206	NE	ARG	B	26	37.353	-4.694	33.713	1.00	22.81
	ATOM	2208	CZ	ARG	B	26	38.526	-5.167	33.274	1.00	20.92

	ATOM	2328	CA	ASN	B	35	31.791	-4.730	25.152	1.00	8.99	C
	ATOM	2330	CB	ASN	B	35	30.278	-4.762	25.446	1.00	9.72	C
	ATOM	2333	CG	ASN	B	35	29.970	-4.723	26.949	1.00	11.37	C
	ATOM	2334	OD1	ASN	B	35	29.559	-5.741	27.574	1.00	14.30	C
5	ATOM	2335	ND2	ASN	B	35	30.186	-3.573	27.551	1.00	7.97	CON
	ATOM	2338	C	ASN	B	35	32.351	-3.341	25.400	1.00	9.17	C
	ATOM	2339	O	ASN	B	35	33.129	-3.115	26.332	1.00	9.29	C
	ATOM	2340	N	LEU	B	36	31.917	-2.413	24.552	1.00	9.41	N
	ATOM	2342	CA	LEU	B	36	32.345	-1.026	24.581	1.00	9.12	CC
10	ATOM	2344	CB	LEU	B	36	33.308	-0.779	23.414	1.00	8.17	CC
	ATOM	2347	CG	LEU	B	36	33.652	0.670	23.053	1.00	9.08	C
	ATOM	2349	CD1	LEU	B	36	34.294	1.387	24.199	1.00	9.44	C
	ATOM	2353	CD2	LEU	B	36	34.560	0.729	21.837	1.00	10.64	C
	ATOM	2357	C	LEU	B	36	31.099	-0.186	24.369	1.00	9.10	C
15	ATOM	2358	O	LEU	B	36	30.382	-0.391	23.385	1.00	8.92	CO
	ATOM	2359	N	THR	B	37	30.824	0.737	25.279	1.00	9.13	N
	ATOM	2361	CA	THR	B	37	29.653	1.596	25.151	1.00	8.93	C
	ATOM	2363	CB	THR	B	37	28.725	1.447	26.372	1.00	9.51	C
	ATOM	2365	OG1	THR	B	37	28.238	0.095	26.458	1.00	10.29	OC
20	ATOM	2367	CG2	THR	B	37	27.474	2.316	26.234	1.00	10.22	CC
	ATOM	2371	C	THR	B	37	30.041	3.056	25.034	1.00	8.89	C
	ATOM	2372	O	THR	B	37	30.857	3.557	25.814	1.00	8.97	C
	ATOM	2373	N	PHE	B	38	29.450	3.724	24.042	1.00	7.97	ON
	ATOM	2375	CA	PHE	B	38	29.584	5.161	23.853	1.00	8.58	C
25	ATOM	2377	CB	PHE	B	38	29.827	5.456	22.386	1.00	8.85	C
	ATOM	2380	CG	PHE	B	38	31.134	4.951	21.847	1.00	7.28	C
	ATOM	2381	CD1	PHE	B	38	32.340	5.237	22.482	1.00	7.91	C
	ATOM	2383	CE1	PHE	B	38	33.544	4.811	21.942	1.00	8.72	C
	ATOM	2385	CZ	PHE	B	38	33.555	4.102	20.756	1.00	11.58	C
30	ATOM	2387	CE2	PHE	B	38	32.366	3.817	20.120	1.00	9.53	C
	ATOM	2389	CD2	PHE	B	38	31.163	4.243	20.661	1.00	7.95	C
	ATOM	2391	C	PHE	B	38	28.269	5.844	24.273	1.00	8.73	C
	ATOM	2392	O	PHE	B	38	27.216	5.431	23.811	1.00	9.98	C
	ATOM	2393	N	PRO	B	39	28.293	6.842	25.163	1.00	8.76	NC
35	ATOM	2394	CA	PRO	B	39	27.036	7.460	25.636	1.00	8.86	C
	ATOM	2396	CB	PRO	B	39	27.497	8.386	26.780	1.00	9.34	C
	ATOM	2399	CG	PRO	B	39	28.785	7.866	27.199	1.00	9.35	C
	ATOM	2402	CD	PRO	B	39	29.448	7.348	25.916	1.00	9.58	C
	ATOM	2405	C	PRO	B	39	26.209	8.230	24.627	1.00	9.13	C
40	ATOM	2406	O	PRO	B	39	24.991	8.328	24.796	1.00	8.44	C
	ATOM	2407	N	LYS	B	40	26.834	8.794	23.602	1.00	9.49	ON
	ATOM	2409	CA	LYS	B	40	26.061	9.548	22.618	1.00	9.43	C
	ATOM	2411	CB	LYS	B	40	25.784	10.967	23.094	1.00	10.36	C
	ATOM	2414	CG	LYS	B	40	24.760	11.685	22.232	1.00	12.43	C
45	ATOM	2417	CD	LYS	B	40	24.661	13.182	22.550	1.00	16.76	C
	ATOM	2420	CE	LYS	B	40	24.030	13.456	23.916	1.00	22.56	C
	ATOM	2423	NZ	LYS	B	40	24.148	14.904	24.336	1.00	28.45	N
	ATOM	2427	C	LYS	B	40	26.748	9.529	21.265	1.00	9.14	C
	ATOM	2428	O	LYS	B	40	27.597	10.355	20.962	1.00	9.57	C
50	ATOM	2429	N	VAL	B	41	26.393	8.544	20.458	1.00	9.12	C
	ATOM	2431	CA	VAL	B	41	26.969	8.438	19.131	1.00	9.08	C
	ATOM	2433	CB	VAL	B	41	26.967	6.970	18.603	1.00	9.12	C
	ATOM	2435	CG1	VAL	B	41	27.769	6.059	19.553	1.00	8.74	C
	ATOM	2439	CG2	VAL	B	41	25.556	6.453	18.410	1.00	8.61	C
55	ATOM	2443	C	VAL	B	41	26.243	9.323	18.136	1.00	9.87	C
	ATOM	2444	O	VAL	B	41	25.107	9.759	18.350	1.00	9.30	C
	ATOM	2445	N	LEU	B	42	26.907	9.582	17.022	1.00	10.23	N
	ATOM	2447	CA	LEU	B	42	26.261	10.288	15.932	1.00	11.90	C
	ATOM	2449	CB	LEU	B	42	27.303	10.820	14.948	1.00	11.82	C
60	ATOM	2452	CG	LEU	B	42	28.246	11.835	15.604	1.00	13.37	C
	ATOM	2454	CD1	LEU	B	42	29.484	12.076	14.765	1.00	17.49	C
	ATOM	2458	CD2	LEU	B	42	27.504	13.150	15.923	1.00	13.74	C
	ATOM	2462	C	LEU	B	42	25.303	9.312	15.257	1.00	13.53	C

	ATOM	2586	O	TYR	B	49	33.035	-3.120	13.301	1.00	8.75	O
	ATOM	2587	N	ARG	B	50	34.838	-1.764	13.149	1.00	8.80	N
	ATOM	2589	CA	ARG	B	50	35.811	-2.821	12.915	1.00	8.50	C
5	ATOM	2591	CB	ARG	B	50	36.725	-2.427	11.771	1.00	8.52	C
	ATOM	2594	CG	ARG	B	50	37.615	-3.545	11.308	1.00	9.29	C
	ATOM	2597	CD	ARG	B	50	38.349	-3.220	10.048	1.00	9.53	C
	ATOM	2600	NE	ARG	B	50	39.382	-2.205	10.191	1.00	8.80	N
	ATOM	2602	CZ	ARG	B	50	40.631	-2.476	10.566	1.00	11.23	C
10	ATOM	2603	NH1	ARG	B	50	40.986	-3.721	10.901	1.00	11.51	N
	ATOM	2606	NH2	ARG	B	50	41.533	-1.506	10.650	1.00	13.88	N
	ATOM	2609	C	ARG	B	50	36.627	-2.987	14.186	1.00	8.77	C
	ATOM	2610	O	ARG	B	50	37.063	-2.001	14.787	1.00	8.89	O
	ATOM	2611	N	VAL	B	51	36.797	-4.230	14.609	1.00	9.25	N
15	ATOM	2613	CA	VAL	B	51	37.467	-4.543	15.860	1.00	9.05	C
	ATOM	2615	CB	VAL	B	51	36.503	-5.312	16.778	1.00	9.51	C
	ATOM	2617	CG1	VAL	B	51	37.193	-5.808	18.029	1.00	10.94	C
	ATOM	2621	CG2	VAL	B	51	35.356	-4.425	17.179	1.00	9.48	C
	ATOM	2625	C	VAL	B	51	38.729	-5.378	15.614	1.00	9.31	C
20	ATOM	2626	O	VAL	B	51	38.692	-6.344	14.865	1.00	9.58	O
	ATOM	2627	N	VAL	B	52	39.827	-5.004	16.274	1.00	8.69	N
	ATOM	2629	CA	VAL	B	52	41.136	-5.638	16.112	1.00	8.61	C
	ATOM	2631	CB	VAL	B	52	42.132	-4.664	15.406	1.00	8.82	C
	ATOM	2633	CG1	VAL	B	52	43.432	-5.350	15.057	1.00	8.22	C
25	ATOM	2637	CG2	VAL	B	52	41.503	-4.031	14.166	1.00	8.20	C
	ATOM	2641	C	VAL	B	52	41.680	-6.010	17.490	1.00	8.96	C
	ATOM	2642	O	VAL	B	52	41.759	-5.166	18.367	1.00	9.37	O
	ATOM	2643	N	VAL	B	53	42.050	-7.276	17.677	1.00	8.28	N
	ATOM	2645	CA	VAL	B	53	42.521	-7.758	18.973	1.00	9.11	C
30	ATOM	2647	CB	VAL	B	53	41.645	-8.918	19.482	1.00	9.88	C
	ATOM	2649	CG1	VAL	B	53	42.248	-9.531	20.746	1.00	10.11	C
	ATOM	2653	CG2	VAL	B	53	40.207	-8.420	19.731	1.00	10.75	C
	ATOM	2657	C	VAL	B	53	43.965	-8.214	18.850	1.00	9.11	C
	ATOM	2658	O	VAL	B	53	44.254	-9.170	18.105	1.00	9.06	O
35	ATOM	2659	N	ASN	B	54	44.873	-7.543	19.563	1.00	8.85	N
	ATOM	2661	CA	ASN	B	54	46.310	-7.827	19.435	1.00	9.61	C
	ATOM	2663	CB	ASN	B	54	46.676	-9.140	20.109	1.00	10.03	C
	ATOM	2666	CG	ASN	B	54	47.031	-8.996	21.586	1.00	11.16	C
	ATOM	2667	OD1	ASN	B	54	47.247	-10.014	22.267	1.00	16.89	C
40	ATOM	2668	ND2	ASN	B	54	47.126	-7.781	22.076	1.00	9.72	C
	ATOM	2671	C	ASN	B	54	46.747	-7.870	17.956	1.00	10.26	C
	ATOM	2672	O	ASN	B	54	47.522	-8.745	17.548	1.00	11.02	O
	ATOM	2673	N	GLY	B	55	46.238	-6.928	17.168	1.00	10.54	N
	ATOM	2675	CA	GLY	B	55	46.575	-6.793	15.760	1.00	9.95	C
45	ATOM	2678	C	GLY	B	55	45.844	-7.707	14.792	1.00	9.84	C
	ATOM	2679	O	GLY	B	55	45.998	-7.522	13.579	1.00	9.89	O
	ATOM	2680	N	SER	B	56	45.036	-8.629	15.310	1.00	9.14	N
	ATOM	2682	CA	SER	B	56	44.226	-9.538	14.506	1.00	9.52	C
	ATOM	2684	CB	SER	B	56	44.022	-10.867	15.235	1.00	10.16	C
50	ATOM	2687	OG	SER	B	56	43.162	-11.730	14.503	1.00	10.98	O
	ATOM	2689	C	SER	B	56	42.858	-8.888	14.232	1.00	9.17	C
	ATOM	2690	O	SER	B	56	42.065	-8.653	15.148	1.00	8.64	O
	ATOM	2691	N	ASP	B	57	42.613	-8.558	12.976	1.00	9.04	N
	ATOM	2693	CA	ASP	B	57	41.358	-7.950	12.530	1.00	8.46	C
55	ATOM	2695	CB	ASP	B	57	41.559	-7.526	11.067	1.00	8.57	C
	ATOM	2698	CG	ASP	B	57	40.364	-6.842	10.457	1.00	9.39	C
	ATOM	2699	OD1	ASP	B	57	40.383	-6.708	9.193	1.00	9.52	O
	ATOM	2700	OD2	ASP	B	57	39.385	-6.414	11.106	1.00	9.56	O
	ATOM	2701	C	ASP	B	57	40.201	-8.950	12.628	1.00	8.72	C
60	ATOM	2702	O	ASP	B	57	40.218	-10.003	11.966	1.00	8.92	O
	ATOM	2703	N	LEU	B	58	39.217	-8.665	13.478	1.00	9.29	N
	ATOM	2705	CA	LEU	B	58	38.021	-9.508	13.542	1.00	9.33	C
	ATOM	2707	CB	LEU	B	58	37.508	-9.582	14.977	1.00	9.53	C
	ATOM	2710	CG	LEU	B	58	38.564	-9.973	16.005	1.00	9.47	C

	ATOM	2712	CD1	LEU	B	58	37.925	-10.156	17.379	1.00	11.17	C
	ATOM	2716	CD2	LEU	B	58	39.325	-11.242	15.604	1.00	11.86	C
	ATOM	2720	C	LEU	B	58	36.897	-9.037	12.608	1.00	9.41	C
5	ATOM	2721	O	LEU	B	58	35.797	-9.607	12.608	1.00	10.79	C
	ATOM	2722	N	GLY	B	59	37.166	-8.006	11.826	1.00	9.53	CON
	ATOM	2724	CA	GLY	B	59	36.245	-7.517	10.815	1.00	9.73	CC
	ATOM	2727	C	GLY	B	59	35.209	-6.570	11.375	1.00	9.94	CC
	ATOM	2728	O	GLY	B	59	35.355	-6.046	12.482	1.00	9.32	CO
10	ATOM	2729	N	VAL	B	60	34.133	-6.407	10.614	1.00	10.86	ON
	ATOM	2731	CA	VAL	B	60	33.165	-5.356	10.840	1.00	11.57	CC
	ATOM	2733	CB	VAL	B	60	33.048	-4.476	9.571	1.00	11.80	CC
	ATOM	2735	CG1	VAL	B	60	32.330	-3.170	9.845	1.00	13.91	CCC
	ATOM	2739	CG2	VAL	B	60	32.361	-5.227	8.443	1.00	13.46	CC
15	ATOM	2743	C	VAL	B	60	31.806	-5.894	11.230	1.00	11.41	CC
	ATOM	2744	O	VAL	B	60	31.409	-7.016	10.849	1.00	11.80	CON
	ATOM	2745	N	GLU	B	61	31.090	-5.084	11.998	1.00	10.41	NC
	ATOM	2747	CA	GLU	B	61	29.728	-5.416	12.391	1.00	11.68	CC
	ATOM	2749	CB	GLU	B	61	29.701	-6.352	13.600	1.00	11.99	CC
20	ATOM	2752	CG	GLU	B	61	28.316	-6.803	14.051	1.00	16.37	CC
	ATOM	2755	CD	GLU	B	61	27.469	-7.353	12.931	1.00	18.62	CC
	ATOM	2756	OE1	GLU	B	61	26.499	-6.673	12.533	1.00	19.03	OO
	ATOM	2757	OE2	GLU	B	61	27.791	-8.452	12.418	1.00	20.11	OC
	ATOM	2758	C	GLU	B	61	28.994	-4.105	12.643	1.00	12.02	CC
25	ATOM	2759	O	GLU	B	61	29.616	-3.087	12.944	1.00	10.24	CON
	ATOM	2760	N	SER	B	62	27.673	-4.129	12.540	1.00	13.34	CC
	ATOM	2762	CA	SER	B	62	26.908	-2.919	12.741	1.00	13.52	CC
	ATOM	2764	CB	SER	B	62	26.526	-2.289	11.407	1.00	14.00	CC
	ATOM	2767	OG	SER	B	62	25.717	-3.176	10.647	1.00	13.96	OC
30	ATOM	2769	C	SER	B	62	25.631	-3.142	13.513	1.00	13.46	CC
	ATOM	2770	O	SER	B	62	24.940	-2.159	13.807	1.00	14.34	CC
	ATOM	2771	N	ASN	B	63	25.333	-4.389	13.869	1.00	12.74	CON
	ATOM	2773	CA	ASN	B	63	24.037	-4.692	14.474	1.00	14.23	CC
	ATOM	2775	CB	ASN	B	63	23.557	-6.123	14.156	1.00	15.00	CC
35	ATOM	2778	CG	ASN	B	63	22.193	-6.441	14.798	1.00	16.84	CC
	ATOM	2779	OD1	ASN	B	63	21.441	-5.531	15.129	1.00	21.06	CC
	ATOM	2780	ND2	ASN	B	63	21.895	-7.727	15.010	1.00	21.14	CC
	ATOM	2783	C	ASN	B	63	24.089	-4.438	15.971	1.00	13.99	CC
	ATOM	2784	O	ASN	B	63	24.093	-5.385	16.770	1.00	13.96	CC
40	ATOM	2785	N	PHE	B	64	24.126	-3.143	16.308	1.00	13.92	NC
	ATOM	2787	CA	PHE	B	64	24.126	-2.622	17.673	1.00	12.95	CC
	ATOM	2789	CB	PHE	B	64	25.518	-2.121	18.080	1.00	12.24	CC
	ATOM	2792	CG	PHE	B	64	26.621	-3.056	17.698	1.00	10.08	CC
	ATOM	2793	CD1	PHE	B	64	26.707	-4.313	18.267	1.00	9.82	CC
45	ATOM	2795	CE1	PHE	B	64	27.717	-5.176	17.900	1.00	10.20	CC
	ATOM	2797	CZ	PHE	B	64	28.622	-4.784	16.947	1.00	10.37	CC
	ATOM	2799	CE2	PHE	B	64	28.533	-3.542	16.383	1.00	10.15	CC
	ATOM	2801	CD2	PHE	B	64	27.547	-2.689	16.752	1.00	9.91	CC
	ATOM	2803	C	PHE	B	64	23.135	-1.462	17.777	1.00	12.85	CC
50	ATOM	2804	O	PHE	B	64	23.282	-0.433	17.128	1.00	12.07	ON
	ATOM	2805	N	ALA	B	65	22.124	-1.637	18.613	1.00	13.52	C
	ATOM	2807	CA	ALA	B	65	21.038	-0.677	18.719	1.00	13.11	C
	ATOM	2809	CB	ALA	B	65	19.990	-1.165	19.701	1.00	13.88	C
	ATOM	2813	C	ALA	B	65	21.554	0.641	19.215	1.00	13.59	C
55	ATOM	2814	O	ALA	B	65	22.471	0.668	20.026	1.00	13.52	ON
	ATOM	2815	N	VAL	B	66	20.985	1.727	18.716	1.00	13.44	NC
	ATOM	2817	CA	VAL	B	66	21.223	3.015	19.337	1.00	13.23	CC
	ATOM	2819	CB	VAL	B	66	21.412	4.126	18.322	1.00	13.46	CC
	ATOM	2821	CG1	VAL	B	66	21.554	5.453	19.053	1.00	14.25	CC
60	ATOM	2825	CG2	VAL	B	66	22.634	3.839	17.457	1.00	12.99	CC
	ATOM	2829	C	VAL	B	66	20.007	3.276	20.232	1.00	13.98	CC
	ATOM	2830	O	VAL	B	66	18.860	3.239	19.765	1.00	14.47	ON
	ATOM	2831	N	THR	B	67	20.241	3.517	21.512	1.00	13.83	ON
	ATOM	2833	CA	THR	B	67	19.134	3.762	22.438	1.00	15.29	C

	ATOM	2958	CE1	PHE	B	76	34.715	-4.318	20.939	1.00	9.26	C
	ATOM	2960	CZ	PHE	B	76	35.131	-3.763	22.112	1.00	8.99	C
	ATOM	2962	CE2	PHE	B	76	35.143	-4.500	23.259	1.00	10.57	C
	ATOM	2964	CD2	PHE	B	76	34.745	-5.806	23.246	1.00	10.62	C
5	ATOM	2966	C	PHE	B	76	32.260	-9.502	20.999	1.00	9.01	C
	ATOM	2967	O	PHE	B	76	32.893	-10.149	20.169	1.00	9.14	O
	ATOM	2968	N	LEU	B	77	31.268	-10.038	21.708	1.00	9.74	N
	ATOM	2970	CA	LEU	B	77	30.894	-11.444	21.525	1.00	9.62	C
	ATOM	2972	CB	LEU	B	77	29.844	-11.840	22.565	1.00	9.60	C
10	ATOM	2975	CG	LEU	B	77	30.361	-12.157	23.981	1.00	11.85	C
	ATOM	2977	CD1	LEU	B	77	31.102	-11.029	24.640	1.00	13.22	C
	ATOM	2981	CD2	LEU	B	77	29.174	-12.593	24.854	1.00	12.68	C
	ATOM	2985	C	LEU	B	77	30.400	-11.713	20.077	1.00	9.32	C
	ATOM	2986	O	LEU	B	77	30.481	-12.843	19.574	1.00	10.46	O
15	ATOM	2987	N	GLN	B	78	29.907	-10.671	19.415	1.00	9.10	N
	ATOM	2989	CA	GLN	B	78	29.450	-10.748	18.032	1.00	10.10	C
	ATOM	2991	CB	GLN	B	78	28.517	-9.569	17.697	1.00	10.43	C
	ATOM	2994	CG	GLN	B	78	27.143	-9.734	18.379	1.00	9.66	C
	ATOM	2997	CD	GLN	B	78	26.224	-8.538	18.295	1.00	11.71	C
20	ATOM	2998	OE1	GLN	B	78	25.803	-8.019	19.329	1.00	11.16	O
	ATOM	2999	NE2	GLN	B	78	25.871	-8.113	17.071	1.00	13.17	C
	ATOM	3002	C	GLN	B	78	30.612	-10.816	17.042	1.00	11.05	C
	ATOM	3003	O	GLN	B	78	30.379	-11.128	15.859	1.00	13.23	O
	ATOM	3004	N	TYR	B	79	31.826	-10.506	17.500	1.00	10.96	N
25	ATOM	3006	CA	TYR	B	79	33.026	-10.570	16.654	1.00	12.02	C
	ATOM	3008	CB	TYR	B	79	33.915	-9.339	16.842	1.00	11.81	C
	ATOM	3011	CG	TYR	B	79	33.345	-8.005	16.481	1.00	10.03	C
	ATOM	3012	CD1	TYR	B	79	33.681	-7.372	15.280	1.00	8.25	C
	ATOM	3014	CE1	TYR	B	79	33.169	-6.116	14.965	1.00	8.39	C
30	ATOM	3016	CZ	TYR	B	79	32.322	-5.483	15.842	1.00	8.21	C
	ATOM	3017	OH	TYR	B	79	31.827	-4.250	15.505	1.00	8.87	O
	ATOM	3019	CE2	TYR	B	79	31.972	-6.104	17.033	1.00	10.02	C
	ATOM	3021	CD2	TYR	B	79	32.487	-7.347	17.345	1.00	9.93	C
	ATOM	3023	C	TYR	B	79	33.940	-11.728	16.985	1.00	13.56	C
35	ATOM	3024	O	TYR	B	79	34.689	-12.190	16.120	1.00	13.49	O
	ATOM	3025	N	ASN	B	80	33.919	-12.162	18.242	1.00	15.17	N
	ATOM	3027	CA	ASN	B	80	34.925	-13.076	18.751	1.00	14.48	C
	ATOM	3029	CB	ASN	B	80	35.582	-12.444	19.978	1.00	14.14	C
	ATOM	3032	CG	ASN	B	80	36.890	-13.105	20.361	1.00	14.62	C
40	ATOM	3033	OD1	ASN	B	80	37.651	-13.558	19.509	1.00	16.67	O
	ATOM	3034	ND2	ASN	B	80	37.153	-13.163	21.664	1.00	13.88	N
	ATOM	3037	C	ASN	B	80	34.352	-14.437	19.084	1.00	14.66	C
	ATOM	3038	O	ASN	B	80	34.712	-15.056	20.083	1.00	13.88	O
	ATOM	3039	N	LYS	B	81	33.412	-14.879	18.257	1.00	15.51	N
45	ATOM	3041	CA	LYS	B	81	32.859	-16.233	18.368	1.00	16.46	C
	ATOM	3043	CB	LYS	B	81	33.943	-17.263	17.992	1.00	16.91	C
	ATOM	3046	CG	LYS	B	81	34.644	-16.962	16.663	1.00	20.41	C
	ATOM	3049	CD	LYS	B	81	35.616	-18.076	16.175	1.00	26.52	C
	ATOM	3052	CE	LYS	B	81	36.392	-18.798	17.293	1.00	31.18	C
50	ATOM	3055	NZ	LYS	B	81	37.525	-19.720	16.822	1.00	38.55	N
	ATOM	3059	C	LYS	B	81	32.216	-16.559	19.735	1.00	15.91	C
	ATOM	3060	O	LYS	B	81	32.322	-17.678	20.227	1.00	16.16	O
	ATOM	3061	N	GLY	B	82	31.522	-15.575	20.307	1.00	15.03	N
	ATOM	3063	CA	GLY	B	82	30.735	-15.736	21.512	1.00	15.26	C
55	ATOM	3066	C	GLY	B	82	31.492	-15.553	22.800	1.00	15.69	C
	ATOM	3067	O	GLY	B	82	30.960	-15.833	23.857	1.00	15.34	O
	ATOM	3068	N	TYR	B	83	32.721	-15.052	22.708	1.00	16.90	N
	ATOM	3070	CA	TYR	B	83	33.550	-14.806	23.873	1.00	17.19	C
	ATOM	3072	CB	TYR	B	83	34.815	-15.667	23.808	1.00	18.20	C
60	ATOM	3075	CG	TYR	B	83	34.583	-17.155	23.934	1.00	23.38	C
	ATOM	3076	CD1	TYR	B	83	34.448	-17.745	25.181	1.00	28.89	C
	ATOM	3078	CE1	TYR	B	83	34.239	-19.105	25.310	1.00	31.05	C
	ATOM	3080	C2	TYR	B	83	34.170	-19.896	24.184	1.00	32.68	C

	ATOM	3081	OH	TYR	B	83	33.962	-21.259	24.329	1.00	36.36	O
	ATOM	3083	CE2	TYR	B	83	34.305	-19.340	22.930	1.00	30.78	C
	ATOM	3085	CD2	TYR	B	83	34.506	-17.972	22.810	1.00	28.62	C
5	ATOM	3087	C	TYR	B	83	34.006	-13.355	23.925	1.00	16.15	C
	ATOM	3088	O	TYR	B	83	34.169	-12.715	22.894	1.00	15.94	O
	ATOM	3089	N	GLY	B	84	34.190	-12.828	25.134	1.00	15.91	N
	ATOM	3091	CA	GLY	B	84	34.842	-11.539	25.278	1.00	14.80	C
	ATOM	3094	C	GLY	B	84	36.359	-11.675	25.047	1.00	13.96	C
10	ATOM	3095	O	GLY	B	84	36.882	-12.737	24.675	1.00	14.24	O
	ATOM	3096	N	VAL	B	85	37.071	-10.586	25.311	1.00	11.64	N
	ATOM	3098	CA	VAL	B	85	38.497	-10.478	25.027	1.00	12.19	C
	ATOM	3100	CB	VAL	B	85	38.761	-9.214	24.159	1.00	11.76	C
	ATOM	3102	CG1	VAL	B	85	40.254	-9.019	23.892	1.00	12.27	C
15	ATOM	3106	CG2	VAL	B	85	37.970	-9.236	22.852	1.00	12.61	C
	ATOM	3110	C	VAL	B	85	39.258	-10.315	26.329	1.00	12.06	C
	ATOM	3111	O	VAL	B	85	38.954	-9.435	27.133	1.00	11.39	O
	ATOM	3112	N	ALA	B	86	40.263	-11.154	26.554	1.00	11.94	N
	ATOM	3114	CA	ALA	B	86	41.077	-11.026	27.752	1.00	12.43	C
20	ATOM	3116	CB	ALA	B	86	42.221	-12.031	27.709	1.00	12.46	C
	ATOM	3120	C	ALA	B	86	41.630	-9.606	27.895	1.00	12.58	C
	ATOM	3121	O	ALA	B	86	42.145	-9.034	26.921	1.00	11.12	O
	ATOM	3122	N	ASP	B	87	41.542	-9.046	29.101	1.00	12.96	N
	ATOM	3124	CA	ASP	B	87	41.977	-7.664	29.319	1.00	13.26	C
25	ATOM	3126	CB	ASP	B	87	41.413	-7.038	30.599	1.00	12.94	C
	ATOM	3129	CG	ASP	B	87	41.973	-7.621	31.863	1.00	15.55	C
	ATOM	3130	OD1	ASP	B	87	42.925	-8.435	31.811	1.00	16.54	O
	ATOM	3131	OD2	ASP	B	87	41.478	-7.304	32.971	1.00	17.64	O
	ATOM	3132	C	ASP	B	87	43.473	-7.446	29.177	1.00	13.14	C
30	ATOM	3133	O	ASP	B	87	43.923	-6.303	29.211	1.00	13.88	O
	ATOM	3134	N	THR	B	88	44.222	-8.529	28.986	1.00	13.35	N
	ATOM	3136	CA	THR	B	88	45.648	-8.426	28.770	1.00	13.89	C
	ATOM	3138	CB	THR	B	88	46.376	-9.702	29.223	1.00	13.97	C
	ATOM	3140	OG1	THR	B	88	45.720	-10.853	28.692	1.00	14.23	O
35	ATOM	3142	CG2	THR	B	88	46.312	-9.865	30.728	1.00	15.70	C
	ATOM	3146	C	THR	B	88	45.972	-8.179	27.308	1.00	14.14	C
	ATOM	3147	O	THR	B	88	47.138	-7.960	26.978	1.00	16.23	O
	ATOM	3148	N	LYS	B	89	44.961	-8.217	26.444	1.00	13.28	N
	ATOM	3150	CA	LYS	B	89	45.163	-7.962	25.021	1.00	13.04	C
40	ATOM	3152	CB	LYS	B	89	44.317	-8.922	24.185	1.00	13.74	C
	ATOM	3155	CG	LYS	B	89	44.531	-10.371	24.562	1.00	15.15	C
	ATOM	3158	CD	LYS	B	89	43.821	-11.309	23.612	1.00	18.37	C
	ATOM	3161	CE	LYS	B	89	43.980	-12.771	24.006	1.00	20.95	C
	ATOM	3164	NZ	LYS	B	89	43.412	-13.640	22.933	1.00	24.82	N
45	ATOM	3168	C	LYS	B	89	44.798	-6.531	24.667	1.00	12.47	C
	ATOM	3169	O	LYS	B	89	44.022	-5.884	25.369	1.00	12.82	O
	ATOM	3170	N	THR	B	90	45.377	-6.023	23.589	1.00	11.85	N
	ATOM	3172	CA	THR	B	90	45.017	-4.714	23.072	1.00	11.57	C
	ATOM	3174	CB	THR	B	90	46.177	-4.184	22.273	1.00	11.83	C
	ATOM	3176	OG1	THR	B	90	47.280	-3.912	23.164	1.00	14.04	O
50	ATOM	3178	CG2	THR	B	90	45.838	-2.872	21.626	1.00	13.14	C
	ATOM	3182	C	THR	B	90	43.780	-4.827	22.180	1.00	11.03	C
	ATOM	3183	O	THR	B	90	43.684	-5.748	21.355	1.00	10.60	O
	ATOM	3184	N	ILE	B	91	42.839	-3.893	22.344	1.00	10.28	N
	ATOM	3186	CA	ILE	B	91	41.640	-3.871	21.530	1.00	10.28	C
55	ATOM	3188	CB	ILE	B	91	40.361	-4.011	22.361	1.00	10.29	C
	ATOM	3190	CG1	ILE	B	91	40.407	-5.261	23.238	1.00	11.66	C
	ATOM	3193	CD1	ILE	B	91	39.298	-5.354	24.285	1.00	13.60	C
	ATOM	3197	CG2	ILE	B	91	39.141	-4.060	21.408	1.00	10.59	C
60	ATOM	3201	C	ILE	B	91	41.597	-2.536	20.815	1.00	10.62	C
	ATOM	3202	O	ILE	B	91	41.700	-1.489	21.442	1.00	10.66	O
	ATOM	3203	N	GLN	B	92	41.490	-2.563	19.497	1.00	10.49	N
	ATOM	3205	CA	GLN	B	92	41.313	-1.326	18.764	1.00	10.71	C
	ATOM	3207	CB	GLN	B	92	42.410	-1.146	17.719	1.00	11.65	C

	ATOM	3210	CG	GLN	B	92	43.778	-0.942	18.282	1.00	15.30	C
	ATOM	3213	CD	GLN	B	92	44.809	-1.227	17.215	1.00	21.75	C
	ATOM	3214	OE1	GLN	B	92	45.030	-2.382	16.855	1.00	23.06	CON
	ATOM	3215	NE2	GLN	B	92	45.388	-0.176	16.654	1.00	29.53	C
5	ATOM	3218	C	GLN	B	92	39.964	-1.384	18.063	1.00	9.76	C
	ATOM	3219	O	GLN	B	92	39.599	-2.416	17.499	1.00	9.52	CON
	ATOM	3220	N	VAL	B	93	39.225	-0.286	18.109	1.00	8.49	C
	ATOM	3222	CA	VAL	B	93	37.922	-0.204	17.447	1.00	8.46	CC
	ATOM	3224	CB	VAL	B	93	36.754	-0.031	18.452	1.00	8.40	CC
10	ATOM	3226	CG1	VAL	B	93	35.408	-0.068	17.715	1.00	8.33	CCC
	ATOM	3230	CG2	VAL	B	93	36.818	-1.110	19.565	1.00	9.21	CCC
	ATOM	3234	C	VAL	B	93	37.954	0.979	16.498	1.00	8.80	CC
	ATOM	3235	O	VAL	B	93	38.313	2.100	16.913	1.00	9.38	CO
	ATOM	3236	N	PHE	B	94	37.585	0.735	15.235	1.00	7.89	ON
15	ATOM	3238	CA	PHE	B	94	37.514	1.756	14.207	1.00	8.32	CC
	ATOM	3240	CB	PHE	B	94	38.303	1.336	12.954	1.00	8.53	CC
	ATOM	3243	CG	PHE	B	94	39.774	1.165	13.207	1.00	8.61	CC
	ATOM	3244	CD1	PHE	B	94	40.658	2.188	12.898	1.00	10.93	CC
	ATOM	3246	CE1	PHE	B	94	42.001	2.046	13.153	1.00	11.25	CC
20	ATOM	3248	CZ	PHE	B	94	42.482	0.899	13.732	1.00	10.46	CC
	ATOM	3250	CE2	PHE	B	94	41.644	-0.131	14.027	1.00	10.75	CC
	ATOM	3252	CD2	PHE	B	94	40.273	0.003	13.776	1.00	10.71	CC
	ATOM	3254	C	PHE	B	94	36.070	1.974	13.819	1.00	8.61	CC
	ATOM	3255	O	PHE	B	94	35.314	1.013	13.648	1.00	9.56	CON
25	ATOM	3256	N	VAL	B	95	35.687	3.231	13.647	1.00	8.45	ON
	ATOM	3258	CA	VAL	B	95	34.370	3.546	13.104	1.00	9.19	CC
	ATOM	3260	CB	VAL	B	95	33.767	4.834	13.699	1.00	9.11	CC
	ATOM	3262	CG1	VAL	B	95	34.614	6.052	13.439	1.00	11.33	CC
	ATOM	3266	CG2	VAL	B	95	32.342	5.013	13.203	1.00	10.43	CC
30	ATOM	3270	C	VAL	B	95	34.535	3.607	11.594	1.00	9.06	ON
	ATOM	3271	O	VAL	B	95	35.480	4.251	11.094	1.00	9.36	ON
	ATOM	3272	N	VAL	B	96	33.660	2.896	10.888	1.00	10.07	ON
	ATOM	3274	CA	VAL	B	96	33.731	2.760	9.435	1.00	10.75	CC
	ATOM	3276	CB	VAL	B	96	33.633	1.287	9.027	1.00	11.31	CC
35	ATOM	3278	CG1	VAL	B	96	33.699	1.117	7.507	1.00	12.23	CC
	ATOM	3282	CG2	VAL	B	96	34.726	0.475	9.716	1.00	11.29	CC
	ATOM	3286	C	VAL	B	96	32.598	3.544	8.811	1.00	11.08	CC
	ATOM	3287	O	VAL	B	96	31.425	3.388	9.170	1.00	11.36	ON
	ATOM	3288	N	ILE	B	97	32.948	4.403	7.866	1.00	11.39	ON
40	ATOM	3290	CA	ILE	B	97	31.959	5.265	7.236	1.00	12.42	CC
	ATOM	3292	CB	ILE	B	97	32.677	6.496	6.644	1.00	12.48	CC
	ATOM	3294	CG1	ILE	B	97	33.614	7.145	7.677	1.00	12.52	CC
	ATOM	3297	CD1	ILE	B	97	32.936	7.612	8.958	1.00	14.19	CC
	ATOM	3301	CG2	ILE	B	97	31.669	7.488	6.082	1.00	12.90	CC
45	ATOM	3305	C	ILE	B	97	31.234	4.501	6.130	1.00	13.56	ON
	ATOM	3306	O	ILE	B	97	31.898	3.883	5.308	1.00	13.06	ON
	ATOM	3307	N	PRO	B	98	29.898	4.524	6.113	1.00	15.09	CC
	ATOM	3308	CA	PRO	B	98	29.132	3.809	5.086	1.00	16.07	CC
	ATOM	3310	CB	PRO	B	98	27.696	3.899	5.600	1.00	15.86	CC
50	ATOM	3313	CG	PRO	B	98	27.661	5.122	6.370	1.00	16.80	CC
	ATOM	3316	CD	PRO	B	98	29.007	5.184	7.076	1.00	14.97	CC
	ATOM	3319	C	PRO	B	98	29.267	4.451	3.718	1.00	17.70	CC
	ATOM	3320	O	PRO	B	98	29.605	5.631	3.592	1.00	17.04	ON
	ATOM	3321	N	ASP	B	99	29.014	3.649	2.696	1.00	20.10	ON
55	ATOM	3323	CA	ASP	B	99	29.082	4.091	1.303	1.00	20.04	CC
	ATOM	3325	CB	ASP	B	99	28.029	5.172	1.061	1.00	21.12	CC
	ATOM	3328	CG	ASP	B	99	26.612	4.657	1.337	1.00	23.44	CC
	ATOM	3329	OD1	ASP	B	99	26.291	3.537	0.874	1.00	27.92	CO
	ATOM	3330	OD2	ASP	B	99	25.761	5.269	2.020	1.00	28.30	OC
60	ATOM	3331	C	ASP	B	99	30.494	4.496	0.860	1.00	19.38	OC
	ATOM	3332	O	ASP	B	99	30.646	5.358	-0.015	1.00	19.12	ON
	ATOM	3333	N	THR	B	100	31.521	3.879	1.460	1.00	17.96	NC
	ATOM	3335	CA	THR	B	100	32.917	4.061	1.030	1.00	18.27	NC

ATOM	3337	CB	THR	B	100	33.757	4.812	2.091	1.00	18.29	C	
ATOM	3339	OG1	THR	B	100	33.964	3.969	3.249	1.00	16.21	C	
ATOM	3341	CG2	THR	B	100	33.041	6.065	2.595	1.00	18.22	C	
ATOM	3345	C	THR	B	100	33.606	2.725	0.744	1.00	18.55	C	
5	ATOM	3346	O	THR	B	100	34.839	2.644	0.724	1.00	18.17	C
ATOM	3347	N	GLY	B	101	32.813	1.676	0.537	1.00	19.93	C	
ATOM	3349	CA	GLY	B	101	33.346	0.341	0.306	1.00	19.59	C	
ATOM	3352	C	GLY	B	101	34.217	-0.119	1.467	1.00	19.66	C	
0	ATOM	3353	O	GLY	B	101	35.181	-0.861	1.285	1.00	19.57	C
ATOM	3354	N	ASN	B	102	33.856	0.343	2.661	1.00	19.84	C	
ATOM	3356	CA	ASN	B	102	34.580	0.071	3.906	1.00	19.99	C	
ATOM	3358	CB	ASN	B	102	34.517	-1.406	4.266	1.00	20.27	C	
ATOM	3361	CG	ASN	B	102	33.148	-1.823	4.683	1.00	22.54	C	
5	ATOM	3362	OD1	ASN	B	102	32.856	-1.970	5.871	1.00	26.86	C
ATOM	3363	ND2	ASN	B	102	32.293	-2.035	3.709	1.00	25.85	C	
ATOM	3366	C	ASN	B	102	36.016	0.557	3.943	1.00	19.98	C	
ATOM	3367	O	ASN	B	102	36.805	0.124	4.782	1.00	19.09	C	
ATOM	3368	N	SER	B	103	36.343	1.504	3.076	1.00	20.60	C	
ATOM	3370	CA	SER	B	103	37.707	1.990	2.986	1.00	19.63	C	
0	ATOM	3372	CB	SER	B	103	38.016	2.353	1.541	1.00	20.16	C
ATOM	3375	OG	SER	B	103	37.253	3.481	1.156	1.00	22.36	C	
ATOM	3377	C	SER	B	103	37.979	3.214	3.870	1.00	18.01	C	
ATOM	3378	O	SER	B	103	39.137	3.525	4.144	1.00	18.98	C	
ATOM	3379	N	GLU	B	104	36.936	3.918	4.294	1.00	15.67	C	
15	ATOM	3381	CA	GLU	B	104	37.135	5.116	5.120	1.00	13.43	C
ATOM	3383	CB	GLU	B	104	36.265	6.297	4.656	1.00	12.81	C	
ATOM	3386	CG	GLU	B	104	36.679	7.618	5.313	1.00	13.27	C	
ATOM	3389	CD	GLU	B	104	35.732	8.776	5.029	1.00	14.64	C	
ATOM	3390	OE1	GLU	B	104	34.919	8.679	4.069	1.00	15.08	C	
30	ATOM	3391	OE2	GLU	B	104	35.814	9.803	5.742	1.00	14.12	C
ATOM	3392	C	GLU	B	104	36.797	4.780	6.558	1.00	12.23	C	
ATOM	3393	O	GLU	B	104	35.659	4.435	6.855	1.00	11.79	C	
ATOM	3394	N	GLU	B	105	37.793	4.856	7.439	1.00	11.65	C	
ATOM	3396	CA	GLU	B	105	37.573	4.534	8.845	1.00	10.60	C	
35	ATOM	3398	CB	GLU	B	105	37.830	3.047	9.102	1.00	10.23	C
ATOM	3401	CG	GLU	B	105	39.288	2.653	8.998	1.00	11.51	C	
ATOM	3404	CD	GLU	B	105	39.569	1.177	9.250	1.00	13.25	C	
ATOM	3405	OE1	GLU	B	105	40.772	0.827	9.367	1.00	14.63	C	
40	ATOM	3406	OE2	GLU	B	105	38.617	0.366	9.341	1.00	11.87	C
ATOM	3407	C	GLU	B	105	38.476	5.381	9.732	1.00	9.65	C	
ATOM	3408	O	GLU	B	105	39.492	5.931	9.272	1.00	10.58	C	
ATOM	3409	N	TYR	B	106	38.112	5.465	11.014	1.00	9.46	C	
ATOM	3411	CA	TYR	B	106	38.842	6.260	12.004	1.00	9.17	C	
45	ATOM	3413	CB	TYR	B	106	38.119	7.593	12.258	1.00	9.00	C
ATOM	3416	CG	TYR	B	106	37.989	8.388	10.990	1.00	9.70	C	
ATOM	3417	CD1	TYR	B	106	39.000	9.227	10.565	1.00	10.02	C	
ATOM	3419	CE1	TYR	B	106	38.900	9.908	9.378	1.00	11.83	C	
ATOM	3421	CZ	TYR	B	106	37.797	9.755	8.584	1.00	10.67	C	
50	ATOM	3422	OH	TYR	B	106	37.719	10.438	7.368	1.00	13.02	C
ATOM	3424	CE2	TYR	B	106	36.784	8.909	8.958	1.00	10.26	C	
ATOM	3426	CD2	TYR	B	106	36.887	8.224	10.166	1.00	9.29	C	
ATOM	3428	C	TYR	B	106	38.949	5.516	13.318	1.00	9.17	C	
ATOM	3429	O	TYR	B	106	37.963	4.945	13.774	1.00	8.39	C	
ATOM	3430	N	ILE	B	107	40.116	5.546	13.955	1.00	8.89	C	
55	ATOM	3432	CA	ILE	B	107	40.211	4.966	15.288	1.00	8.90	C
ATOM	3434	CB	ILE	B	107	41.652	5.037	15.848	1.00	9.24	C	
ATOM	3436	CG1	ILE	B	107	41.770	4.245	17.155	1.00	10.76	C	
ATOM	3439	CD1	ILE	B	107	41.571	2.765	16.979	1.00	12.64	C	
ATOM	3443	CG2	ILE	B	107	42.088	6.484	16.051	1.00	9.73	C	
60	ATOM	3447	C	ILE	B	107	39.211	5.679	16.194	1.00	9.06	C
ATOM	3448	O	ILE	B	107	39.102	6.912	16.171	1.00	9.11	O	
ATOM	3449	N	ILE	B	108	38.448	4.908	16.958	1.00	8.10	N	
ATOM	3451	CA	ILE	B	108	37.466	5.505	17.858	1.00	8.46	C	

	ATOM	3453	CB	ILE	B	108	36.038	5.362	17.263	1.00	7.92	C
	ATOM	3455	CG1	ILE	B	108	35.058	6.318	17.933	1.00	8.44	C
	ATOM	3458	CD1	ILE	B	108	35.451	7.735	17.791	1.00	10.51	C
5	ATOM	3462	CG2	ILE	B	108	35.548	3.943	17.367	1.00	7.73	C
	ATOM	3466	C	ILE	B	108	37.577	5.032	19.315	1.00	9.21	O
	ATOM	3467	O	ILE	B	108	37.028	5.675	20.206	1.00	9.94	N
	ATOM	3468	N	ALA	B	109	38.295	3.939	19.566	1.00	8.98	C
	ATOM	3470	CA	ALA	B	109	38.565	3.467	20.929	1.00	8.87	C
10	ATOM	3472	CB	ALA	B	109	37.358	2.771	21.525	1.00	9.01	C
	ATOM	3476	C	ALA	B	109	39.744	2.507	20.935	1.00	8.73	C
	ATOM	3477	O	ALA	B	109	39.957	1.757	19.994	1.00	9.76	C
	ATOM	3478	N	GLU	B	110	40.533	2.576	21.998	1.00	9.54	C
	ATOM	3480	CA	GLU	B	110	41.607	1.634	22.215	1.00	9.46	N
15	ATOM	3482	CB	GLU	B	110	42.946	2.257	21.824	1.00	10.74	C
	ATOM	3485	CG	GLU	B	110	44.120	1.290	21.888	1.00	14.55	C
	ATOM	3488	CD	GLU	B	110	45.414	1.938	21.419	1.00	21.68	C
	ATOM	3489	OE1	GLU	B	110	45.950	2.812	22.134	1.00	25.49	O
	ATOM	3490	OE2	GLU	B	110	45.877	1.587	20.321	1.00	27.22	O
20	ATOM	3491	C	GLU	B	110	41.678	1.209	23.680	1.00	9.73	C
	ATOM	3492	O	GLU	B	110	41.720	2.056	24.582	1.00	10.15	O
	ATOM	3493	N	TRP	B	111	41.703	-0.098	23.894	1.00	9.39	N
	ATOM	3495	CA	TRP	B	111	41.990	-0.671	25.187	1.00	10.20	C
	ATOM	3497	CB	TRP	B	111	41.076	-1.840	25.491	1.00	9.67	C
25	ATOM	3500	CG	TRP	B	111	41.470	-2.528	26.787	1.00	8.52	C
	ATOM	3501	CD1	TRP	B	111	42.308	-3.581	26.928	1.00	9.95	C
	ATOM	3503	NE1	TRP	B	111	42.456	-3.906	28.256	1.00	13.10	N
	ATOM	3505	CE2	TRP	B	111	41.728	-3.017	29.010	1.00	10.98	C
	ATOM	3506	CD2	TRP	B	111	41.102	-2.128	28.116	1.00	9.50	C
30	ATOM	3507	CE3	TRP	B	111	40.282	-1.123	28.636	1.00	10.62	C
	ATOM	3509	CZ3	TRP	B	111	40.114	-1.040	30.002	1.00	12.26	C
	ATOM	3511	CH2	TRP	B	111	40.756	-1.929	30.857	1.00	9.96	C
	ATOM	3513	CZ2	TRP	B	111	41.561	-2.926	30.385	1.00	12.41	C
	ATOM	3515	C	TRP	B	111	43.423	-1.193	25.169	1.00	13.35	C
35	ATOM	3516	O	TRP	B	111	43.775	-2.031	24.344	1.00	11.63	O
	ATOM	3517	N	LYS	B	112	44.263	-0.666	26.056	1.00	16.48	N
	ATOM	3519	CA	LYS	B	112	45.593	-1.223	26.244	1.00	20.35	C
	ATOM	3521	CB	LYS	B	112	46.627	-0.459	25.436	1.00	22.00	C
	ATOM	3524	CG	LYS	B	112	47.926	-1.228	25.265	1.00	26.13	C
40	ATOM	3527	CD	LYS	B	112	49.024	-0.337	24.701	1.00	31.80	C
	ATOM	3530	CE	LYS	B	112	48.724	0.098	23.278	1.00	34.89	C
	ATOM	3533	NZ	LYS	B	112	49.811	0.945	22.716	1.00	40.47	N
	ATOM	3537	C	LYS	B	112	45.946	-1.177	27.725	1.00	22.27	C
	ATOM	3538	O	LYS	B	112	46.204	-0.081	28.253	1.00	23.96	O
45	ATOM	3539	BR	BR1	C	1	32.421	56.008	18.617	1.00	7.69	B
	ATOM	3540	BR	BR1	C	2	29.535	49.785	7.652	1.00	7.89	B
	ATOM	3541	BR	BR1	C	3	14.888	42.517	9.414	1.00	6.57	B
	ATOM	3542	BR	BR1	C	4	25.062	15.958	16.407	1.00	10.90	B
	ATOM	3543	BR	BR1	C	5	33.144	18.262	4.026	1.00	20.03	B
50	ATOM	3544	BR	BR1	C	6	40.800	30.559	10.185	1.00	12.36	B
	ATOM	3545	BR	BR1	C	7	30.248	54.190	19.852	1.00	14.74	B
	ATOM	3546	BR	BR1	C	8	38.772	41.003	24.687	1.00	22.37	B
	ATOM	3547	BR	BR1	C	9	26.990	5.115	28.326	1.00	15.47	B
	ATOM	3548	BR	BR1	C	10	40.148	5.267	23.548	1.00	2.00	B
55	ATOM	3549	BR	BR1	C	11	40.494	-13.035	23.333	1.00	14.97	B
	ATOM	3550	BR	BR1	C	12	26.318	-12.293	15.448	1.00	14.38	B
	ATOM	3551	BR	BR1	C	13	31.199	-18.188	15.135	1.00	9.41	B
	ATOM	3552	BR	BR1	C	14	32.035	-14.040	15.742	1.00	12.63	B
	ATOM	3553	BR	BR1	C	15	29.171	31.139	8.101	1.00	2.00	B
60	ATOM	3554	BR	BR1	C	16	28.318	-4.326	9.061	1.00	2.00	B
	ATOM	3555	O	HOH	D	1	45.016	-8.481	11.093	1.00	14.02	O
	ATOM	3558	O	HOH	D	2	39.945	9.187	15.069	1.00	13.06	O
	ATOM	3561	O	HOH	D	3	37.478	27.672	11.707	1.00	16.80	O
	ATOM	3564	O	HOH	D	4	44.772	-4.577	18.363	1.00	14.26	O

	ATOM	3756	O	HOH	D	68	27.701	41.270	1.720	1.00	29.57	O
	ATOM	3759	O	HOH	D	69	25.980	-9.896	14.868	1.00	27.31	O
	ATOM	3762	O	HOH	D	70	23.821	12.814	14.658	1.00	24.54	O
	ATOM	3765	O	HOH	D	71	35.006	43.534	25.083	1.00	28.85	O
5	ATOM	3768	O	HOH	D	72	35.312	36.253	1.522	1.00	29.21	O
	ATOM	3771	O	HOH	D	73	48.598	-9.901	24.935	1.00	25.55	O
	ATOM	3774	O	HOH	D	74	42.294	-13.685	16.218	1.00	25.18	O
	ATOM	3777	O	HOH	D	75	42.607	12.387	16.515	1.00	31.10	O
	ATOM	3780	O	HOH	D	76	26.330	35.006	4.050	1.00	31.74	O
	ATOM	3783	O	HOH	D	77	32.850	10.209	3.504	1.00	27.19	O
10	ATOM	3786	O	HOH	D	78	30.508	10.747	29.512	1.00	25.87	O
	ATOM	3789	O	HOH	D	79	45.693	19.098	22.237	1.00	30.36	O
	ATOM	3792	O	HOH	D	80	15.634	44.761	11.710	1.00	26.02	O
	ATOM	3795	O	HOH	D	81	18.085	50.959	3.872	1.00	35.51	O
	ATOM	3798	O	HOH	D	82	29.549	1.503	7.572	1.00	29.19	O
	ATOM	3801	O	HOH	D	83	39.725	31.841	30.695	1.00	40.77	O
15	ATOM	3804	O	HOH	D	84	20.283	36.188	-4.205	1.00	39.38	O
	ATOM	3807	O	HOH	D	85	34.763	-11.883	13.146	1.00	21.47	O
	ATOM	3810	O	HOH	D	86	26.410	32.901	7.289	1.00	24.64	O
	ATOM	3813	O	HOH	D	87	44.314	-2.758	11.932	1.00	23.95	O
	ATOM	3816	O	HOH	D	88	30.034	-14.313	17.413	1.00	29.20	O
	ATOM	3819	O	HOH	D	89	26.961	12.263	27.391	1.00	30.17	O
20	ATOM	3822	O	HOH	D	90	28.249	0.678	3.312	1.00	28.11	O
	ATOM	3825	O	HOH	D	91	45.718	32.030	14.220	1.00	34.46	O
	ATOM	3828	O	HOH	D	92	28.299	-9.696	27.995	1.00	24.79	O
	ATOM	3831	O	HOH	D	93	13.832	48.982	7.768	1.00	33.46	O
	ATOM	3834	O	HOH	D	94	43.000	-11.174	31.241	1.00	28.43	O
	ATOM	3837	O	HOH	D	95	35.944	8.335	1.385	1.00	29.40	O
25	ATOM	3840	O	HOH	D	96	29.165	29.895	11.877	1.00	24.28	O
	ATOM	3843	O	HOH	D	97	32.349	31.864	24.473	1.00	30.09	O
	ATOM	3846	O	HOH	D	98	22.954	24.601	11.686	1.00	28.72	O
	ATOM	3849	O	HOH	D	99	31.154	51.462	19.574	1.00	25.81	O
	ATOM	3852	O	HOH	D	100	43.443	12.360	23.615	1.00	24.55	O
	ATOM	3855	O	HOH	D	101	15.670	52.252	4.362	1.00	34.13	O
30	ATOM	3858	O	HOH	D	102	25.701	41.081	26.231	1.00	27.56	O
	ATOM	3861	O	HOH	D	103	37.527	21.694	22.195	1.00	32.29	O
	ATOM	3864	O	HOH	D	104	33.325	-12.660	37.738	1.00	35.18	O
	ATOM	3867	O	HOH	D	105	26.319	5.217	15.262	1.00	26.13	O
	ATOM	3870	O	HOH	D	106	33.848	22.140	26.173	1.00	31.07	O
	ATOM	3873	O	HOH	D	107	35.489	18.857	24.618	1.00	27.43	O
35	ATOM	3876	O	HOH	D	108	42.855	46.462	8.947	1.00	33.41	O
	ATOM	3879	O	HOH	D	109	42.188	5.317	9.853	1.00	30.90	O
	ATOM	3882	O	HOH	D	110	41.401	45.084	19.630	1.00	35.22	O
	ATOM	3885	O	HOH	D	111	45.990	-4.685	27.447	1.00	36.33	O
	ATOM	3888	O	HOH	D	112	44.969	4.979	13.641	1.00	30.89	O
	ATOM	3891	O	HOH	D	113	21.231	24.488	19.771	1.00	29.91	O
40	ATOM	3894	O	HOH	D	114	28.991	22.460	25.768	1.00	32.28	O
	ATOM	3897	O	HOH	D	115	30.182	42.704	28.664	1.00	34.73	O
	ATOM	3900	O	HOH	D	116	38.457	26.788	9.301	1.00	28.25	O
	ATOM	3903	O	HOH	D	117	33.010	8.247	32.080	1.00	30.38	O
	ATOM	3906	O	HOH	D	118	40.296	-12.388	19.763	1.00	29.43	O
	ATOM	3909	O	HOH	D	119	26.522	44.371	25.621	1.00	29.51	O
45	ATOM	3912	O	HOH	D	120	43.804	-4.826	10.570	1.00	33.46	O
	ATOM	3915	O	HOH	D	121	47.448	-11.680	26.748	1.00	37.40	O
	ATOM	3918	O	HOH	D	122	40.716	-13.572	24.920	1.00	24.40	O
	ATOM	3921	O	HOH	D	123	41.998	-1.274	34.849	1.00	32.74	O
	ATOM	3924	O	HOH	D	124	45.154	42.318	18.028	1.00	36.95	O
	ATOM	3927	O	HOH	D	125	30.324	-11.134	10.862	1.00	29.46	O
50	ATOM	3930	O	HOH	D	126	42.517	10.179	15.159	1.00	30.78	O
	ATOM	3933	O	HOH	D	127	48.214	-11.222	16.932	1.00	31.45	O
	ATOM	3936	O	HOH	D	128	23.815	-9.373	14.042	1.00	33.96	O
	ATOM	3939	O	HOH	D	129	31.988	24.965	29.884	1.00	32.47	O
	ATOM	3942	O	HOH	D	130	35.266	30.662	4.339	1.00	37.13	O

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ATOM	3945	O	HOH	D	131	42.057	38.530	10.976	1.00	38.75	O
ATOM	3948	O	HOH	D	132	24.900	3.888	13.671	1.00	41.30	O
ATOM	3951	O	HOH	D	133	44.797	-11.819	18.372	1.00	31.27	O
ATOM	3954	O	HOH	D	134	31.380	27.561	6.462	1.00	38.93	O
5 ATOM	3957	O	HOH	D	135	24.585	-2.131	6.886	1.00	36.52	O
ATOM	3960	O	HOH	D	136	44.178	14.598	21.666	1.00	49.82	O

CLAIMS

1. An Fve polypeptide comprising at least one biological activity of native Fve protein, and being a fragment, homologue, variant or derivative thereof.
2. An Fve polypeptide according to Claim 1, which comprises an immunomodulatory activity.
5
3. An Fve polypeptide according to Claim 1 or 2, which comprises a biological activity selected from the group consisting of: up-regulation of expression of Th1/Tc1 cytokines, preferably IFN- γ and TNF- α , down-regulation of expression of Th2/Tc2 cytokines, preferably IL-4 and IL-13, up-regulation of expression of T regulatory (Tr) cytokines IL-10 and TGF- β , hemagglutination activity, cell aggregation activity,
10 lymphocyte aggregation activity, lymphoproliferation activity, up-regulation of expression of IL-2, IFN- γ , TNF- α , but not IL-4 in CD3 $^{+}$ T cells, interaction with T and NK cells, adjuvant activity, stimulation of CD3 $^{+}$ CD16 $^{+}$ CD56 $^{+}$ natural killer (NK) T cells and CD3 $^{+}$ CD8 $^{+}$ CD18 $^{+ \text{bright}}$ T cells, and up-regulation of allergen specific Th1 immune responses.
15
3. An Fve polypeptide according to Claim 1, 2 or 3, in which the polypeptide comprises between 2 to 20 residues of amino acid sequence flanking the glycine residue corresponding to position 28 of Fve.
4. An Fve polypeptide according to any preceding claim, in which the polypeptide comprises the sequence RGT or the sequence RGD.
20
5. An Fve polypeptide according to any preceding claim, in which the polypeptide has a sequence as set out in Appendix A or Appendix B.

6. An Fve polypeptide comprising an sequence selected from the group consisting of: Fve R27A, Fve T29A, GST-Fve (wild type), GST-Fve R27A, and GST-Fve T29A, and fragments, homologues, variants and derivatives thereof.
7. A polypeptide comprising a first portion comprising at least a fragment of native Fve, or an Fve polypeptide according to any preceding claim, and a second portion comprising at least a fragment of an allergen.
5
8. A polypeptide according to Claim 7, in which the allergen comprises an allergen from a mite, preferably from Family *Glycyphagidae* or Family *Pyroglyphidae*, preferably a group 1 allergen (Der p 1, Der f 1, Blo t 1, Eur m1, Lep d 1), a group 2 allergen (Der p 2, Der f 2, Blo t 2, Eur m 2, Lep d 2), a group 5 allergen (Blo t 5, Der p 5, Der f 5, Eur m 5, Lep d 5) a group 15 allergen (Der p 15, Der f 15, Blot 15, Eur m 15, Lep d 15).
10
9. A Fve polypeptide or a polypeptide according to Claim 7 or 8, which is selected from the group consisting of: Blo t 5-Fve, Blo t 5-FveR27A, Blo t 5-FveT29A, Der p 2-FveT29A, GST-Der p 2-FveR27A, GST-Der p 2-FveT29A, Blo t 5-Der p 2-FveR27A and
15 Blo t 5-Der p 2-FveT29A.
10. A polypeptide according to Claim 7, in which the allergen is selected from the group consisting of: tree pollen allergen, Bet v 1 and Bet v 2 from birch tree; grass pollen allergen, Phl p 1 and Phl p 2 from timothy grass; weed pollen allergen, antigen E from ragweed; major feline antigen, Fel d; major fungal allergen, Asp f1, Asp f2, and Asp f3
20 from *Aspergillus fumigatus*.
11. A polypeptide comprising a first portion comprising at least a fragment of native Fve, or an Fve polypeptide according to any of Claims 1 to 6, and a second portion comprising at least a fragment of a viral antigen selected from the group consisting of: E6 and E7 from HPV; core Ag and E2 from HCV; core and surface antigens from HBV;
25 LMP-1, LMP-2, EBNA-2, EBNA-3 from EBV; and Tax from HTLV-1.

12. A polypeptide according to Claim 11, which comprises HCV Core23-FveT29A, or HPV E7-FveT29A.
13. A polypeptide comprising a first portion comprising at least a fragment of native Fve, or an Fve polypeptide according to any of Claims 1 to 6, and a second portion comprising at least a fragment of a tumour-associated antigen selected from the group consisting of: MAGE-1, MAGE-2, MAGE-3, preferably a sequence, BAGE, GAGE, PRAME, SSX-2, Tyrosinase, MART-1, NY-ESO-1, gp100, TRP-1, TRP-2, A2 melanotope, BCR/ABL, Proteinase-3/Myeloblastin, HER2/neu, CEA, P1A, HK2, PAPA, PSA, PSCA, PSMA, pg75, MUM-1, MUC-1, BTA, GnT-V, β -catenin, CDK4, and P15.
5
14. A polypeptide according to Claim 13, which comprises MAGE3-FveT29A, MART1-FveT29A or CEA-FveT29A.
10
15. A nucleic acid encoding a Fve polypeptide or a polypeptide according to any preceding claim.
16. A nucleic acid according to Claim 15, in which the nucleic acid comprises CGT GGT ACC, or a sequence which differs from the above by virtue of the degeneracy of the genetic code and which encodes a sequence RGT.
15
17. A nucleic acid comprising a first sequence encoding at least a fragment of native Fve, or an Fve polypeptide according to any of Claims 1 to 6, and a second sequence encoding at least a fragment of an allergen.
18. A nucleic acid according to Claim 17, which comprises Blo t 5-Fve, Blo t 5-FveR27A, Blo t 5-FveT29A, Der p 2-FveT29A, GST-Der p 2-FveR27A, GST-Der p 2-FveT29A, Blo t 5-Der p 2-FveR27A or Blo t 5-Der p 2-FveT29A.
20

19. A nucleic acid comprising a first sequence encoding at least a fragment of native Fve, or an Fve polypeptide according to any of Claims 1 to 6, and a second sequence encoding at least a fragment of a viral antigen selected from the group consisting of: E6 and E7 from HPV; core Ag and E2 from HCV; core and surface antigens from HBV; 5 LMP-1, LMP-2, EBNA-2, EBNA-3 from EBV; and Tax from HTLV-1.
20. A nucleic acid according to Claim 19, which comprises HCV Core23-FveT29A, or HPV E7-FveT29A.
21. A nucleic acid comprising a first sequence encoding at least a fragment of native Fve, or an Fve polypeptide according to any of Claims 1 to 6, and a second sequence 10 encoding at least a fragment of a tumour associated antigen selected from the group consisting of: MAGE-1, MAGE-2, MAGE-3, BAGE, GAGE, PRAME, SSX-2, Tyrosinase, MART-1, NY-ESO-1, gp100, TRP-1, TRP-2, A2 melanotope, BCR/ABL, Proteinase-3/Myeloblastin, HER2/neu, CEA, P1A, HK2, PAPA, PSA, PSCA, PSMA, pg75, MUM-1, MUC-1, BTA, GnT-V, β -catenin, CDK4, and P15.
- 15 22. A nucleic acid according to Claim 21, which comprises MAGE3-FveT29A, MART1-FveT29A or CEA-FveT29A.
23. A nucleic acid selected from the group consisting of: Fve R27A, Fve T29A, GST-Fve (wild type), GST-Fve R27A, GST-Fve T29A, Blo t 5-Fve, Blo t 5-FveR27A, Blo t 5-FveT29A, GST-Der p 2-FveR27A, GST-Der p 2-FveT29A, Blo t 5-Der p 2-FveR27A, Blo 20 t 5-Der p 2-FveT29A, and fragments, homologues, variants and derivatives thereof.
24. A vector, preferably an expression vector, comprising a nucleic acid sequence according to any of Claims 15 to 23.
25. A DNA vaccine comprising a nucleic acid encoding Fve, a nucleic acid according to any of Claims 15 to 23, or a vector according to Claim 24.

26. A host cell comprising a nucleic acid encoding Fve, a nucleic acid according to any of Claims 15 to 23, or a vector according to Claim 24.
27. A transgenic non-human organism comprising a nucleic acid encoding Fve, a nucleic acid according to any of Claims 15 to 23, or a vector according to Claim 24.
- 5 28. A transgenic non-human organism according to Claim 27 which is a bacterium, a yeast, a fungus, a plant or an animal, preferably a mouse.
29. A pharmaceutical composition comprising a polypeptide according to any of Claims 1 to 14, a nucleic acid according to any of Claims 15 to 23, a vector according to Claim 24, a DNA vaccine according to Claim 25, or a host cell according to Claim 26,
- 10 together with a pharmaceutically acceptable carrier or diluent.
30. Use of a native Fve polypeptide, or an Fve polypeptide, nucleic acid, vector, DNA vaccine, host cell, transgenic organism, or a pharmaceutical composition according to any of Claims 1 to 29 as an immunomodulator.
- 15 31. Use of a native Fve polypeptide, or an Fve polypeptide, nucleic acid, vector, DNA vaccine, host cell, transgenic organism, or a pharmaceutical composition according to any of Claims 1 to 29 to enhance an immune response in a mammal.
32. Use of a native Fve polypeptide, or an Fve polypeptide, nucleic acid, vector, DNA vaccine, host cell, transgenic organism, or a pharmaceutical composition according to any of Claims 1 to 29 to stimulate proliferation of CD3⁺ CD8⁺ CD18^{+bright} T cells.
- 20 33. Use of a native Fve polypeptide, or an Fve polypeptide, nucleic acid, vector, DNA vaccine, host cell, transgenic organism, or a pharmaceutical composition according to any of Claims 1 to 29 to stimulate proliferation of CD3⁺ CD16⁺ CD56⁺ natural killer (NK) T cells.

34. Use of a native Fve polypeptide, or an Fve polypeptide, nucleic acid, vector, DNA vaccine, host cell, transgenic organism, or a pharmaceutical composition according to any of Claims 1 to 29 to stimulate production of IL-2, IL-10, TGF- β , IFN- γ or TNF- α in CD3 $^{+}$ cells.
- 5 35. Use according to Claim 34, in which production of IL-4 is not stimulated in the CD3 $^{+}$ cells.
36. Use of a native Fve polypeptide, or an Fve polypeptide, nucleic acid, vector, DNA vaccine, host cell, transgenic organism, or a pharmaceutical composition according to any of Claims 1 to 29 as an adjuvant for a vaccine.
- 10 37. Use of a native Fve polypeptide, or an Fve polypeptide, nucleic acid, vector, DNA vaccine, host cell, transgenic organism, or a pharmaceutical composition according to any of Claims 1 to 29 in a method of treatment or prophylaxis of a disease.
- 15 38. Use of a native Fve polypeptide, or an Fve polypeptide, nucleic acid, vector or host cell according to any of Claims 1 to 29 for the preparation of a pharmaceutical composition for the treatment of a disease.
39. A method of treating an individual suffering from a disease or preventing the occurrence of a disease in an individual, the method comprising administering to the individual a therapeutically or prophylactically effective amount of a native Fve polypeptide, or an Fve polypeptide, nucleic acid, vector, DNA vaccine, host cell, transgenic organism, or a pharmaceutical composition according to any of Claims 1 to 29.
- 20 40. A use or method according to any of Claims 37, 38 or 39, in which the disease comprises an atopic disease or allergy.

41. Use of a DNA vaccine according to Claim 25, preferably as dependent on Claim 17 or 18, in a method of treatment or prevention of an allergy.
42. A use or method according to Claim 40 or 41, in which the allergy is selected from the group consisting of: allergic asthma, a seasonal respiratory allergy, a perennial respiratory allergy, allergic rhinitis, hayfever, nonallergic rhinitis, vasomotor rhinitis, irritant rhinitis, an allergy against grass pollen, weed pollen, tree pollen or animal danders, an allergy associated with allergic asthma and a food allergy.
43. A use or method according to Claim 40, 41 or 42, in which the allergy is to a house dust mite from Family Glyphagidae, preferably *Blomia tropicalis* or from Family Pyroglyphidae, preferably *Dermatophagoides pteronyssinus* or *Dermatophagoides farinae*, or to fungi or fungal spores, preferably *Aspergillus fumigatus*, or to tree pollen allergens, preferably from birch tree, or grass pollen allergens, preferably from timothy grass, or weed allergens, preferably ragweed.
44. A use or method according to any of Claims 37, 38 or 39, in which the disease comprises a cancer.
45. Use of a DNA vaccine according to Claim 25, preferably as dependent on Claim 19 or 20, in a method of treatment or prevention of a cancer, or in a method of suppressing tumour progression.
46. Use of a DNA vaccine according to Claim 25, preferably as dependent on Claim 21, in a method of treatment or prevention of a cancer, or in a method of suppressing tumour progression.
47. A use or method according to Claim 44, 45 or 46, in which the cancer comprises a T cell lymphoma, melanoma, lung cancer, colon cancer, breast cancer or prostate cancer.

48. A method of identifying a molecule capable of binding to Fve, the method comprising exposing a native Fve polypeptide, or an Fve polypeptide or nucleic acid, vector, host cell or transgenic organism according to any of Claims 1 to 24, 26 and 27 to a candidate molecule and detecting whether the candidate molecule binds to the native Fve polypeptide, or an Fve polypeptide or nucleic acid, vector, host cell or transgenic organism.

5
49. A method of identifying an agonist or antagonist of an Fve polypeptide, the method comprising: (a) providing a cell or organism; (b) exposing the cell or organism to a native Fve polypeptide, or an Fve polypeptide or nucleic acid, vector, host cell or transgenic organism according to any of Claims 1 to 24, 26 and 27; (c) exposing the cell to a candidate molecule; and (d) detecting an Fve mediated effect.

10
50. A method according to Claim 49, in which the Fve mediated effect is selected from the biological activities set out in Claim 2.

15
51. A method according to Claim 48, 49 or 50, in which the method further comprises isolating or synthesising a selected or identified molecule.

52. A molecule identified or selected using a method according to any of Claims 48 to 51.

53. A native Fve polypeptide, or an Fve polypeptide in crystalline form.

54. A native Fve polypeptide, or an Fve polypeptide in crystalline form according to 20 Claim 53, which has the structural coordinates shown in Appendix C.

55. A model for at least part of Fve made using a crystal according to Claim 53 or 54.

56. A method of screening for a receptor capable of binding to Fve, or designing a ligand capable of modulating the interaction between Fve and an Fve receptor, comprising the use of a model according to Claim 55.
57. A computer readable medium having stored thereon the structure of a crystal according to Claim 53 or 54, or a model according to Claim 55.
58. A ligand identified by the method according to Claim 56.
59. Use of a molecule according to Claim 52 or a ligand according to Claim 58 for the treatment or prevention of a disease in an individual.
60. A pharmaceutical composition comprising a molecule according to Claim 52 or a ligand according to Claim 58 and optionally a pharmaceutically acceptable carrier, diluent, excipient or adjuvant or any combination thereof.
61. A method of treating and/or preventing a disease comprising administering a molecule according to Claim 52 or a ligand according to Claim 58 and/or a pharmaceutical composition according to Claim 60 to an individual in need of such treatment.
- 15 62. A method of amplifying a sub-population of cells, the method comprising: (a) obtaining a population of cells from an individual; (b) amplifying CD3⁺ CD8⁺ and CD18^{+ bright} T cells by exposing the population of cells to a native Fve polypeptide, or an Fve polypeptide or nucleic acid, vector, host cell or transgenic organism according to any of Claims 1 to 24, 26 and 27.
- 20 63. A method according to Claim 62, further comprising the step of: (c) isolating the CD3⁺ CD8⁺ and CD18^{+ bright} T cells.

64. A method of treating an individual suffering from a disease or preventing the occurrence of a disease in an individual, the method comprising amplifying a CD3⁺ CD8⁺ and CD18^{+ bright} T cell by a method according to Claim 62 or 63, and administering the amplified CD3⁺ CD8⁺ and CD18^{+ bright} T cell to an individual.
- 5 65. An amplified population of CD3⁺ CD8⁺ and CD18^{+ bright} T cells obtainable by a method according to Claim 62 or 63.
66. A pharmaceutical composition comprising an amplified population of CD3⁺ CD8⁺ and CD18^{+ bright} T cells according to Claim 65, together with a pharmaceutically acceptable excipient or carrier.
- 10 67. A combination comprising a first component comprising an immunomodulator and a second component comprising at least a fragment of an allergen, a viral antigen or a tumour associated antigen.
68. A combination according to Claim 67 in which the first component is separate from the second component.
- 15 69. A combination according to Claim 67 in which the first component is associated with the second component.
70. A combination according to Claim 67 which is a fusion protein.
71. A combination according to Claim 67, in which the first component comprises a native Fve polypeptide, or a polypeptide according to any of Claims 1 to 14.
- 20 72. A combination according to any of Claims 67 to 71, in which the second component comprises an allergen selected from the group consisting of: a mite allergen, an mite allergen from Family *Glycyphagidae* or Family *Pyroglyphidae*, a group 1 allergen

(Der p 1, Der f 1, Blo t 1, Eur m1, Lep d 1), a group 2 allergen (Der p 2, Der f 2, Blo t 2, Eur m 2, Lep d 2), a group 5 allergen (Blo t 5, Der p 5, Der f 5, Eur m 5, Lep d 5), a group 15 allergen (Der p 15, Der f 15, Blot 15, Eur m 15, Lep d 15), a tree pollen allergen, Bet v 1 and Bet v 2 from birch tree; grass pollen allergen, Phl p 1 and Phl p 2 from timothy grass; weed pollen allergen, antigen E from ragweed; major feline antigen, Fel d; major fungal allergen, Asp f1, Asp f2, and Asp f3 from *Aspergillus fumigatus*.

73. A combination according to any of Claims 67 to 71, in which the second component comprises a viral antigen selected from the group consisting of: E6 and E7 from HPV; core Ag and E2 from HCV; core and surface antigens from HBV; LMP-1, LMP-2, EBNA-2, EBNA-3 from EBV; and Tax from HTLV-1.

74. A combination according to any of Claims 67 to 71, in which the second component comprises a tumour-associated antigen selected from the group consisting of: MAGE-1, MAGE-2, MAGE-3, BAGE, GAGE, PRAME, SSX-2, Tyrosinase, MART-1, NY-ESO-1, gp100, TRP-1, TRP-2, A2 melanotope, BCR/ABL, Proteinase-3/Myeloblastin, HER2/neu, CEA, P1A, HK2, PAPA, PSA, PSCA, PSMA, pg75, MUM-1, MUC-1, BTA, GnT-V, β-catenin, CDK4, and P15.

75. An immunomodulator-antigen conjugate, preferably an immunomodulator-allergen conjugate, an immunomodulator-tumour associated antigen conjugate or a immunomodulator-viral antigen conjugate, in which the immunomodulator preferably comprises an Fve polypeptide.

76. A polypeptide comprising a first portion comprising at least a fragment of native Fve, or an Fve polypeptide according to any of Claims 1 to 6, and a second portion comprising at least a fragment of a viral antigen selected from the group consisting of an antigen from Adenovirus, Parainfluenza 3 virus, Human Immunodeficiency Virus (HIV), Herpes simplex virus, HSV, Respiratory syncytial virus, RSV, or Influenza A, Flu A.

77. A nucleic acid comprising a first sequence encoding at least a fragment of native Fve, or an Fve polypeptide according to any of Claims 1 to 6, and a second sequence encoding at least a fragment of a viral antigen selected from the group consisting of an antigen from from Adenovirus, Parainfluenza 3 virus, Human Immunodeficiency Virus (HIV), Herpes simplex virus, HSV, Respiratory syncytial virus, RSV, or Influenza A, Flu A.
- 5
78. A combination according to any of Claims 67 to 71, in which the second component comprises a tumour-associated antigen selected from the group consisting of antigen from from Adenovirus, Parainfluenza 3 virus, Human Immunodeficiency Virus (HIV), Herpes simplex virus, HSV, Respiratory syncytial virus, RSV, or Influenza A, Flu A.
- 10
79. A nucleic acid sequence, including an Fve nucleic acid sequence, a polypeptide sequence, including a Fve polypeptide sequence, a method of treatment, a method of diagnosis, a host cell, vector, transgenic animal, a transgenic plant, a genetically-modified lactose bacilli, assay, vaccine, pharmaceutical composition or agent substantially as hereinbefore described with reference to and as shown in the accompanying drawings.
- 15

**ABSTRACT
MOLECULES**

We describe an Fve polypeptide being a fragment, homologue, variant or derivative of Fve protein, which comprises at least one biological activity of Fve protein. uses of such 5 a polypeptide, etc, and nucleic acids encoding these, in the treatment and prevention of allergy and cancer are also disclosed.

Figure 1

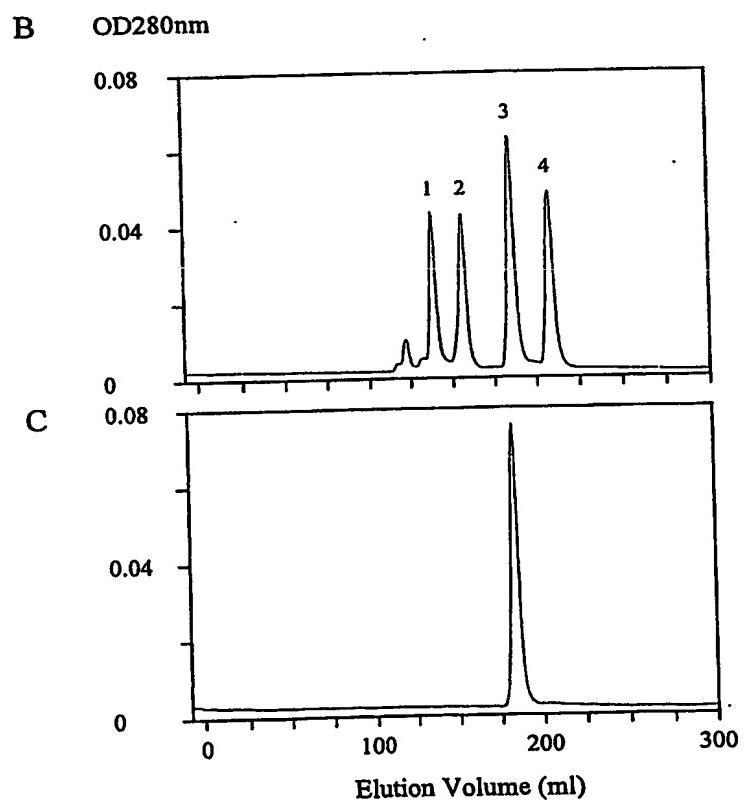
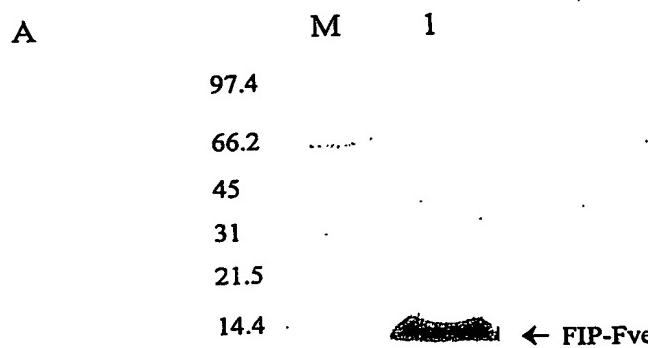


FIGURE 1

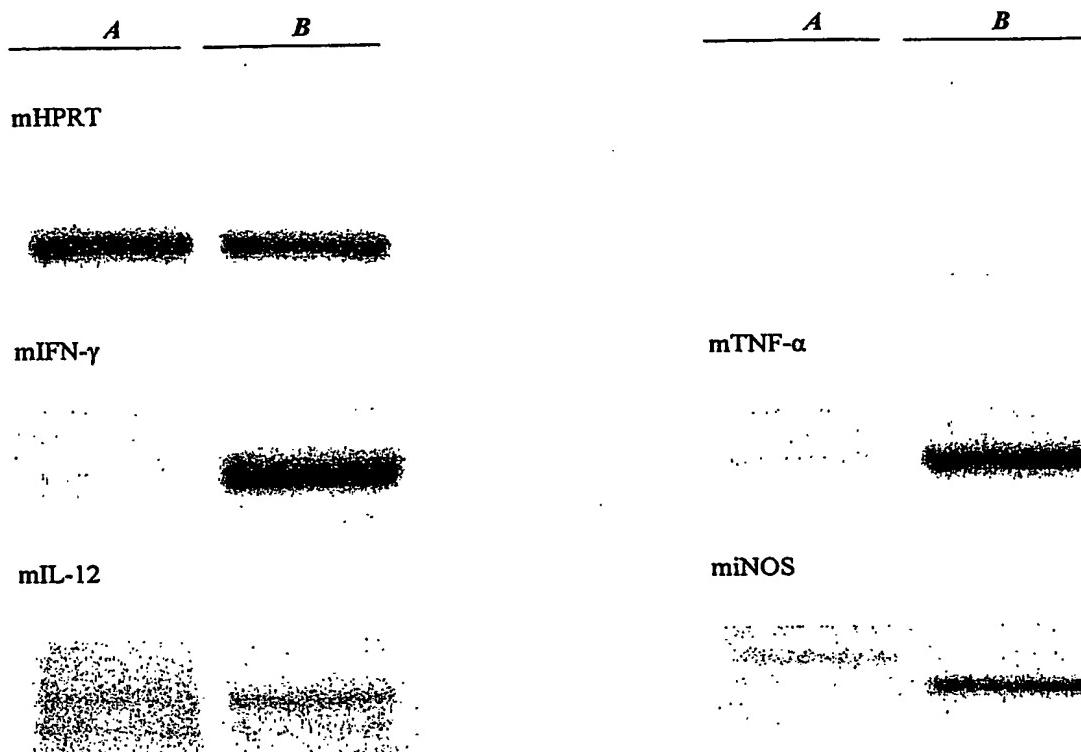


FIGURE 2

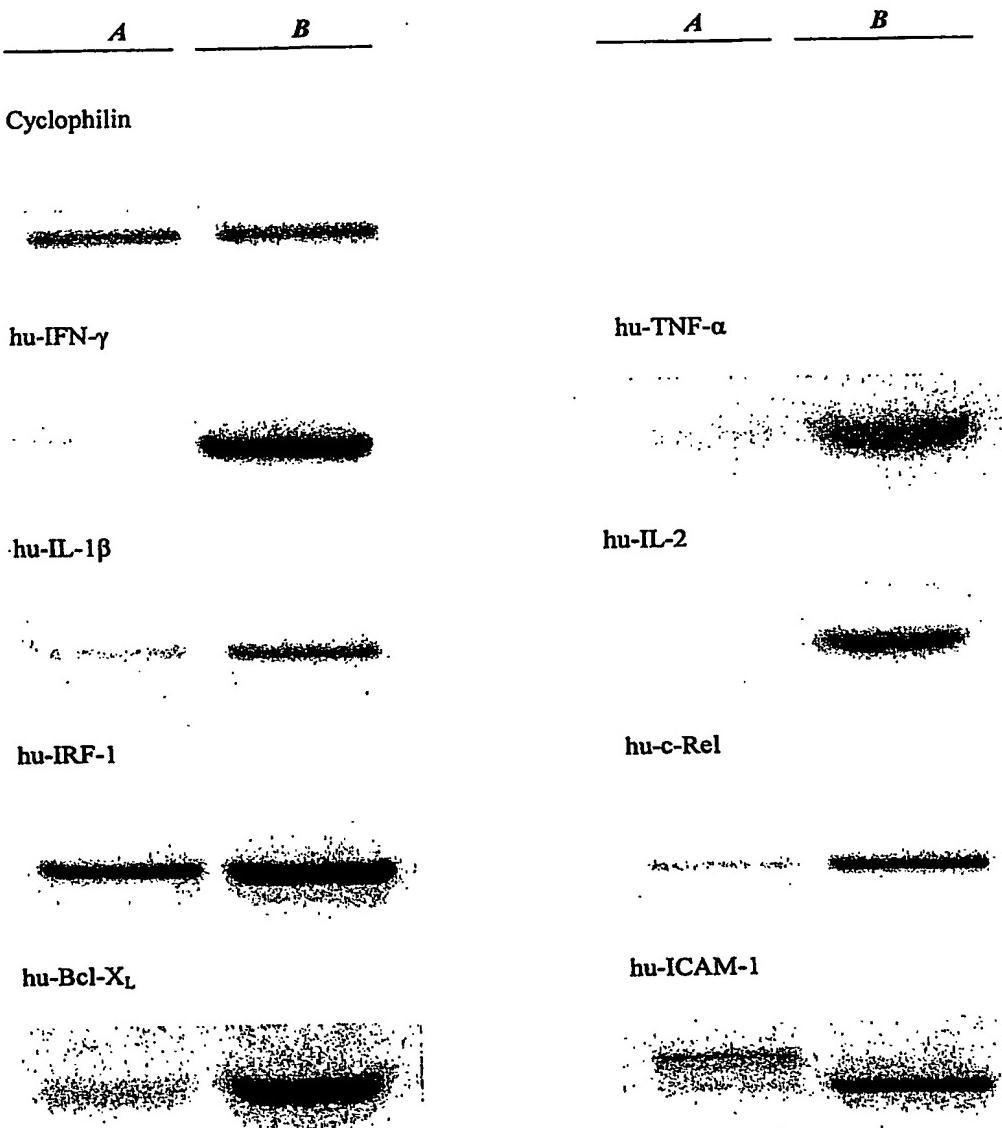


FIGURE 3

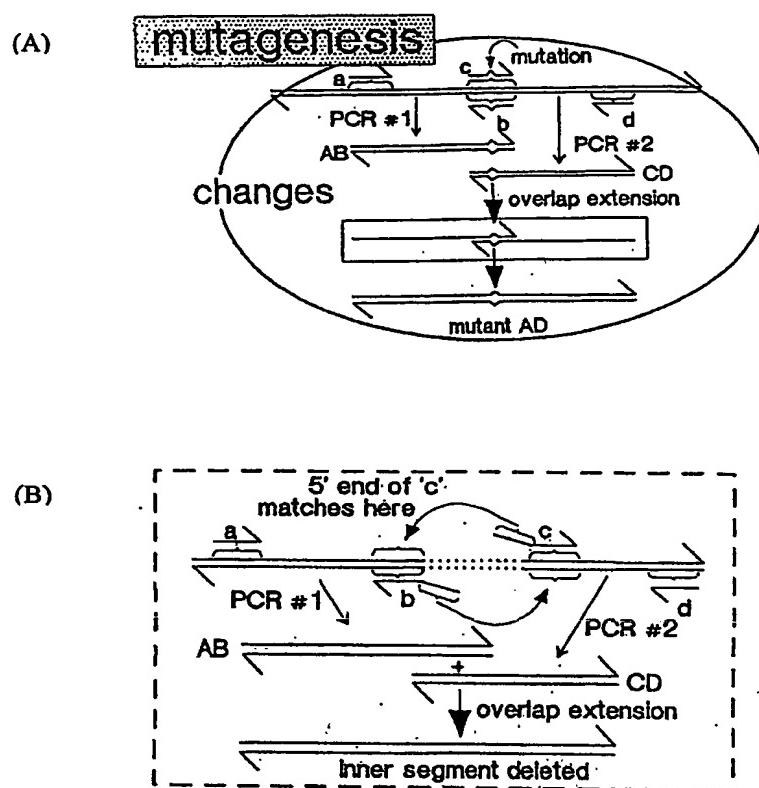
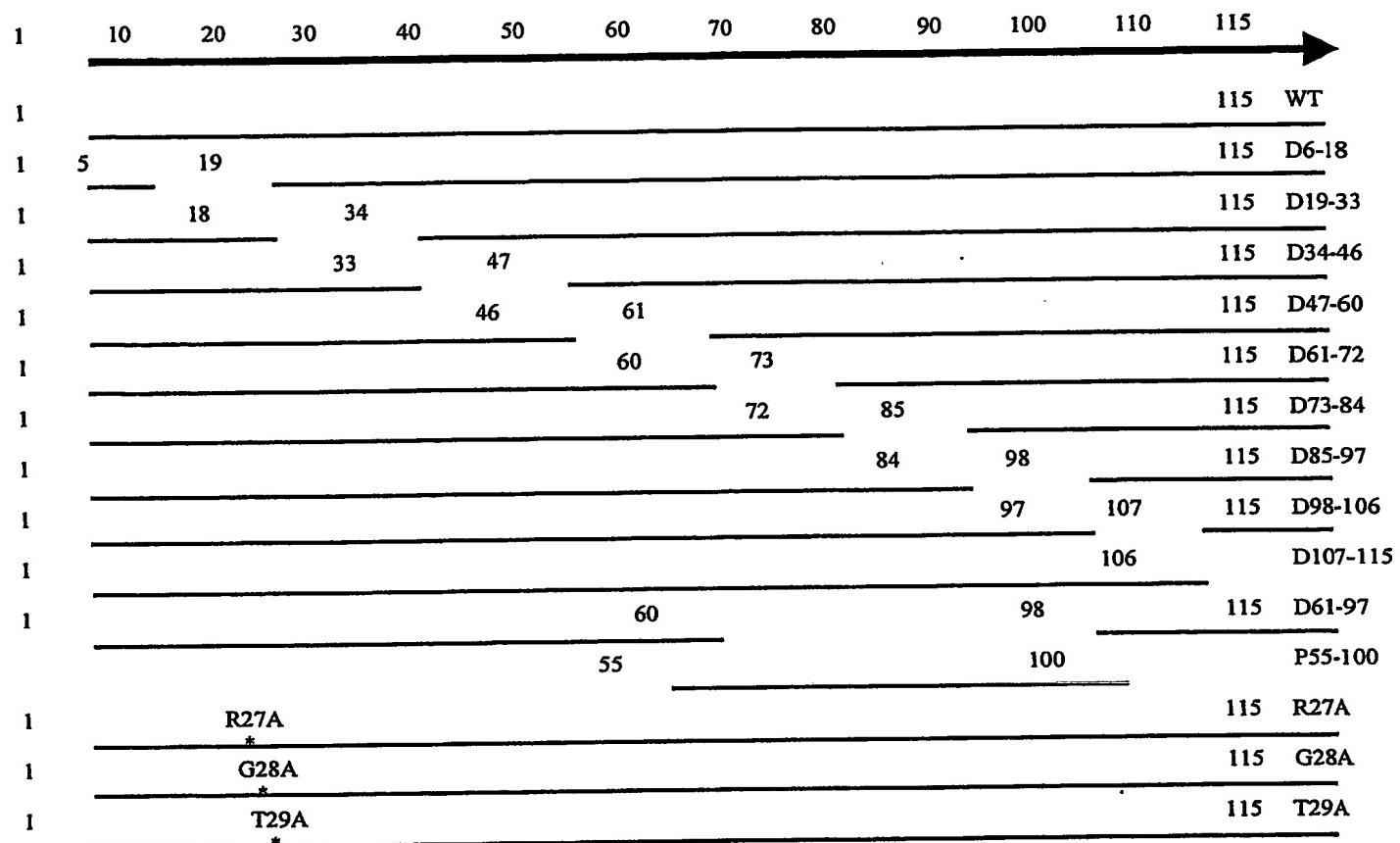


FIGURE 4

**FIGURE 5**

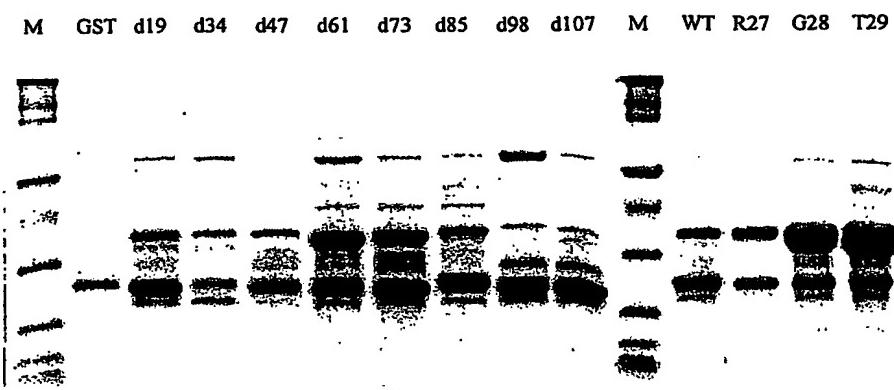


FIGURE 6

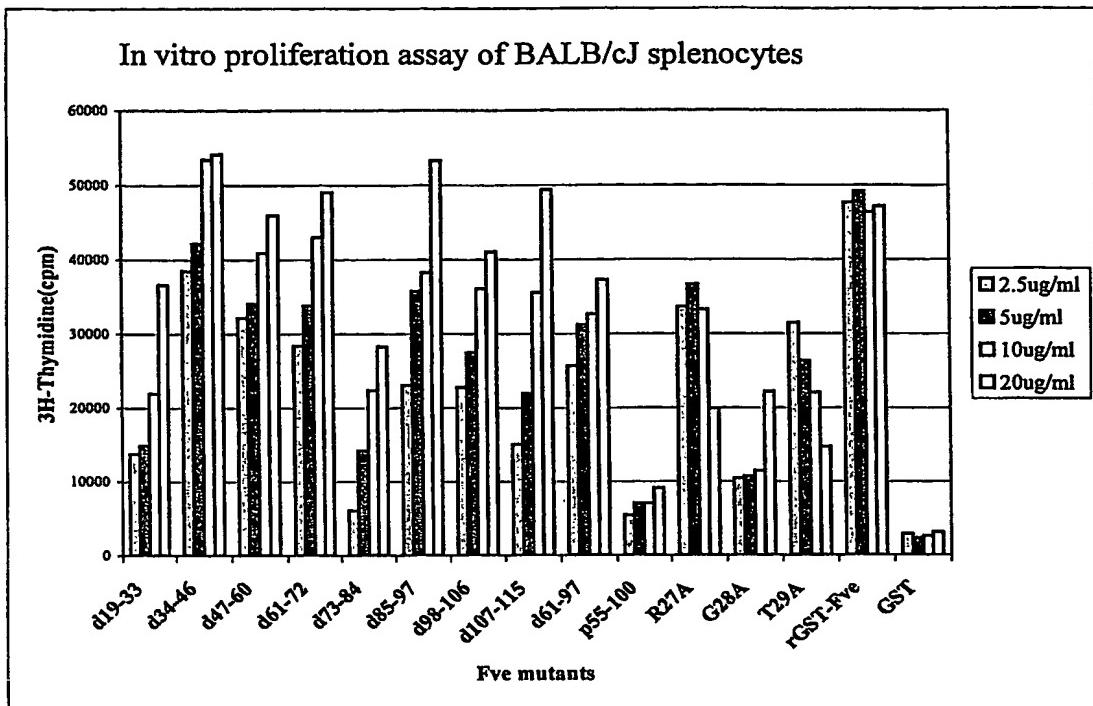


FIGURE 7

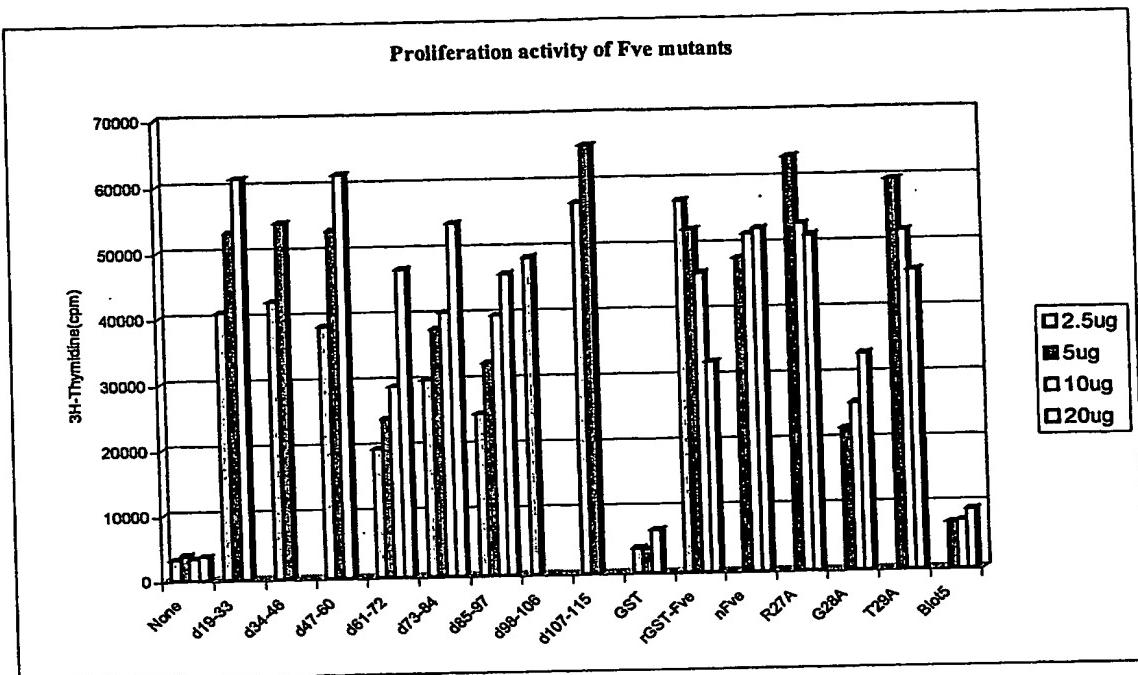
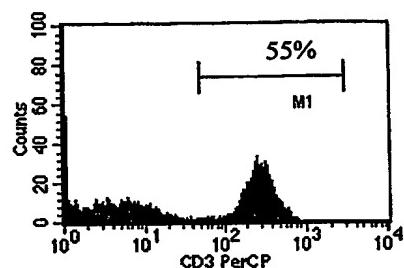
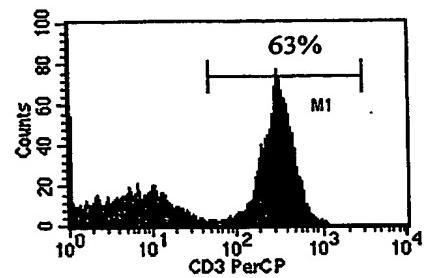


FIGURE 8

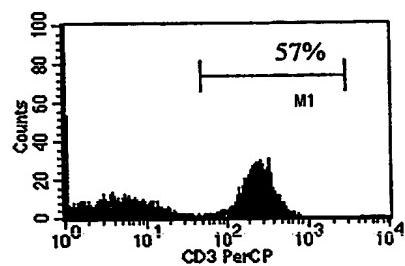
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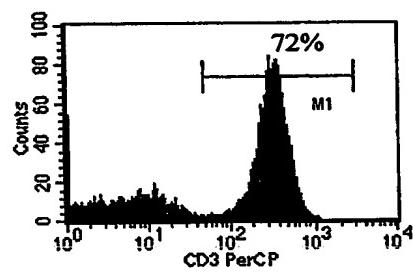
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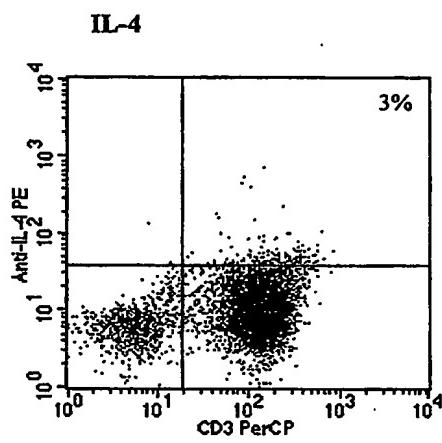
(B).



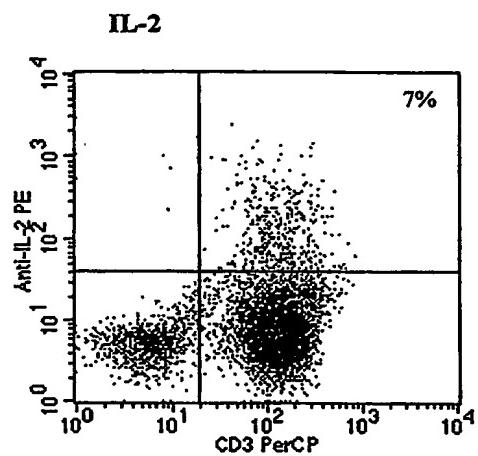
(D).

**FIGURE 9**

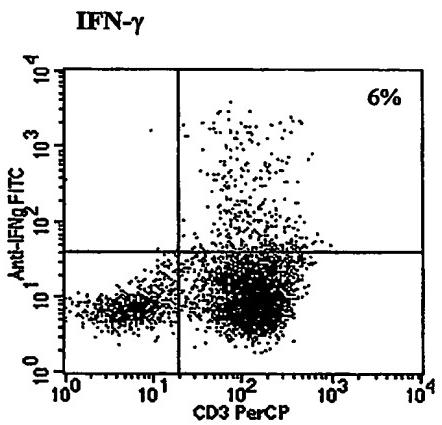
(A).



(B).



(C).



(D).

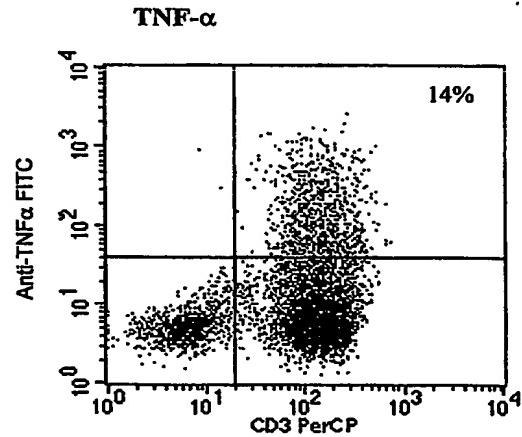
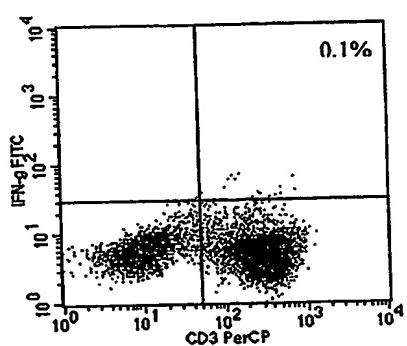


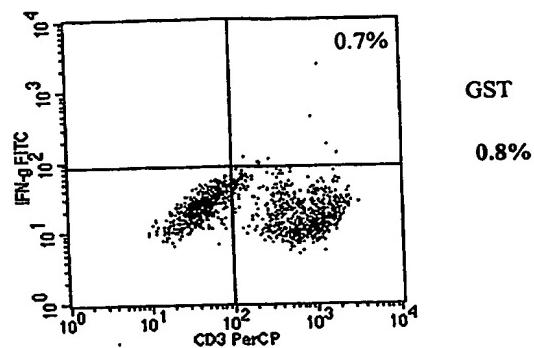
FIGURE 10

IFN- γ production at day 3

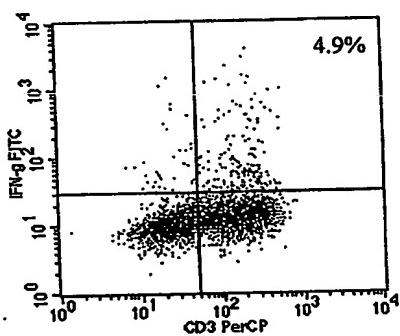
(1a).



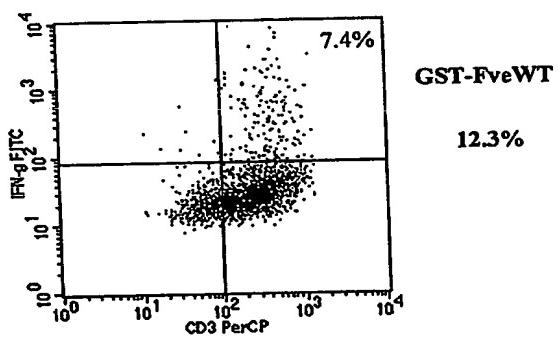
(1b).



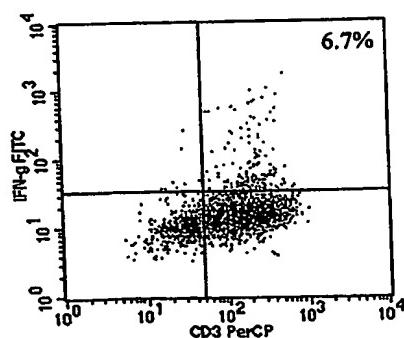
(2a).



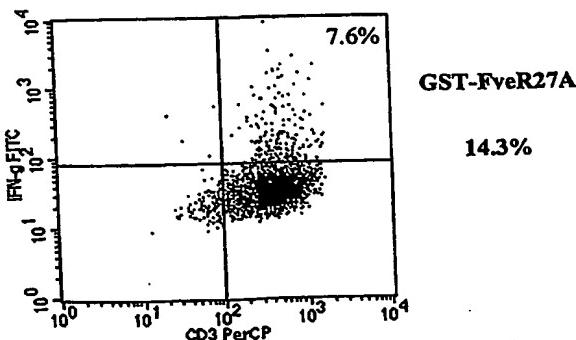
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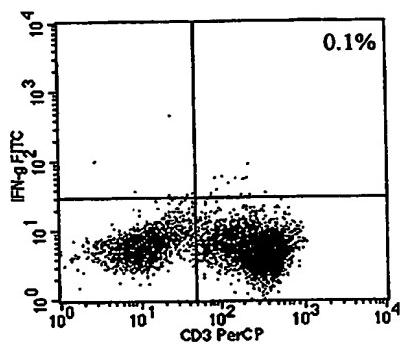
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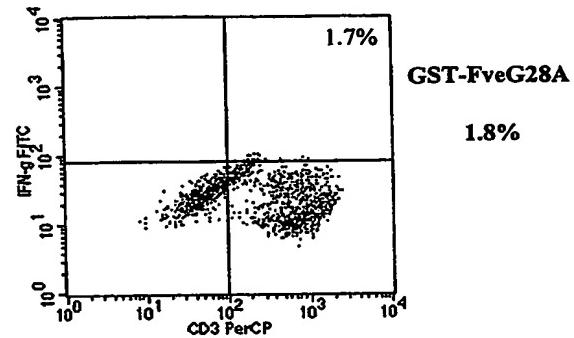
(3b).

**FIGURE 11**

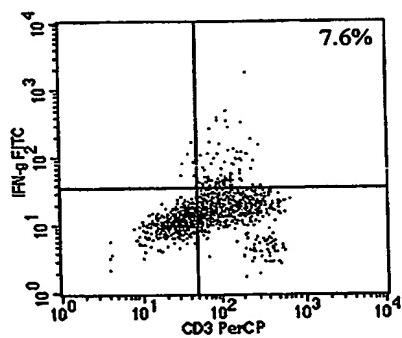
(4a).



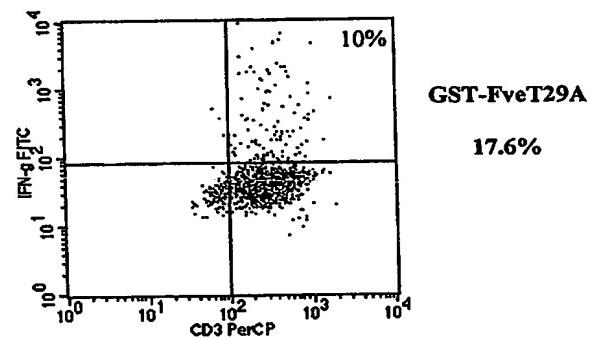
(4b).



(5a).

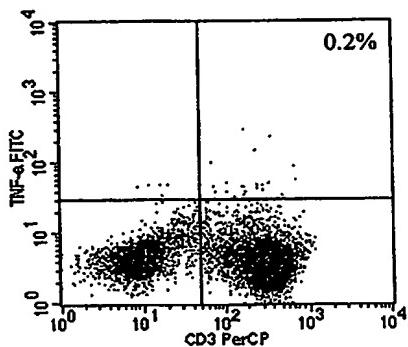


(5b).

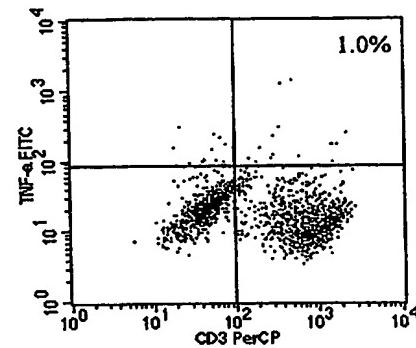
**FIGURE 11 (CONTINUED)**

TNF- α production at day 3

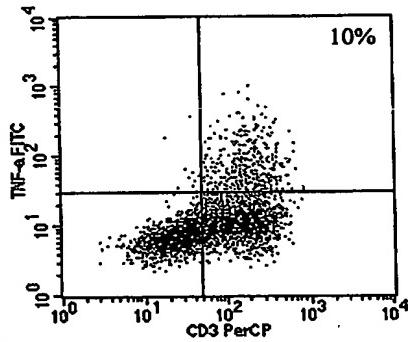
(1a).



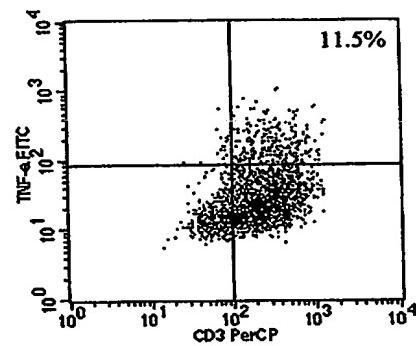
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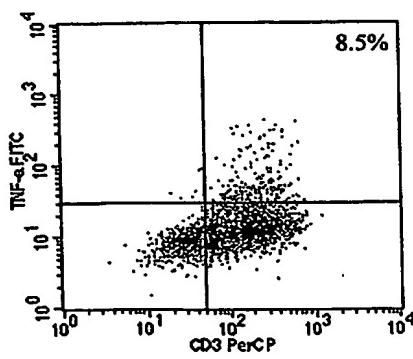
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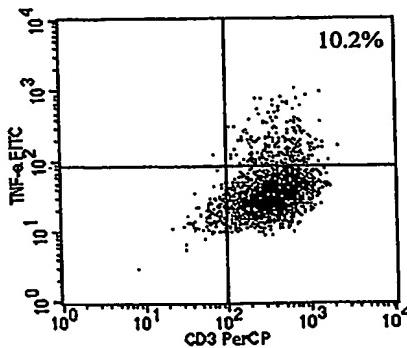
(2b).



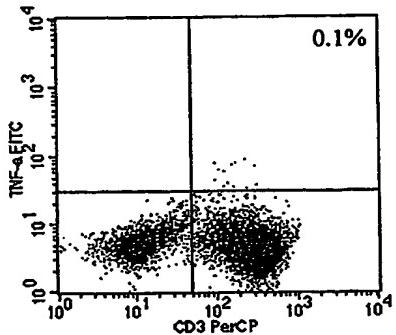
(3a).



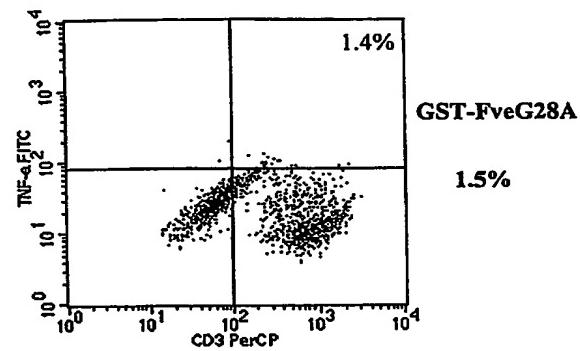
(3b).

**FIGURE 12**

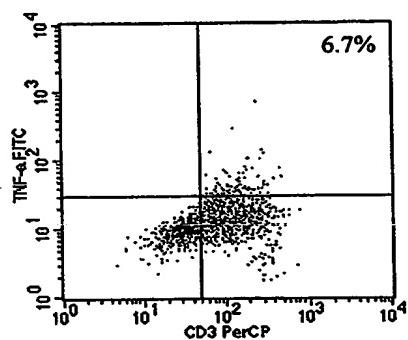
(4a).



(4b).



(5a).



(5b).

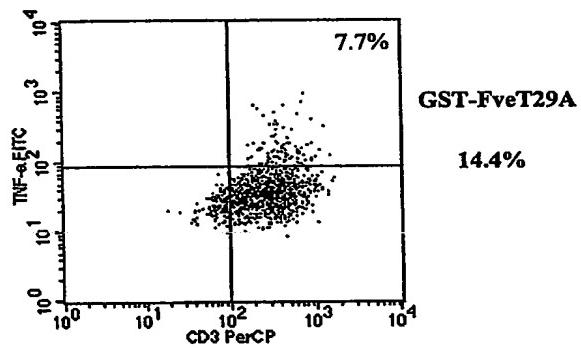


FIGURE 12 (CONTINUED)

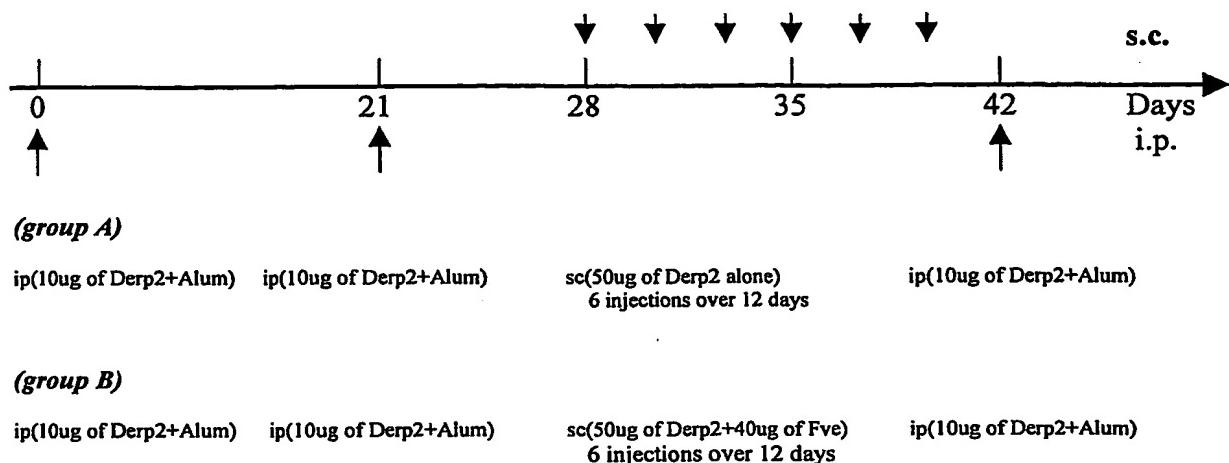


FIGURE 13

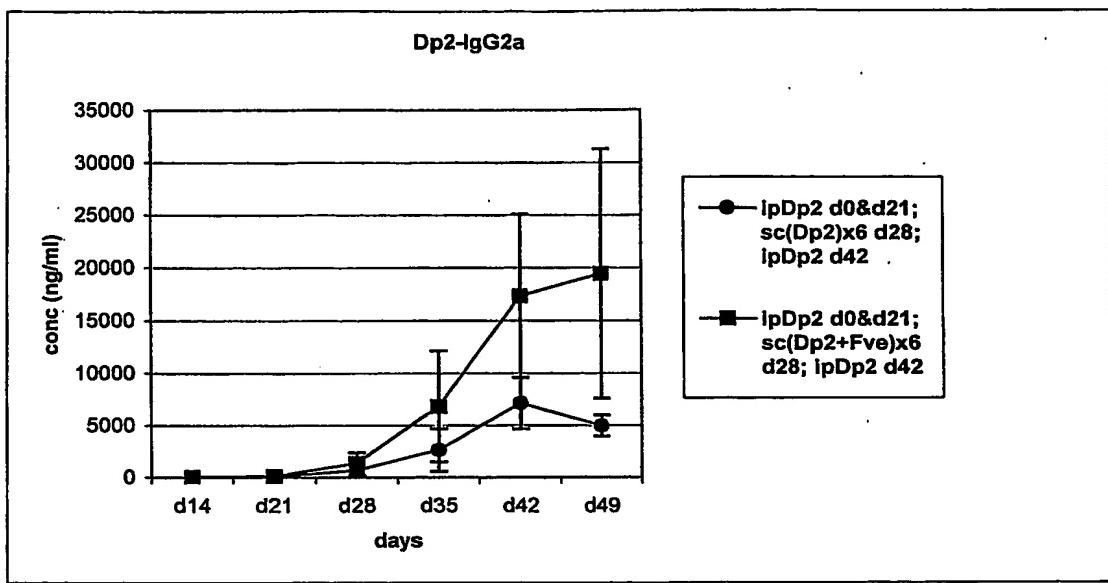
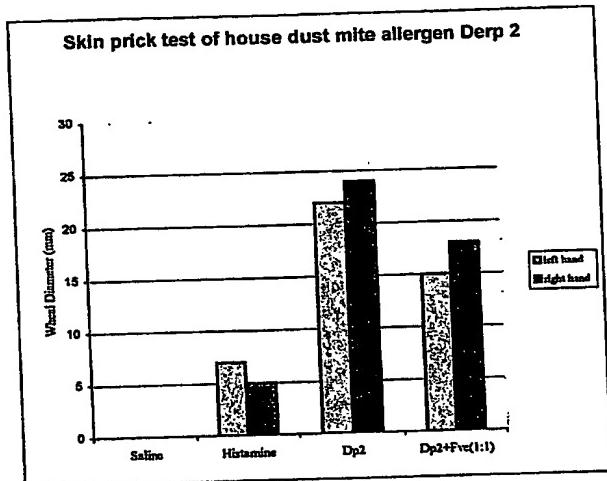


FIGURE 14

(A)



(B)

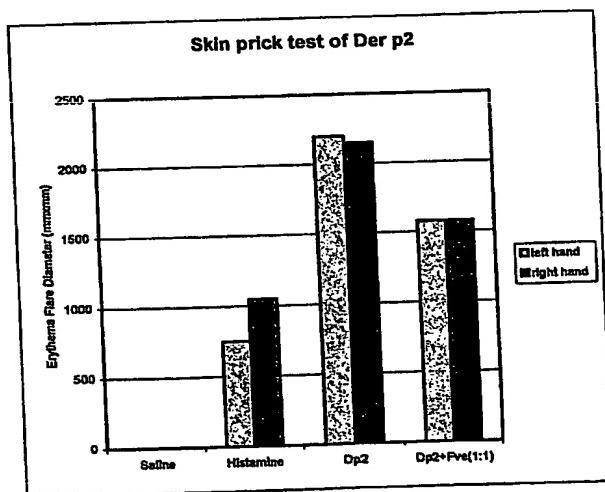


FIGURE 15

Blo t 5	Fve	Bt5-Fve	
Blo t 5	FveR27A	Bt5-FveR27A	
Blo t 5	FveT29A	Bt5-FveT29A	
Der p 2	FveR27A	Dp2-FveR27A	
Der p 2	FveT29A	Dp2-FveT29A	
Blo t 5	Der p 2	FveR27A	Bt5-Dp2-FveR27A
Blo t 5	Der p 2	FveT29A	Bt5-Dp2-FveT29A

FIGURE 16

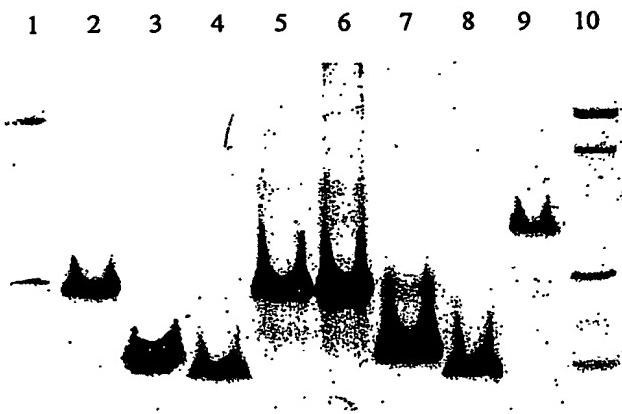
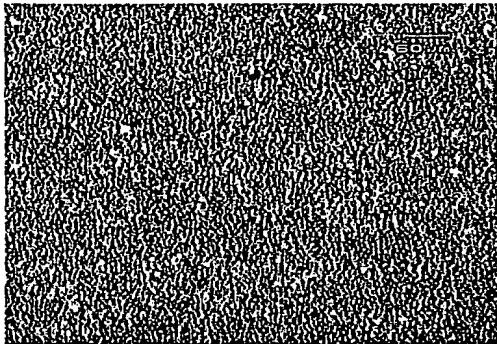


FIGURE 17

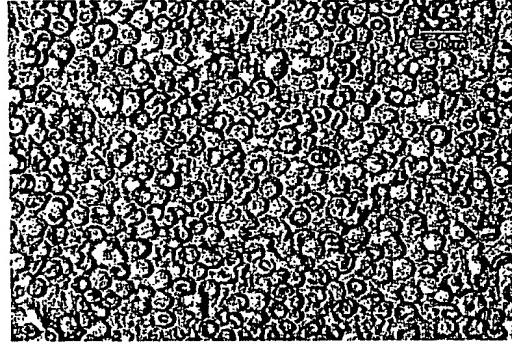
(1a)

Control: Non-stimulated (10x10 magnification)



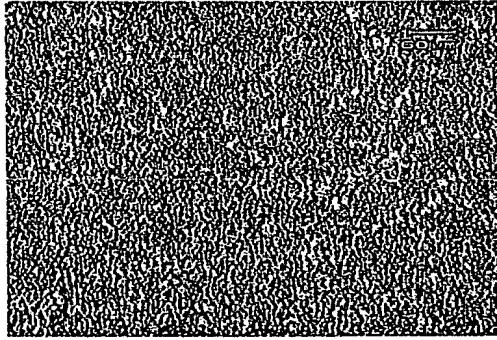
(1b)

Control : Non-stimulated (40x10 magnification)



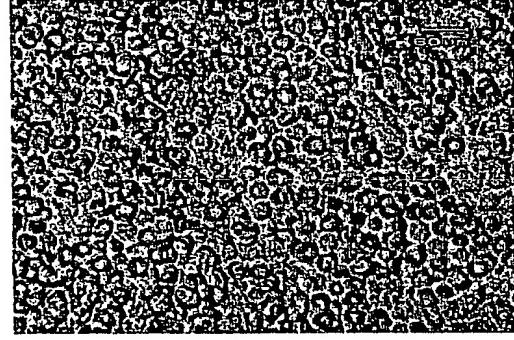
(2a)

20ug of GST 10x10



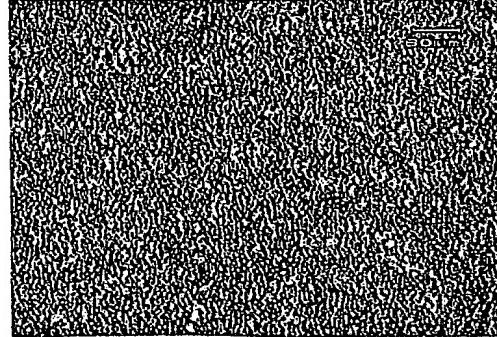
(2b)

20ug of GST 40x10



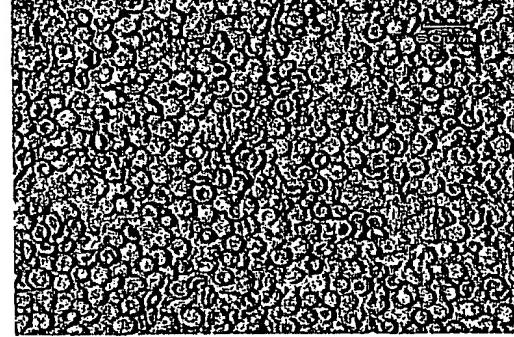
(3a)

20ug of Blo t 5 10x10



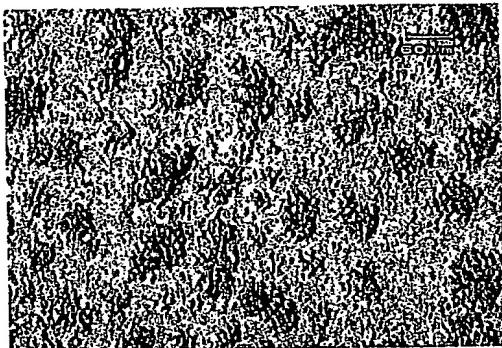
(3b)

20ug of Blo t 5 40x10

**FIGURE 18**

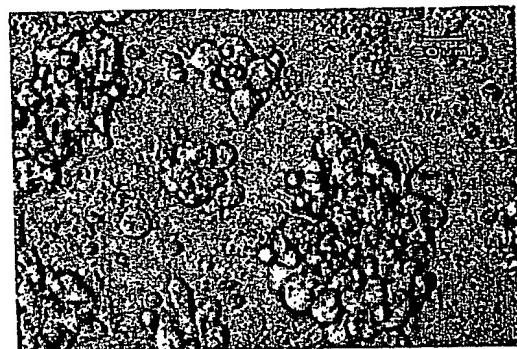
(4a)

20ug of native FIP-Fve 10x10



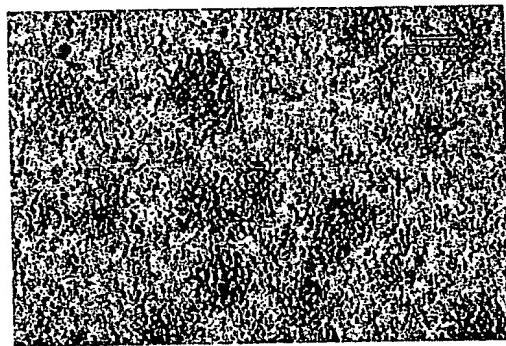
(4b)

20ug of native FIP-Fve 40x10



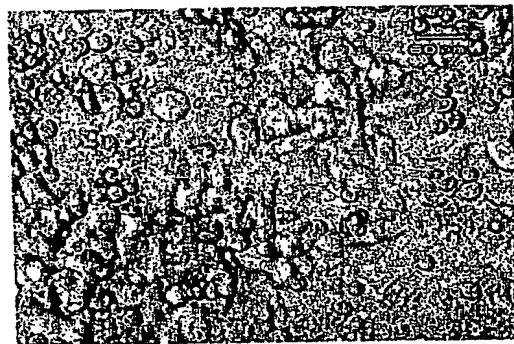
(5a)

20ug of Bt5-Fve 10x10



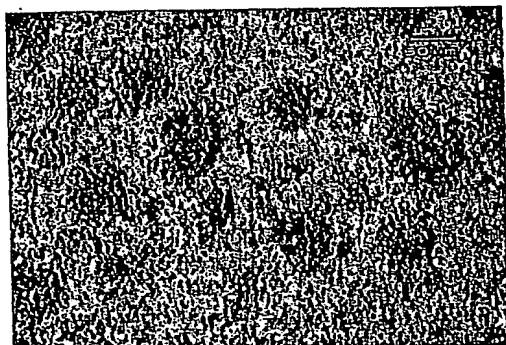
(5b)

20ug of Bt5-Fve 40x10



(6a)

40ug of Bt5-Fve 10x10



(6b)

40ug of Bt5-Fve 40x10

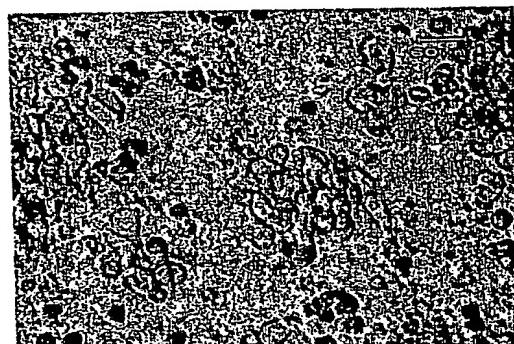
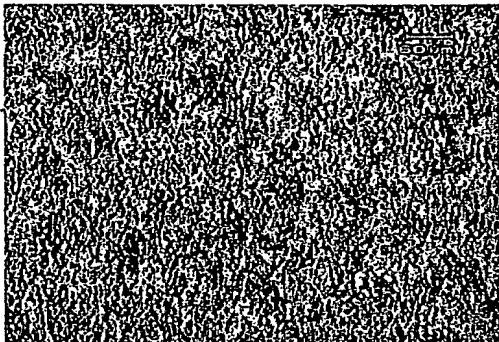


FIGURE 18 (CONTINUED)

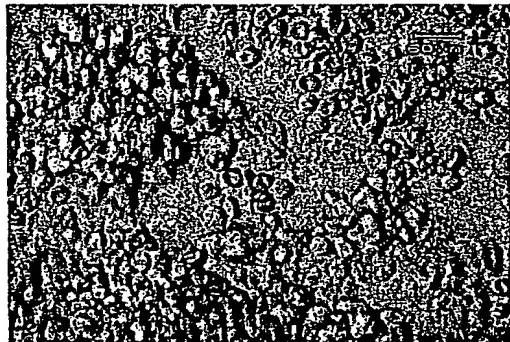
(7a)

40ug of Bt5-FveR27A 10x10



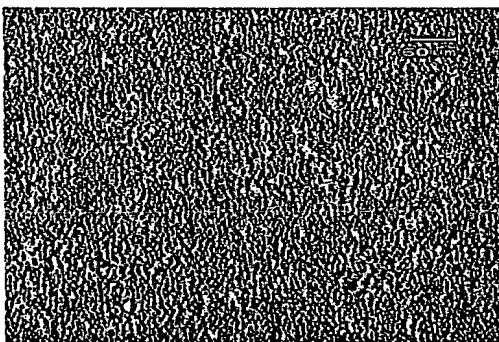
(7b)

40ug of Bt5-FveR27A 40x10



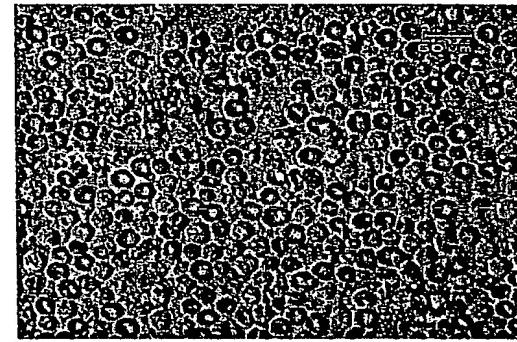
(8a)

20ug of Der p 2 10x10



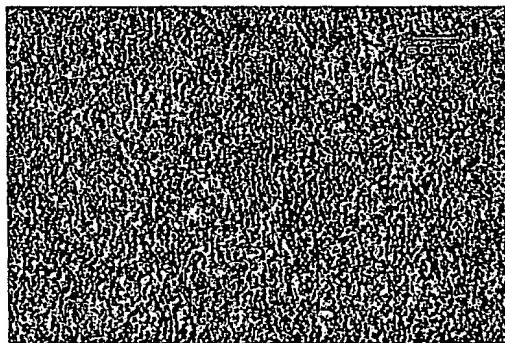
(8b)

20ug of Der p 2 40x10



(9a)

40ug of GST-Dp2-FveR27A 10x10



(9b)

40ug of GST-Dp2-FveR27A 40x10

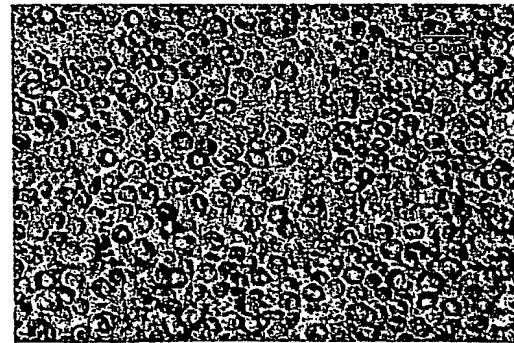
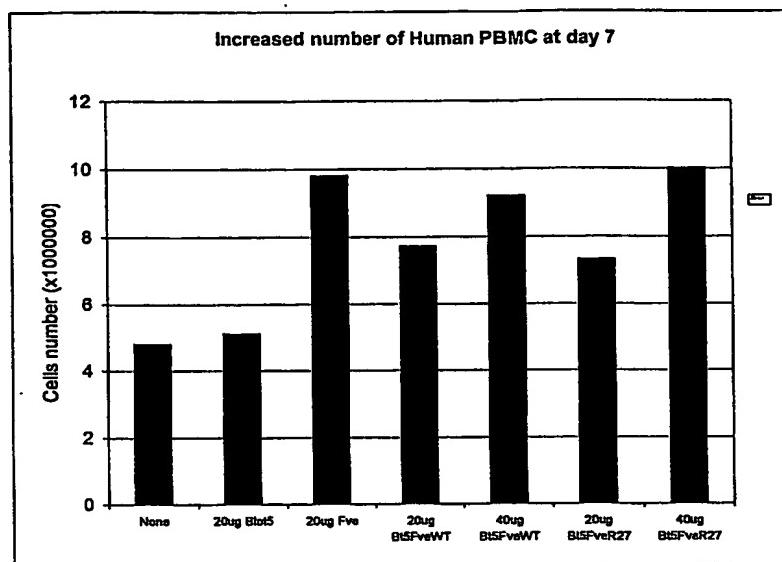


FIGURE 18 (CONTINUED)

(A)



(B)

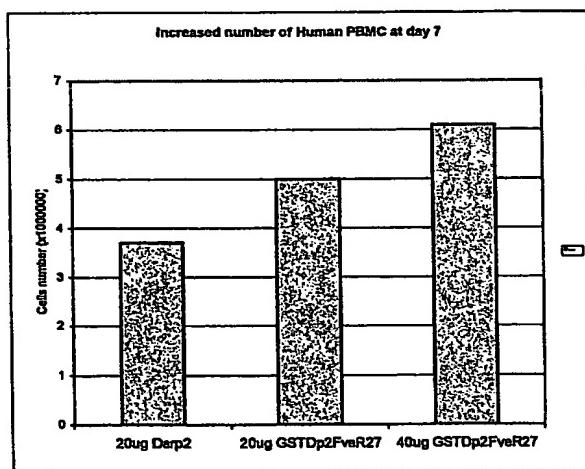


FIGURE 19

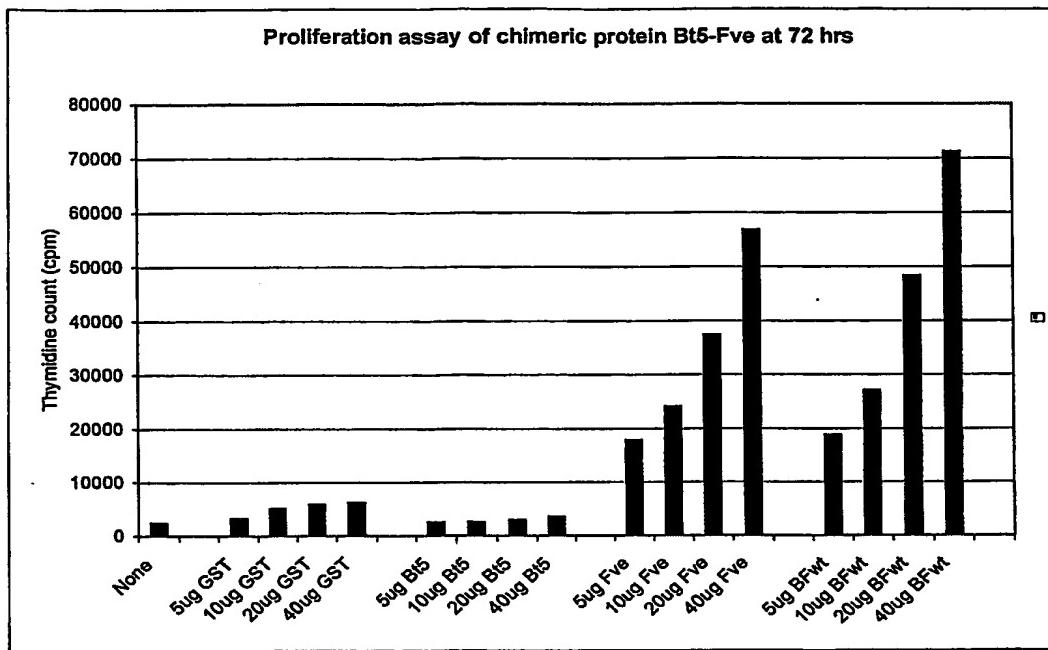


FIGURE 20

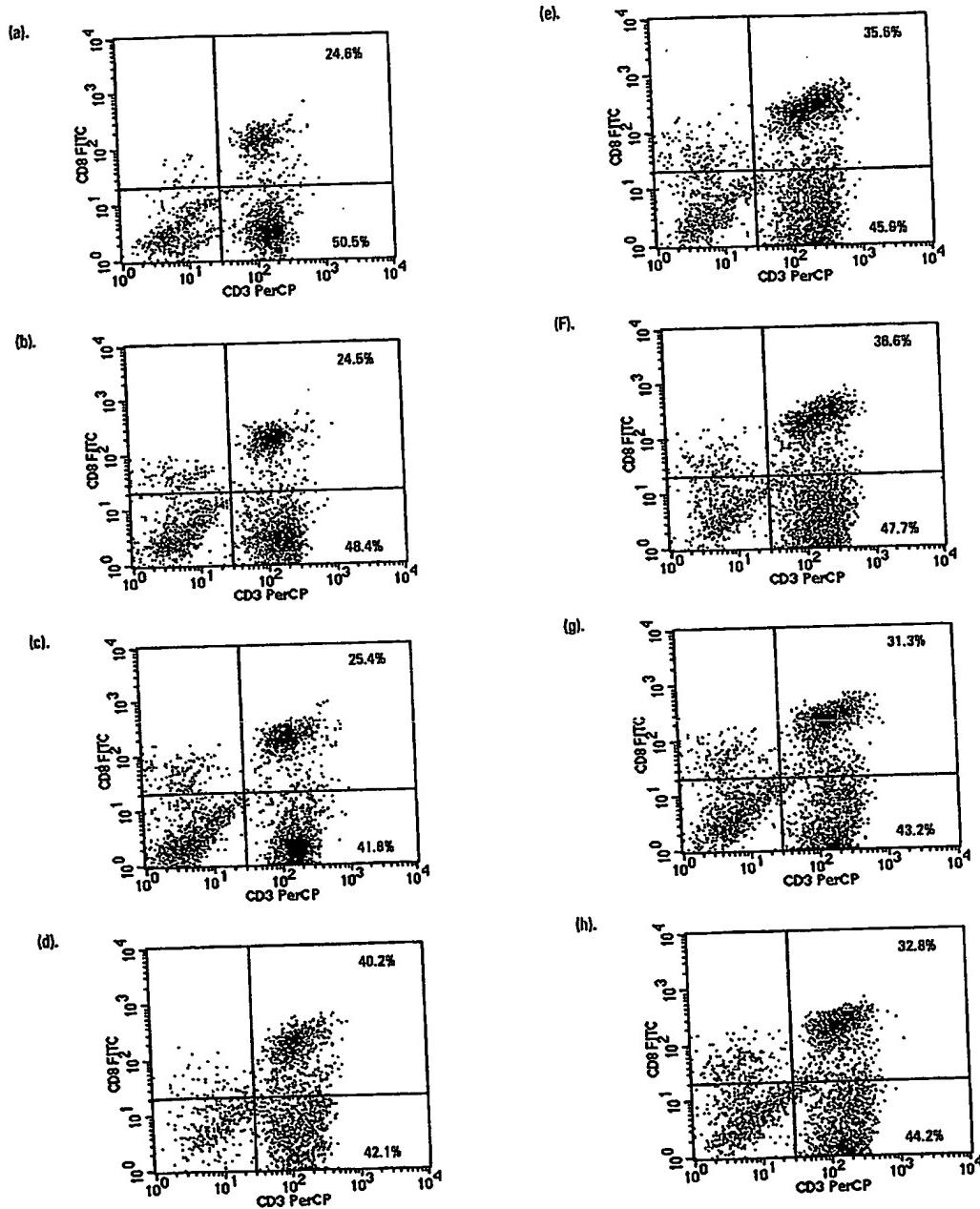
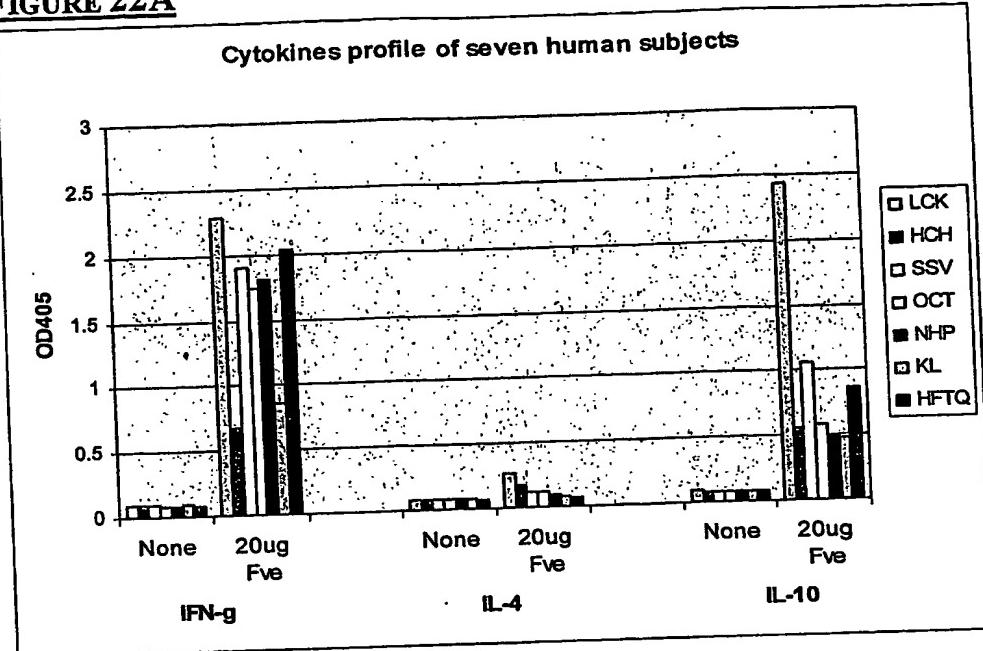
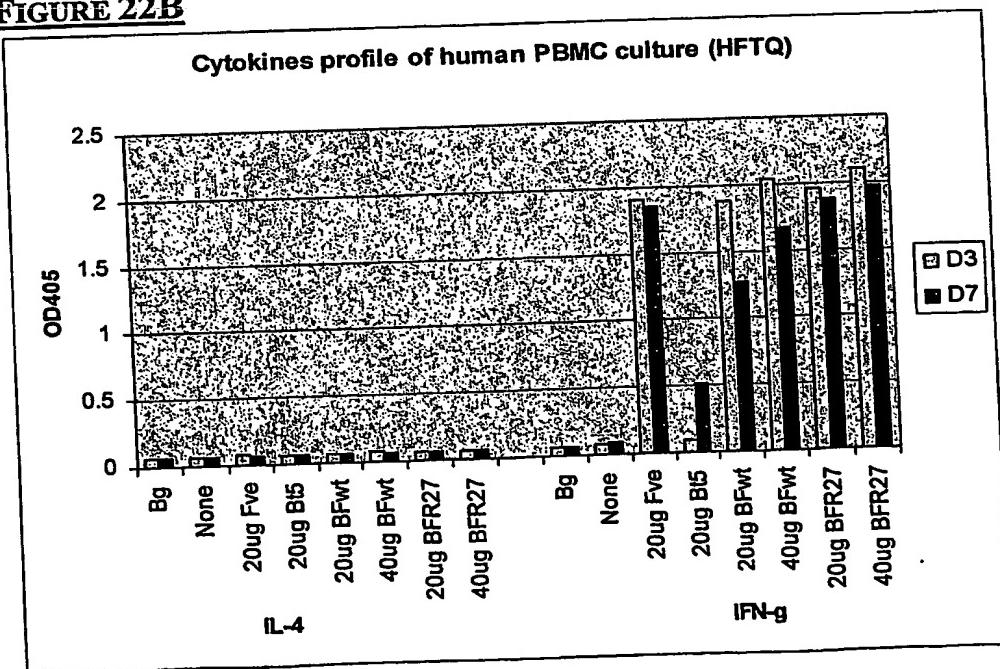
FIGURE 21

FIGURE 22A**FIGURE 22B**

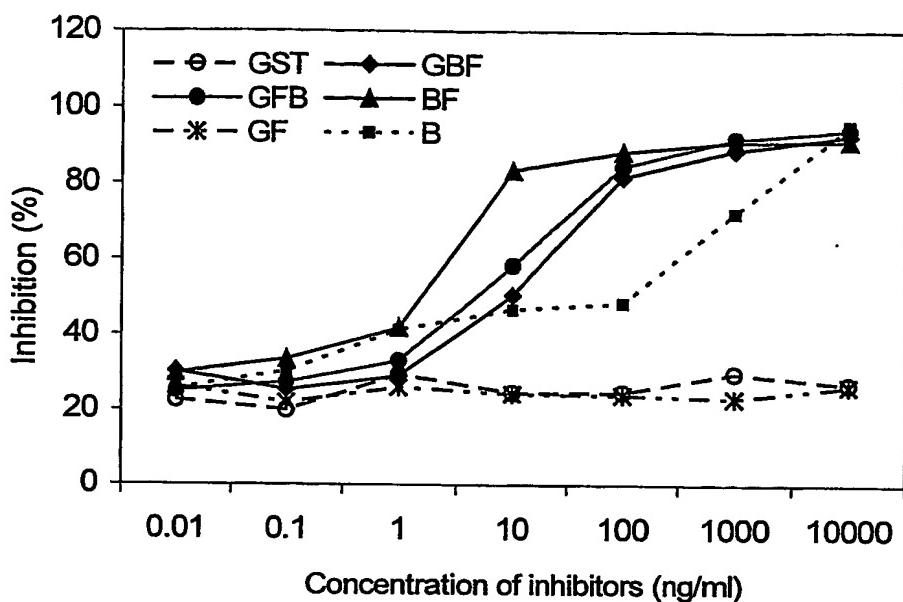
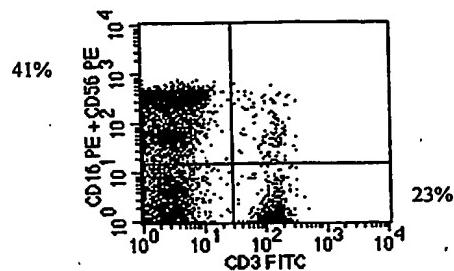
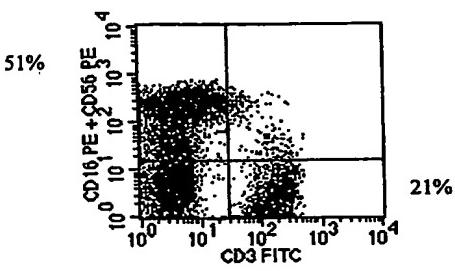


FIGURE 23

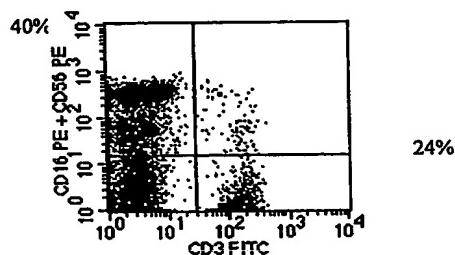
(a).



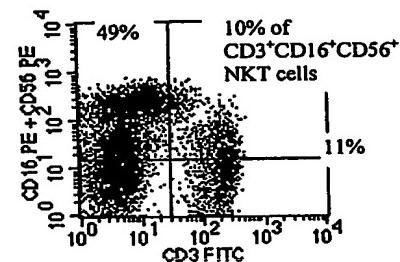
(d).



(b).



(e).



(c).

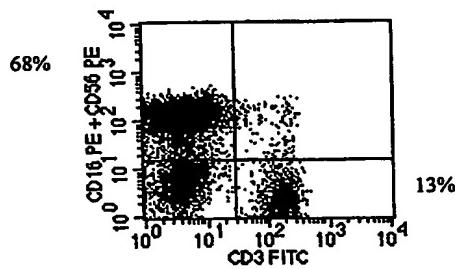
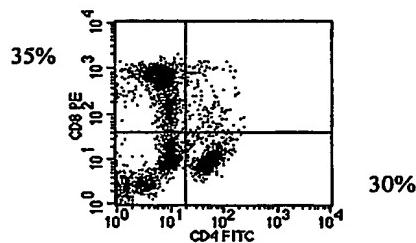
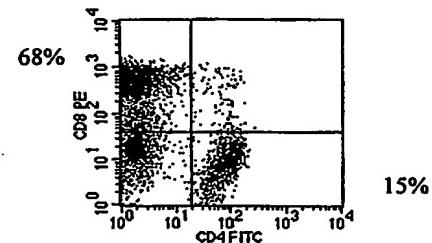


FIGURE 24

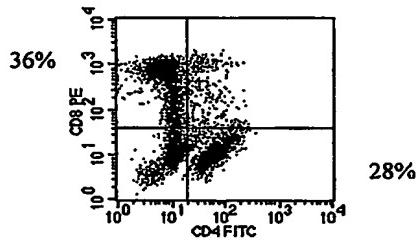
(a).



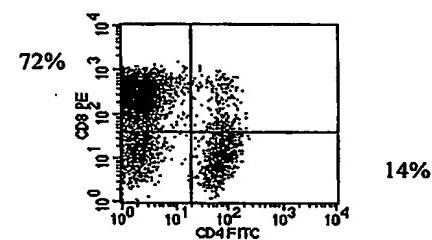
(d).



(b).



(e).



(c).

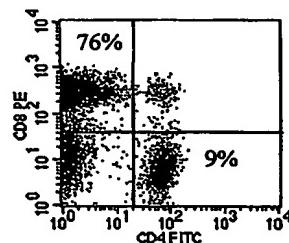
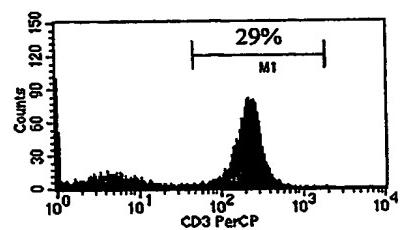
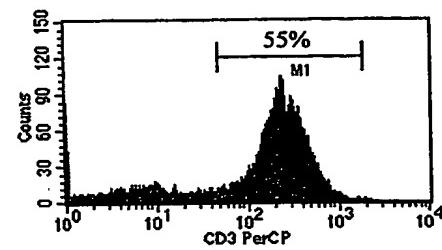


FIGURE 25

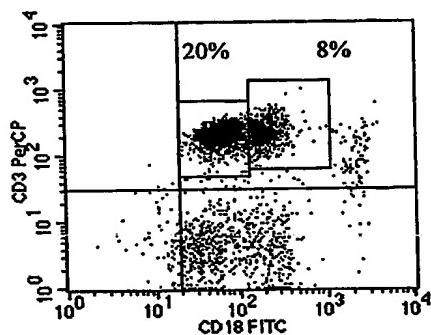
(a).



(b).



(c).



(d).

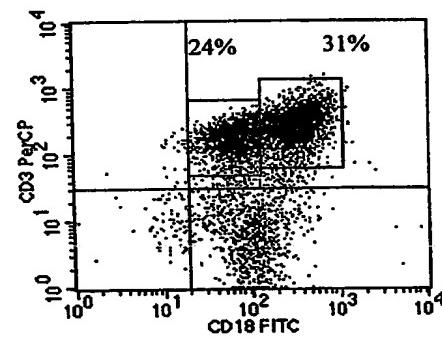
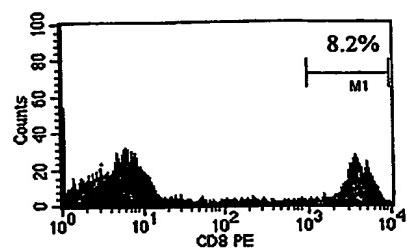
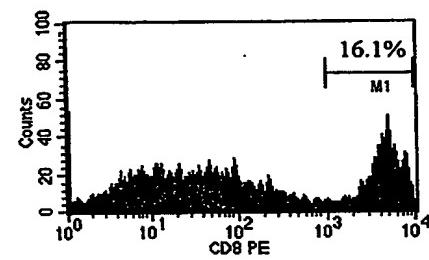


FIGURE 26

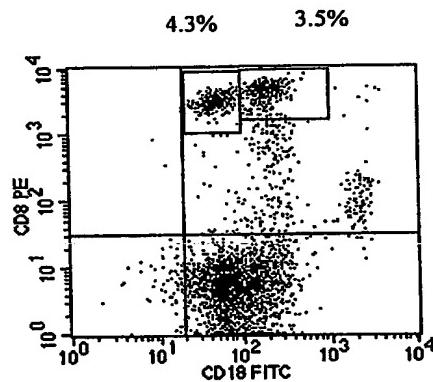
(a).



(b).



(c).



(d).

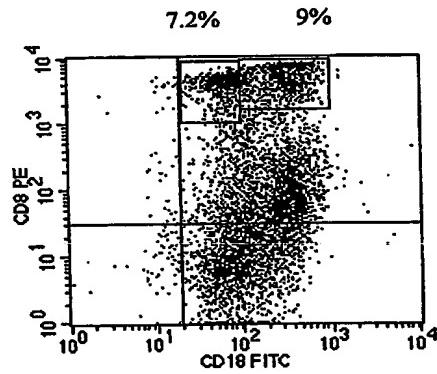


FIGURE 27

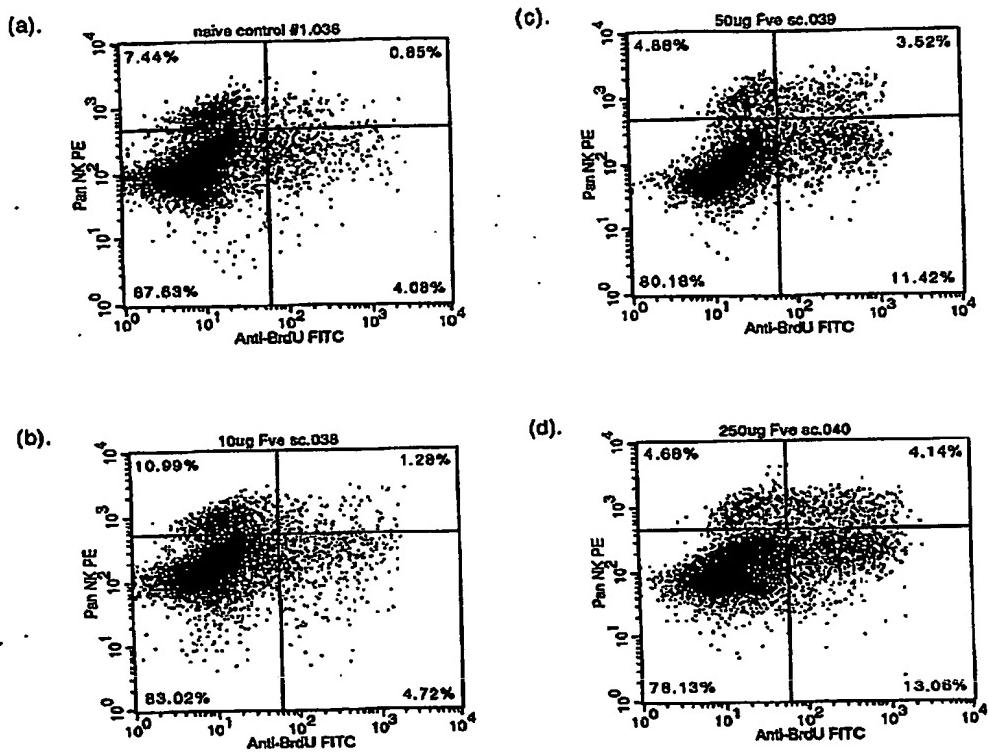


FIGURE 28

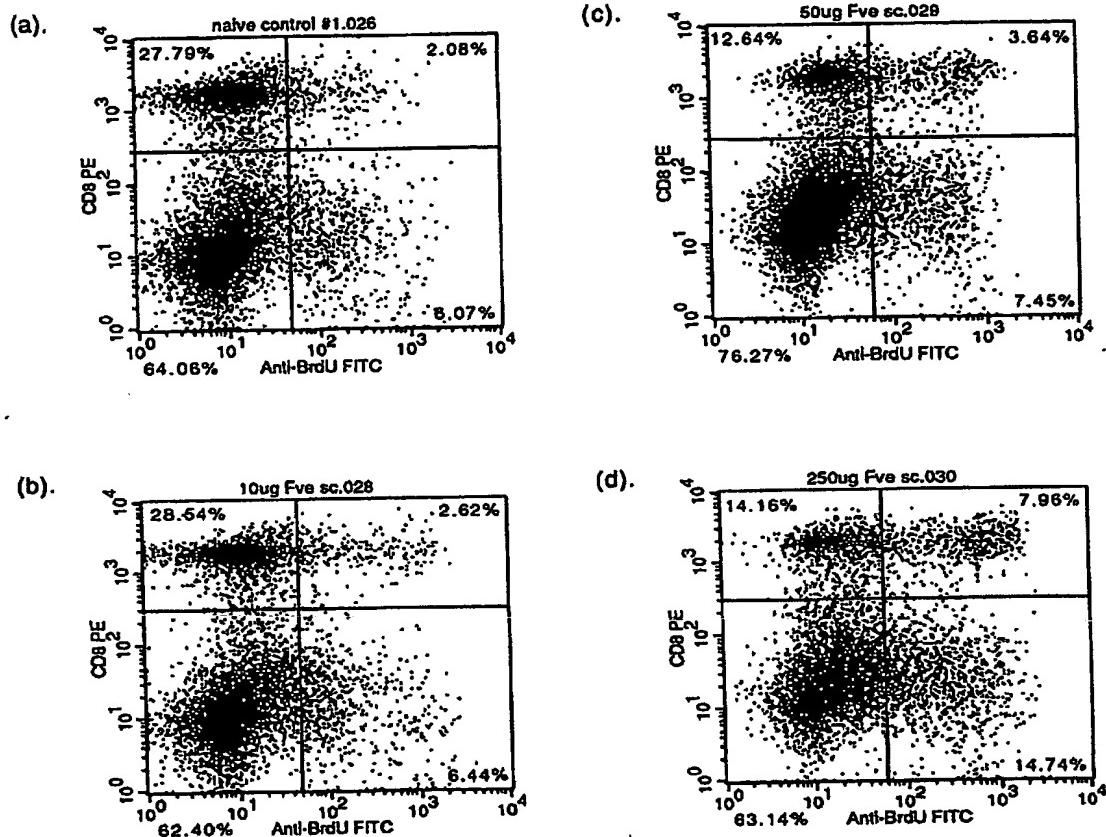


FIGURE 29

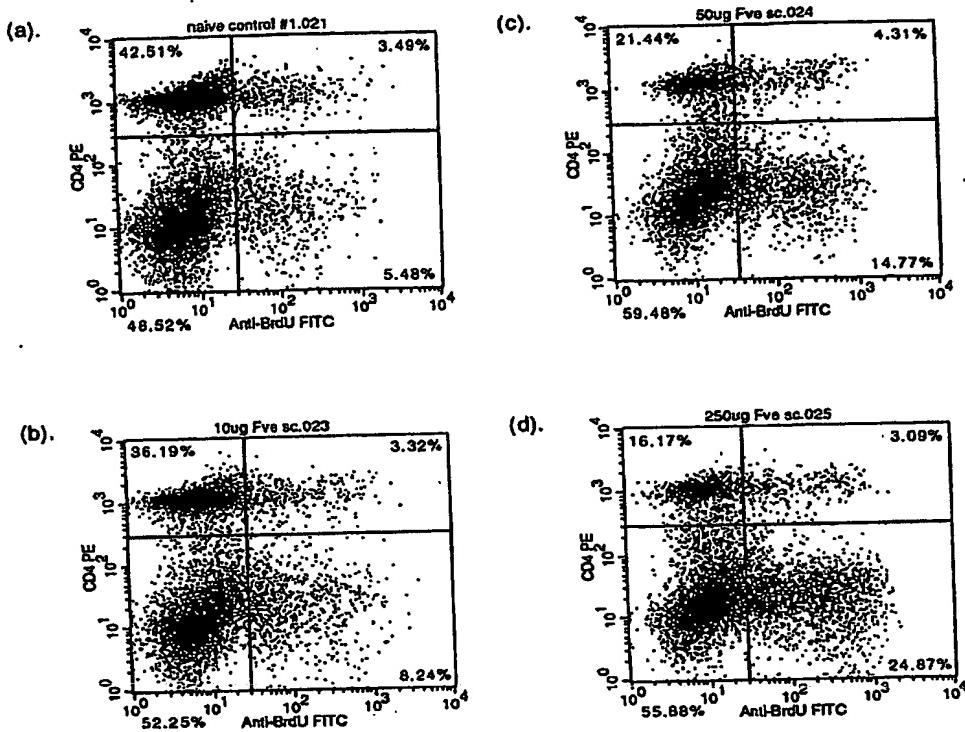


FIGURE 30

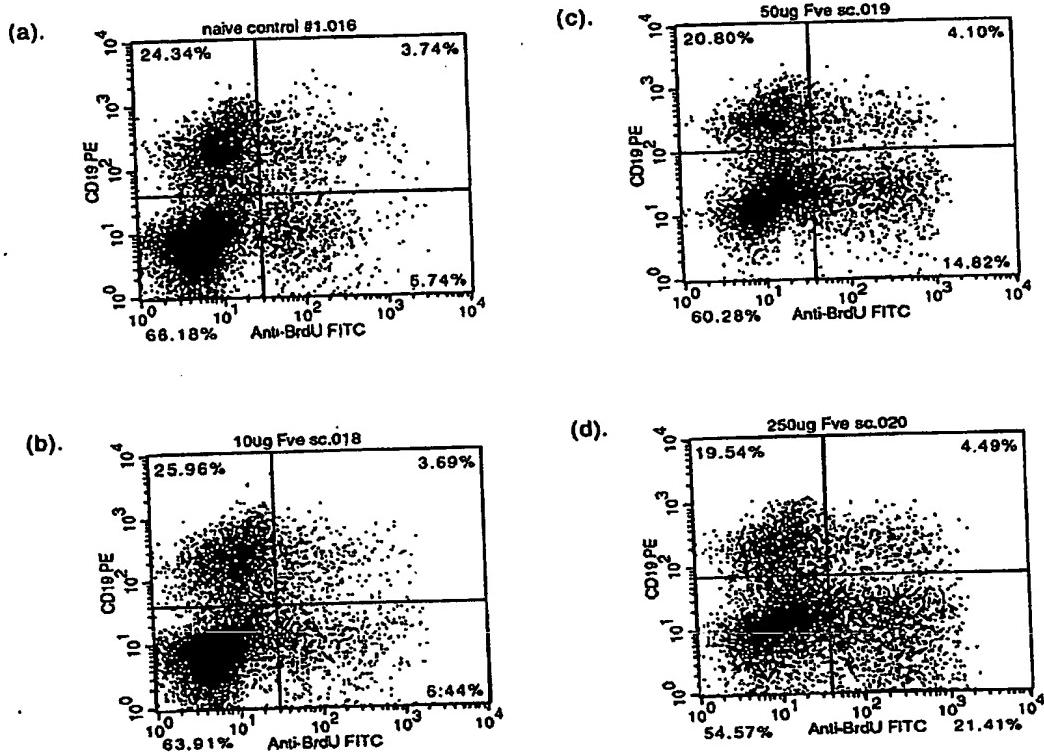


FIGURE 31

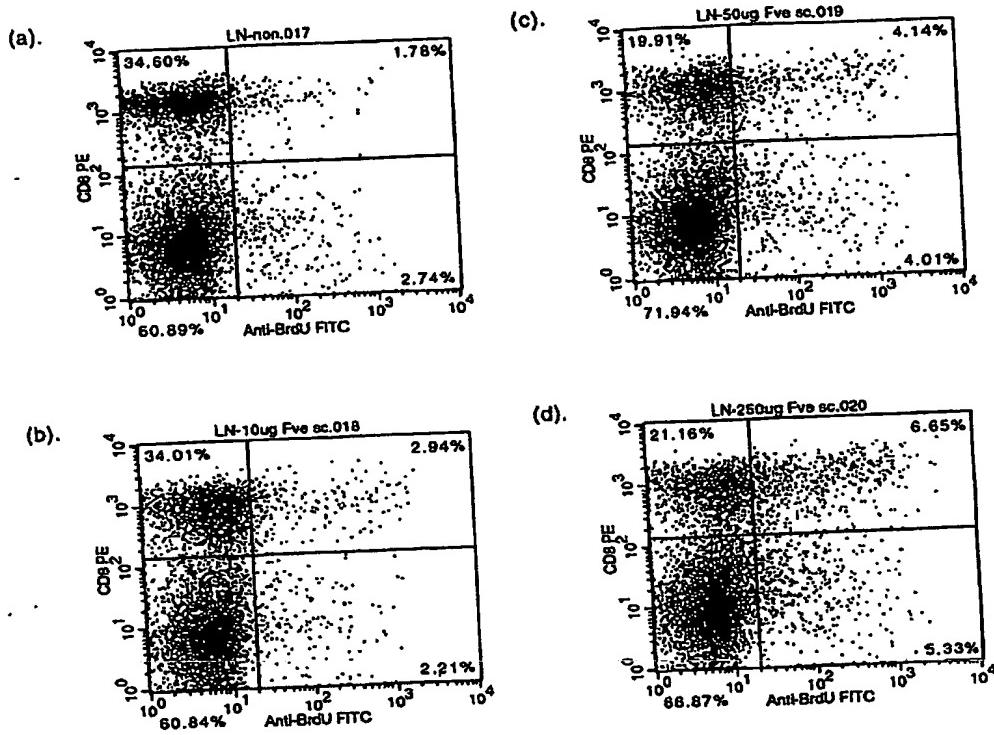


FIGURE 32

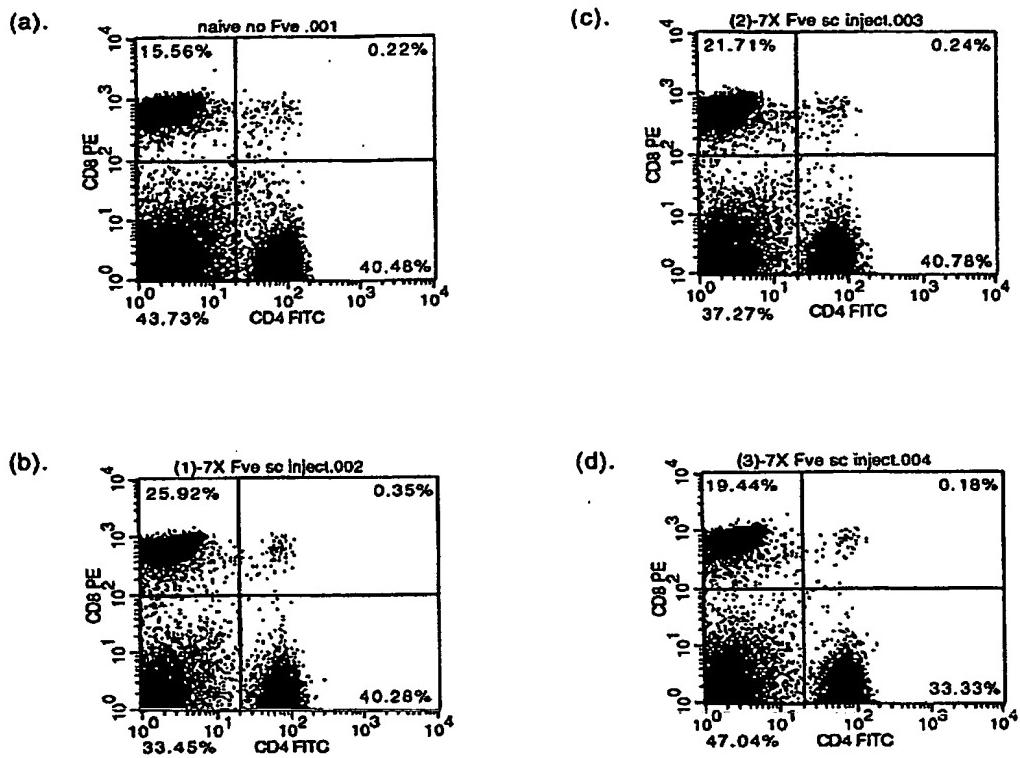
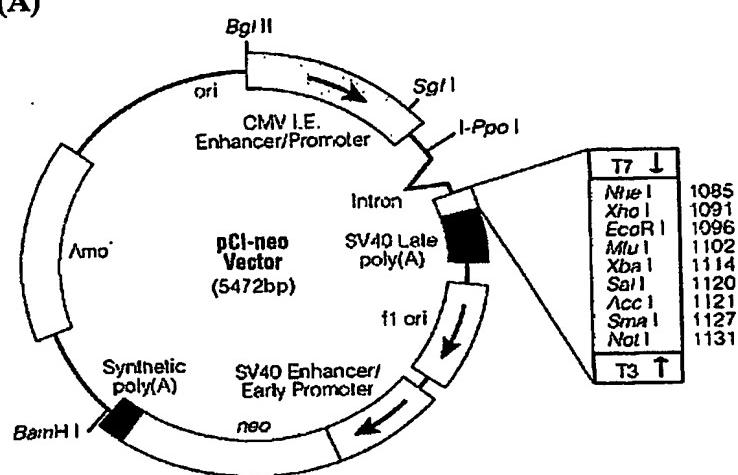


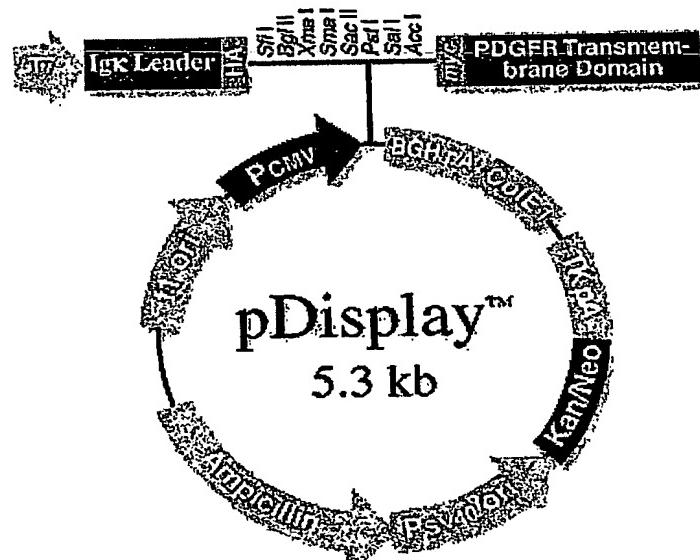
FIGURE 33

(A)



59-4WAWC 21

(B)

**FIGURE 34**

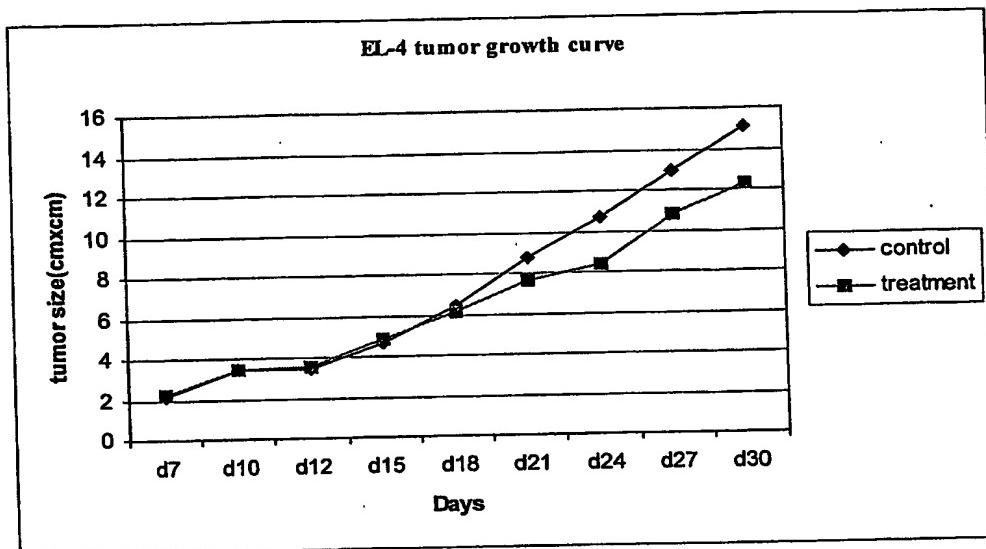
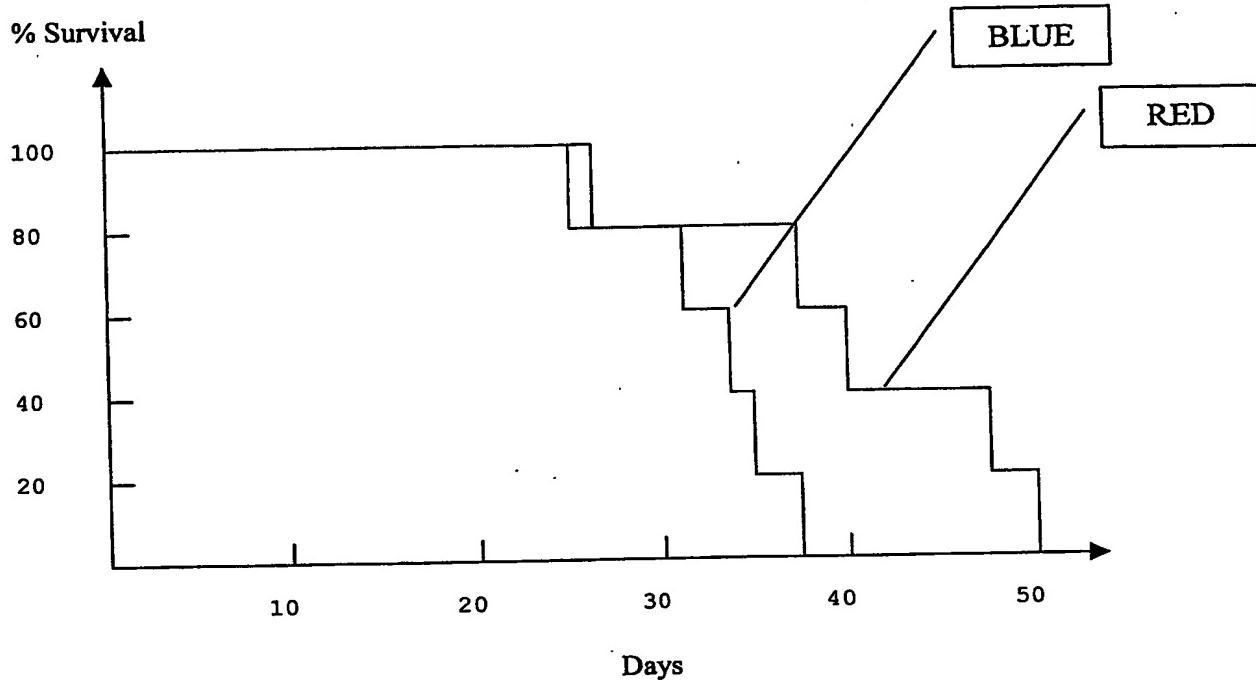


FIGURE 35

40/50

**FIGURE 36**

% survival

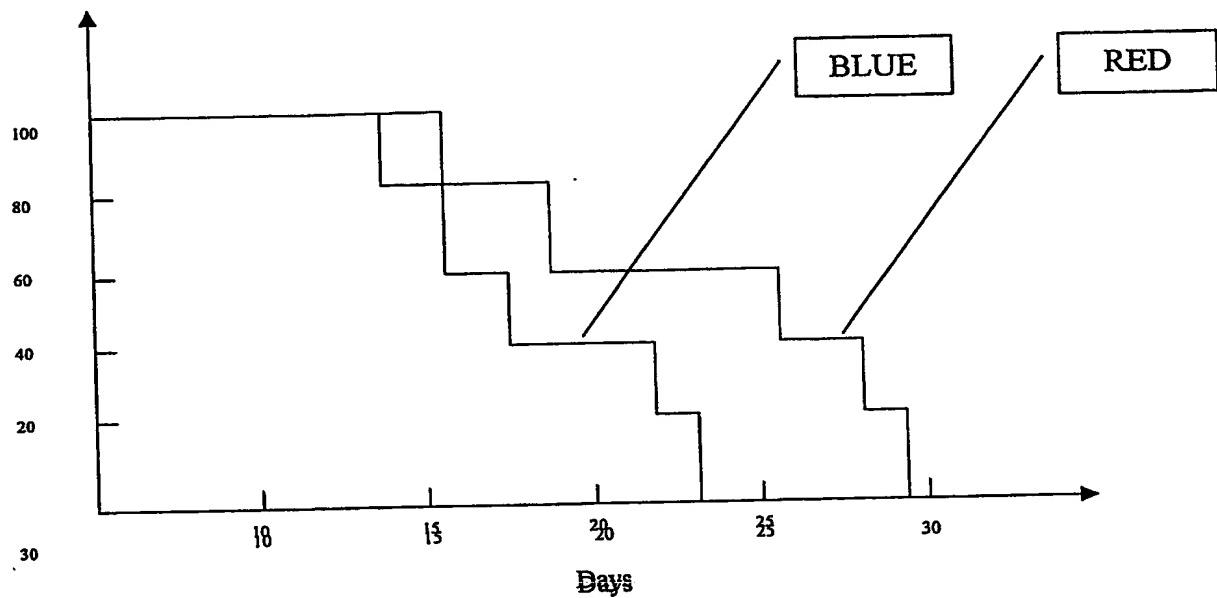


FIGURE 37

42/50

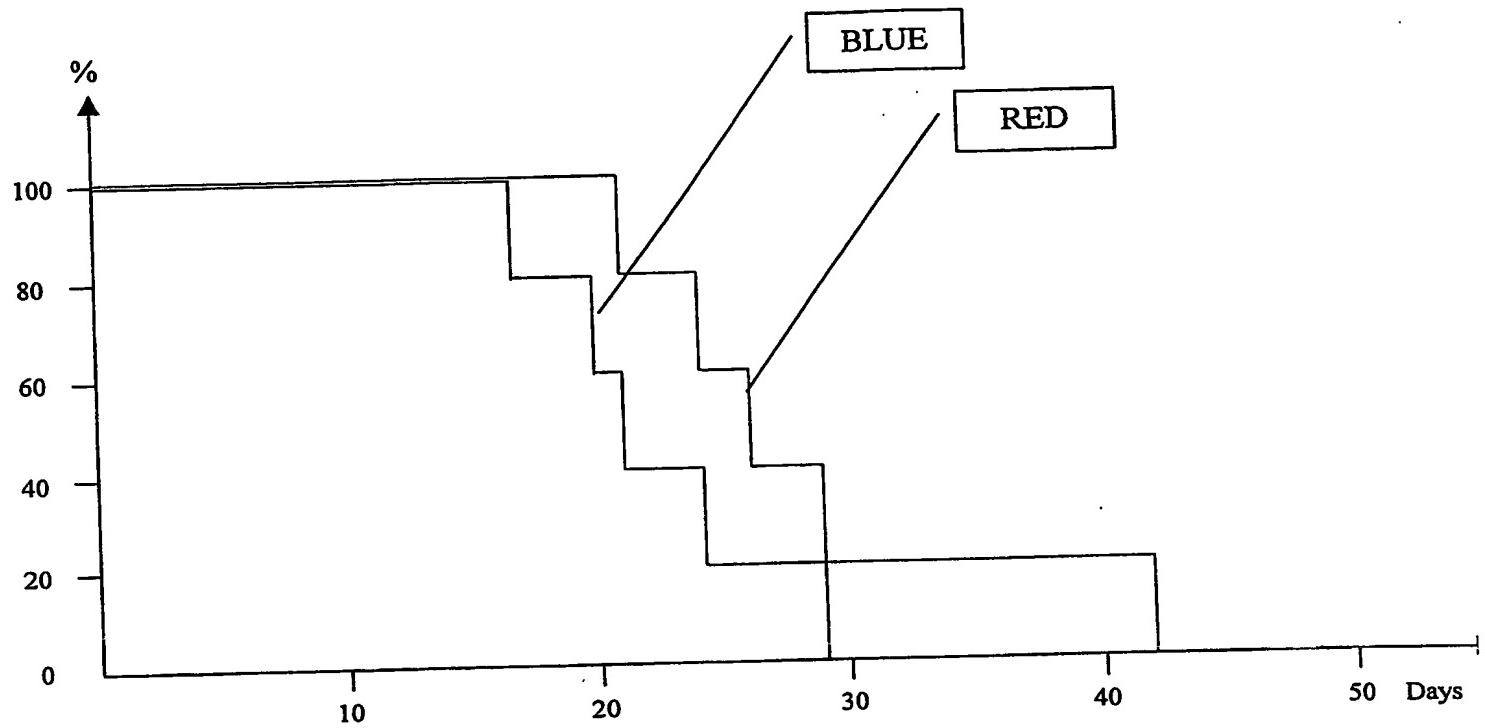


FIGURE 38

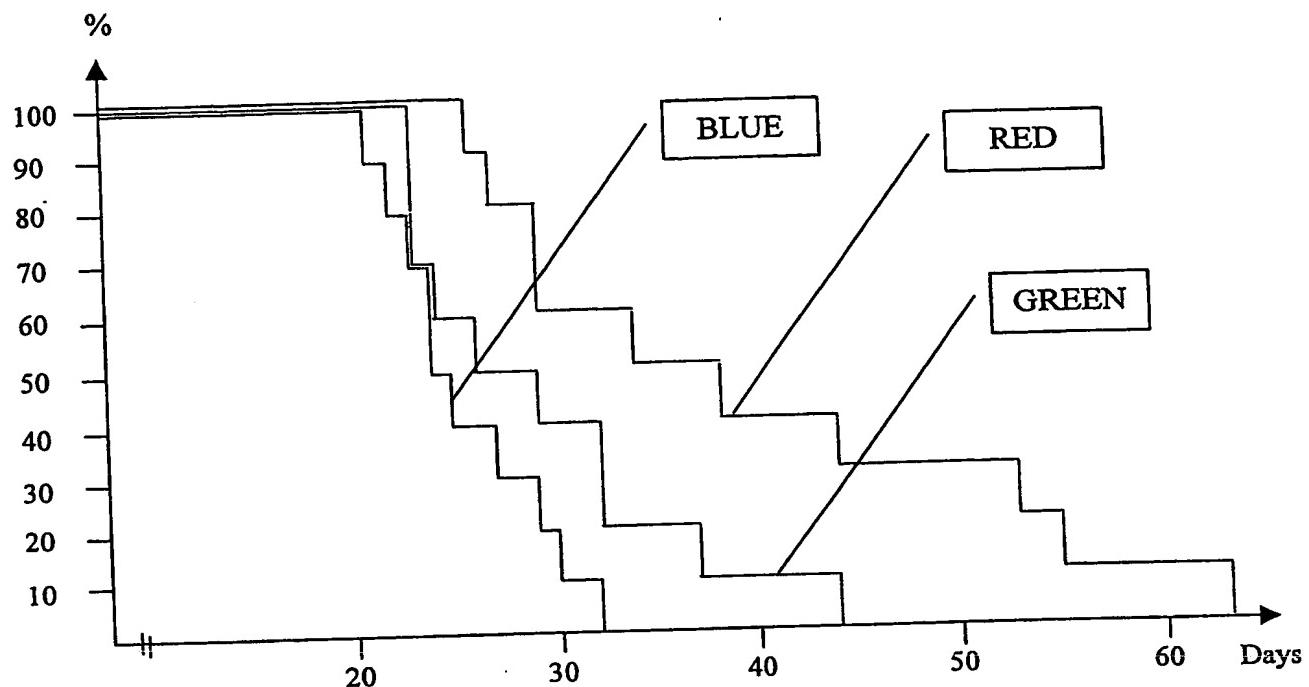


FIGURE 39

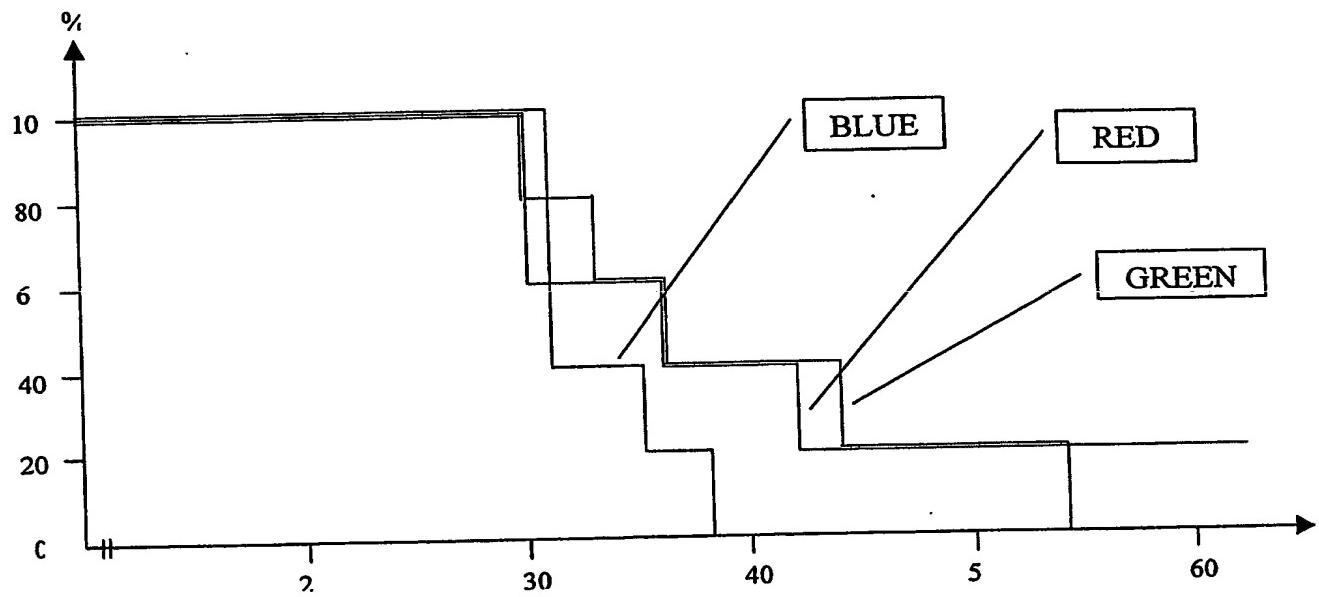
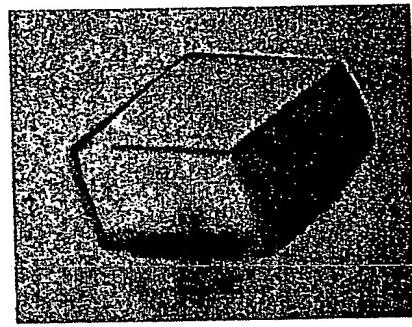
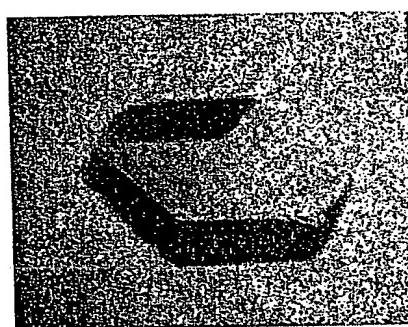


FIGURE 40

FIGURE 41



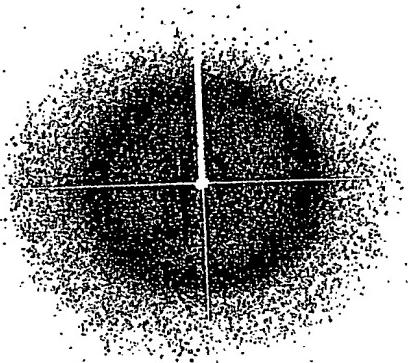


FIGURE 42

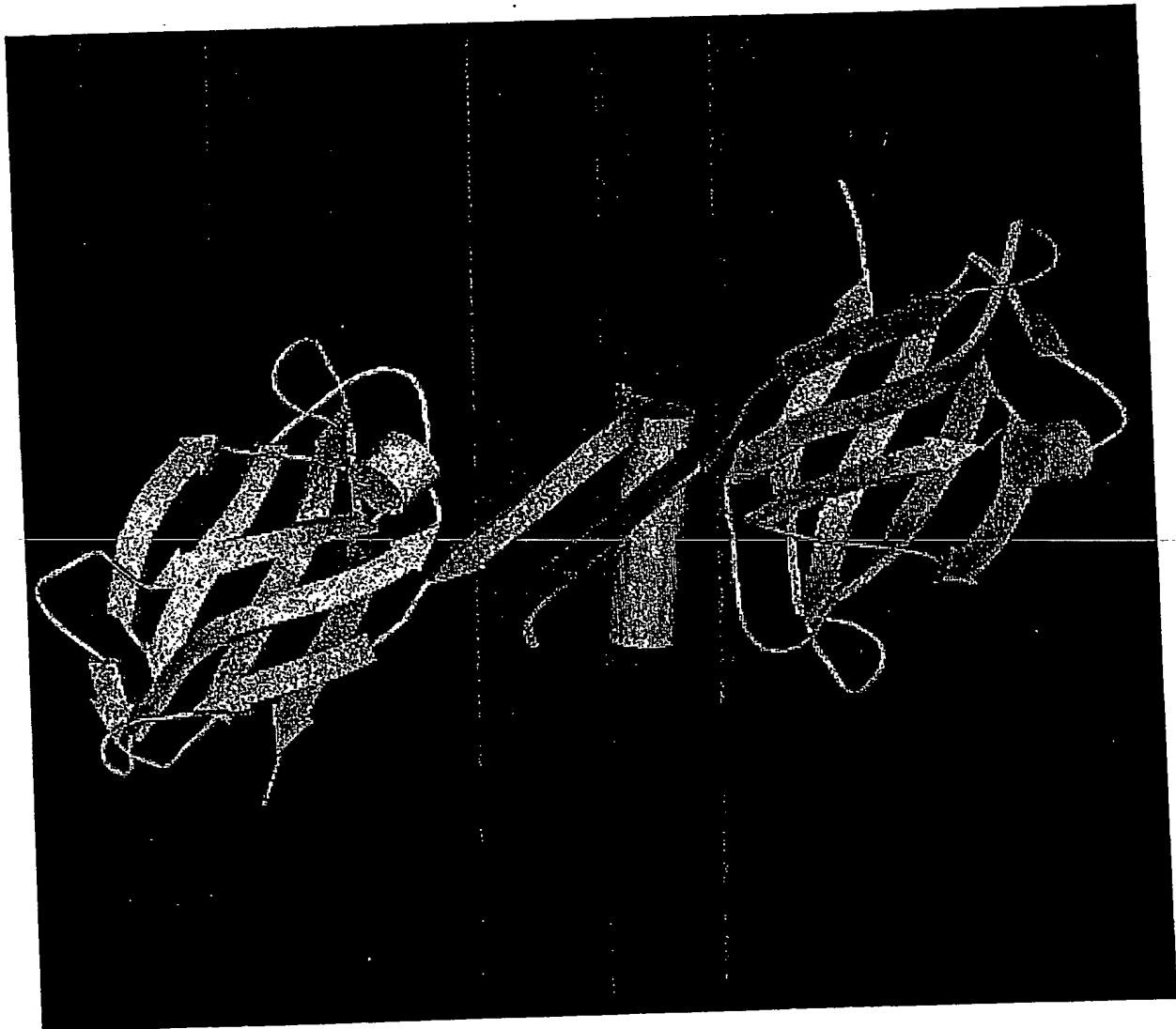
FIGURE 43

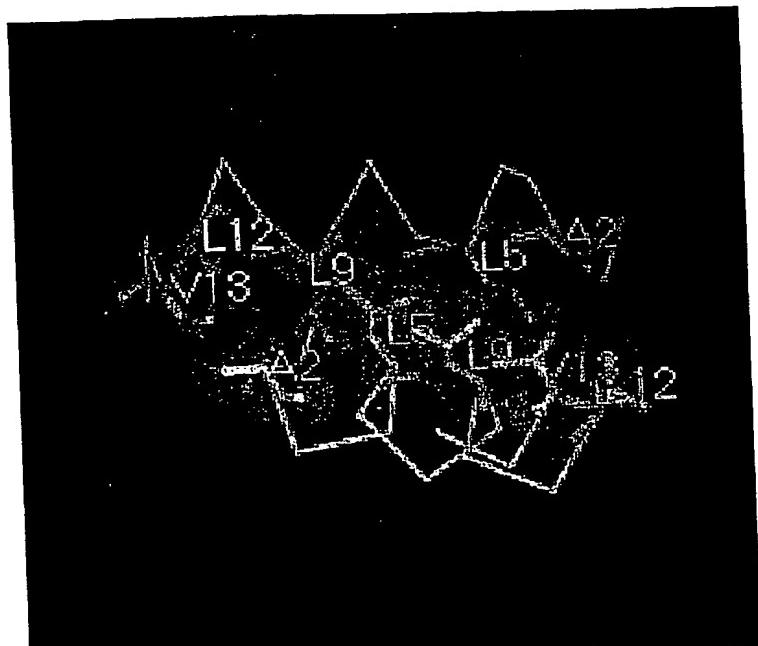
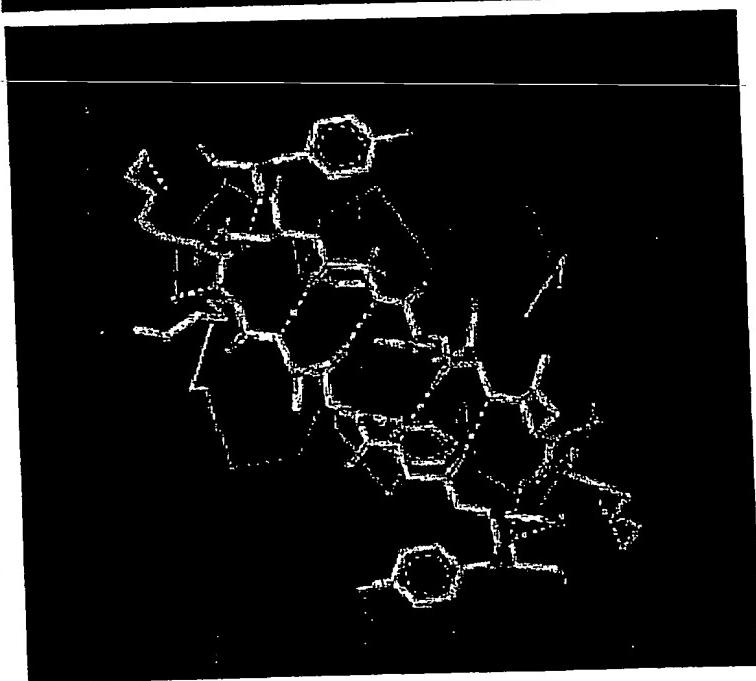
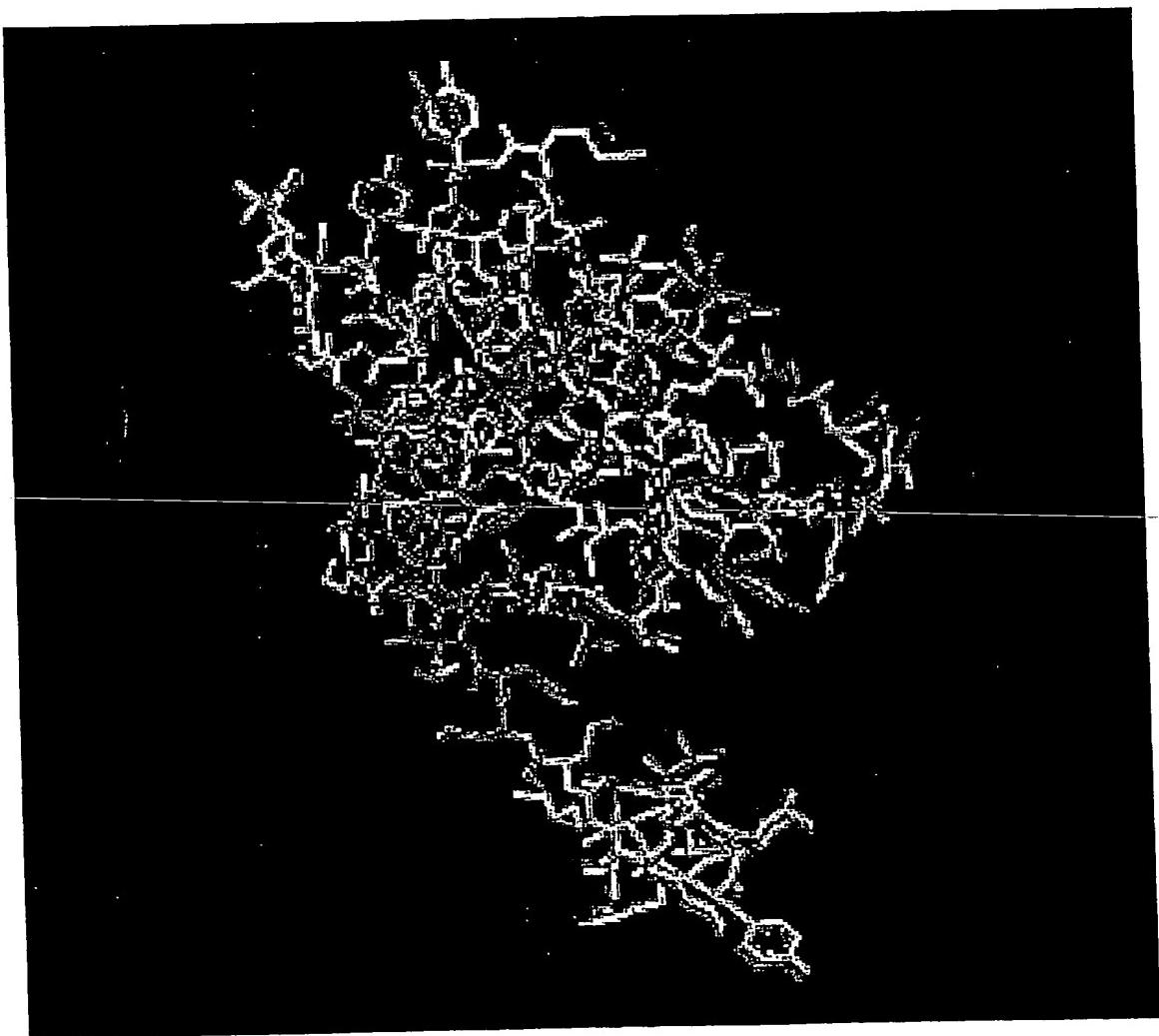
FIGURE 44A**FIGURE 44B**

FIGURE 44C



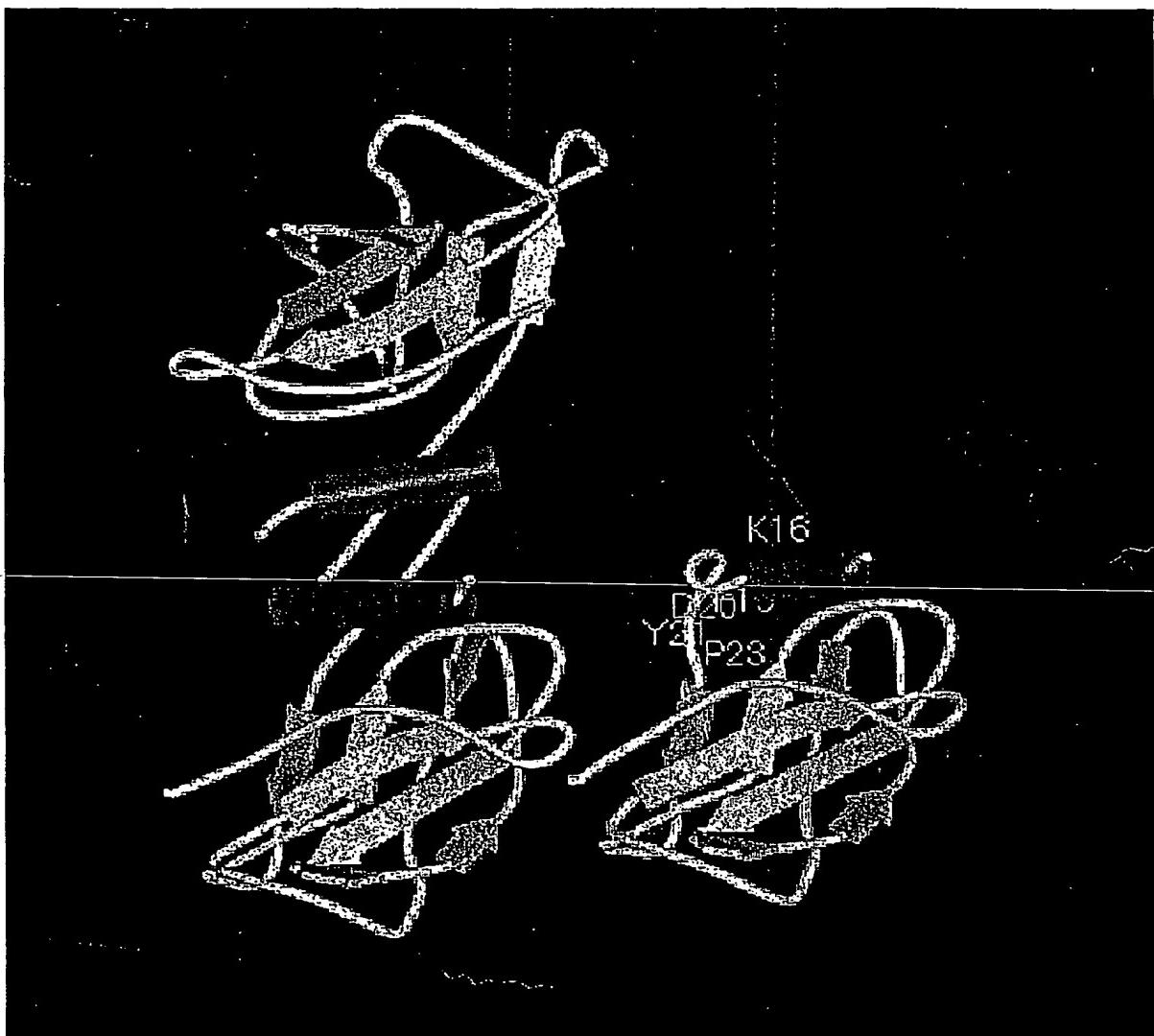


FIGURE 45A

FIGURE 45B

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